

## **SUPPLEMENTARY MATERIALS**

**To the manuscript entitled**

### **The Intron Retention Variant CsClpP3m Is Involved in Leaf Chlorosis in Some Tea Cultivars**

Xueyin Luo<sup>1†</sup>, Mengxian Zhang<sup>1†</sup>, Pei Xu<sup>1</sup>, Guofeng Liu<sup>1,2\*</sup> and Shu Wei<sup>1\*</sup>

<sup>1</sup> State Key Laboratory of Tea Plant Biology and Utilization, Anhui Agricultural University, Hefei, China, <sup>2</sup> Henan Provincial Key Laboratory of Tea Plant Biology, Xinyang Normal University, Xinyang, China

†, Both contributed to this work equally.

\*. For correspondence: Shu Wei, [weishu@ahau.edu.cn](mailto:weishu@ahau.edu.cn);  
Guofeng Liu, [liuguof0219@xynu.edu.cn](mailto:liuguof0219@xynu.edu.cn)

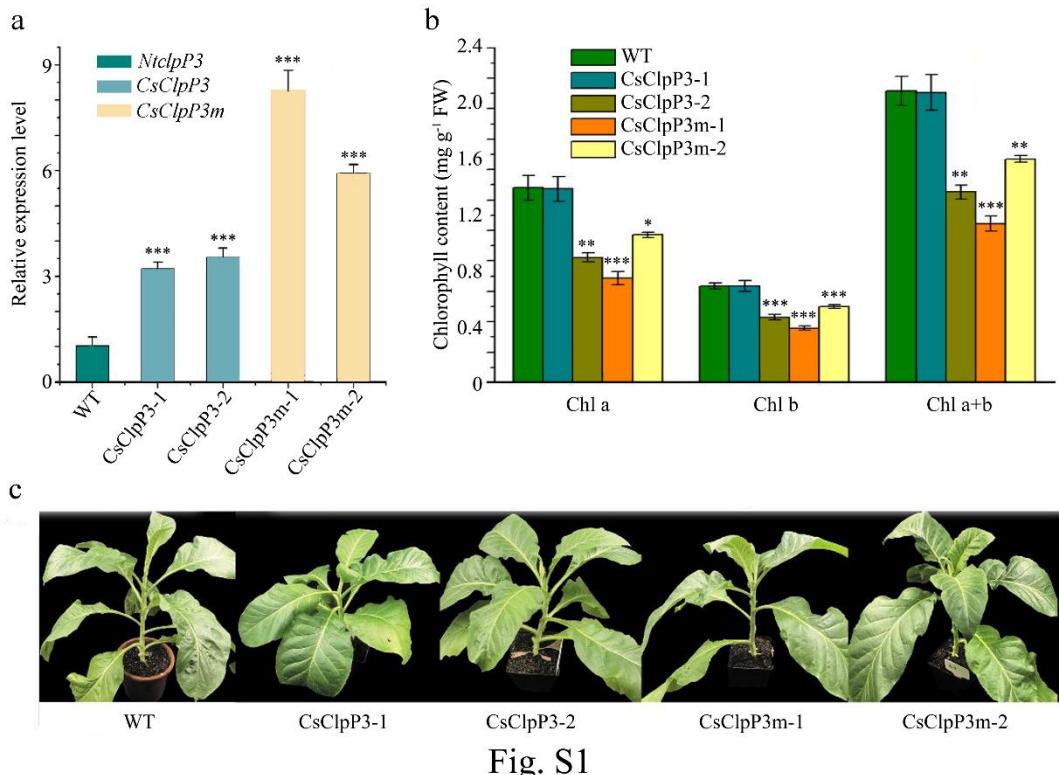


Fig. S1

**Fig. S1: Phenotypic analysis of transgenic tobacco over-expressing *CsClpP3* or *CsClpP3m*.**

**a** Enhanced transcript levels of *CsClpP3* or *CsClpP3m* in transgenic plants. WT, wild type plants; CsClpP3-1 and -2, transgenic lines expressing *CsClpP3*-YJX; CsClpP3m-1 and -2, transgenic lines expressing *CsClpP3m*-YJX. **b** Chlorophyll contents of transgenic plants over-expressing *CsClpP3* or *CsClpP3m*. **c** No visible difference in leaf color among wild type plant and transgenic tobaccos over-expressing *CsClpP3* or *CsClpP3m*. Statistical analysis was performed using Student *t*-test. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ .

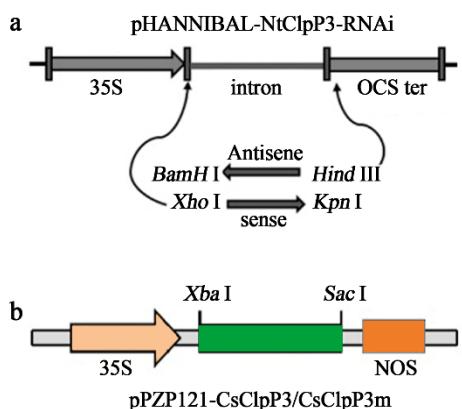


Fig. S2

**Fig. S2:** Construction of RNAi- NtClpP3 expression cassette using pHANNIBAL intermediate vector (**a**) and pPZP121-CsClpP3/CsClpP3m expression cassettes (**b**).

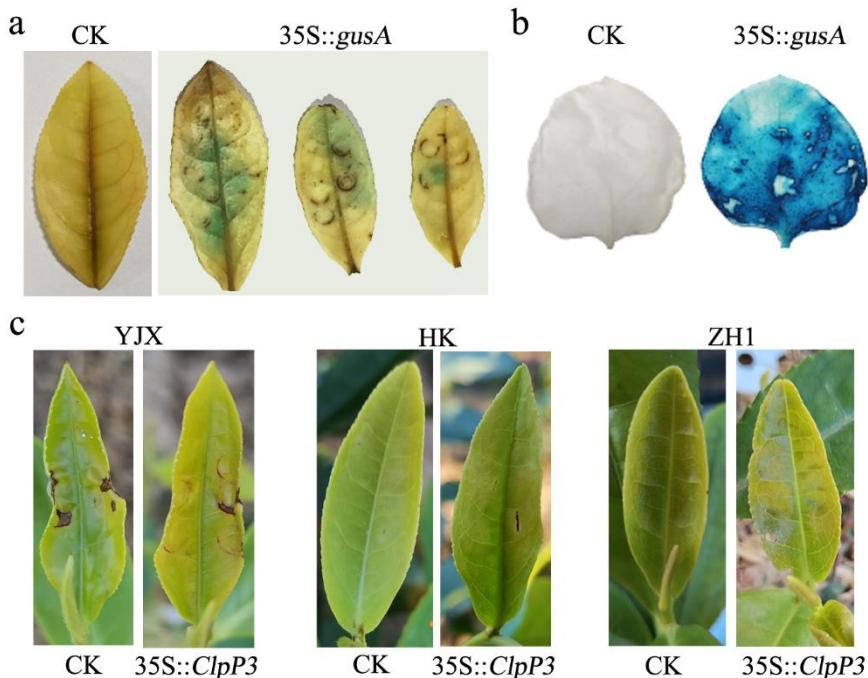


Fig. S3

**Fig. S3: Transient expression of pPZP121-CsClpP3 in chlorosis tea leaves.** **a** Transient expression of pPZP121 in chlorotic young tea leaves; **b** Transient expression of pPZP121 in tobacco leaves; **c** Leaf infiltration of Agrobacterium GV3101 containing pPZP121-CsClpP3 in YJX; **d** Leaf infiltration of GV3101 containing pPZP121-CsClpP3 in HK; **e** Leaf infiltration of GV3101 containing pPZP121-CsClpP3 in ZH1. CK, control using suspension buffer only for leaf infiltration; 35S::*gusA*, pPZP121; 35S::*ClpP3*, pPZP121-CsClpP3. Transient expression was done according to Sparks et al. (2006) with minor modifications. In brief, agrobacteria grown to  $OD_{600} = 0.8\text{-}1.0$  was used for resuspension with non-sucrose IM medium (AS,  $Na_3PO_4$ , MES); the final concentration of resuspended agrobacteria was  $OD_{600} = 1.0$ . Leaves were collected 4 d after infiltration. GUS staining was done according to the standard methods.

**Table S1: Primers used in the experiments.**

Primer name	Primer sequence and usage
<b>CsClp gene cloning</b>	
CsclpP3-F	tctagaGAGAGAGAGAGAGAGACAGAGATGGAG
CsclpP3-R	gagctcGTGATAAATCTTACATTGTGTTGAGG
CsclpP5-F	ggatccATGGCGCATTCTGCGTGT
CsclpP5-R	ctgeagTTACTGATCTGCACTCGCTGGTAGA
CsclpP6-F	ggatccATGGTAGCTTCAGCCATCTCAGCTC
CsclpP6-R	gtcgacTTAGTATTCTGTTCCAAGACCCCATCA
CsclpR2-F	ggatccATGGTTATCTCCCTTCATACAACCTGGT
CsclpR2-R	gtcgacCTAACCAAGACCTGTTCTGCATCT
CsclpR4-F	ggatccATGGAGGTGCCACCATGGC
CsclpR4-R	gtcgacTTAAATAAGTTGTGCTTTTCAGGTAG
CsclpC-F	ggtaccATGGCTGGGGCTTGGTCAG
CsclpC-R	ctcgagCTACACAGTGATTGGTTCTGGCAATGA
<b>Ntclp cloning</b>	
NtclpP3-sense-F	tttgagaggacacgctcgagCAAGGCTGATGTTCTACAATCTGC
NtclpP3-sense-R	ttcattcaattgggtaccTCTTCAGAAGGCAAATTTCTTTG
NtclpP3-antisense-F	gaaatcgataaggcttgatccTCAGAAGGCAAATTTCTTTGC
NtclpP3-antisense-R	tcattaaagcaggacttagaGGCTGATGTTCTACAATCTGCAT
<b>q-RT-PCR in <i>Camellia sinensis</i></b>	
c51485-L(ClP3)	AGCAACAGAGATGAGCATCGAATA
c51485-R(ClP3)	GTAAGGTGCAATTAATCCTGGCTTAC
c48468-L(ClP4)	CATCAACAGCTTCCATAATCCTTGG
c48468-R(ClP4)	TCTTGTGACATTGTCCTTGTATGC
c48000-L(ClP5)	CTTGCCAAACTCCAGGATAATGATC
c48000-R(ClP5)	TTGAGAGGATTCATGATAACACCGT
c33064-L(ClP6)	TCTCTAAGTTGCAGAAATGGTAGCT
c33064-R(ClP6)	TAGTAGCCTGCTCCTCAAGTTTAG
c53238-L(ClP2)	TCAGAATGAGAAGATGGAGACTGTC
c53238-R(ClP2)	ATATCTATGACAGCTCCACTTGACC
c15808-L(ClP2-1)	TCTAACTCATGTCTACACTCTGGGA
c15808-R(ClP2-1)	GTTTACTGTCACAAGCCTGGATTG
c42794-L(ClP3)	TGAGGGTTTGCAATTATGATGCT
c42794-R(ClP3)	ATTATTACCTCCTTGCACGGATGA
c38659-L(ClP4)	TGTTTACTTGGCATGTCAGT
c38659-R(ClP4)	CTCATAACCTAACCTTCGCCATCC
c27589-L(ClP1)	AATAAGATGCCGACACTGGAGGAG
c27589-R(ClP1)	ATTTGGGTACACGTTCTATTGTT
c43453c0-L(ClP1)	AGATCAAAGAAATCGAGCTACAGGT
c43453c0-R(ClP1)	TCGACAATACTGAATCACCCCTT
c51361-L-(ClpD)	AAGCATCATCATCATGTTCTTCACC
c51361-R-(ClpD)	TATGAAAGGAGTTGTTGGGTGAGAT
c62560-L-(ClpB1)	GGACCAGAACAAATTGCAAAAGAAG

c62560-R-(ClpB1)	CAACCCTGATCTAACATAGCTTC
c41573-L-(ClpB3)	TTCCAGTATTATTCCCTCACGCTCTT
c41573-R-(ClpB3)	TCATTGTAGCTGCAGAAGTGATG
<b>Quantitative RT-PCR for <i>Nicotiana tabacum</i><sup>a</sup></b>	
NtClpP3_F-qPCR	TCTCGGAACGGGATGTTTC
NtClpP3_R-qPCR	GATTCCCATCCCAGCAGTTA
NtClpP4_F-qPCR	CCTCACTGTTCCCTCCCTCAA
NtClpP4_R-Qpcr	GAGAAGGGGGTTTTGAAGC
NtClpP5_F-qPCR	TGTTGATCCCACAAAGGACA
NtClpP5_R-qPCR	CCAAGAGGCTGGTGAATCAT
NtClpP6_F-qPCR	GAGAACCCGTTAAAGCTCA
NtClpP6_R-qPCR	AGTTGCGAGGGTCACAAGTT
NtClpR1_F-qPCR	ATAACCCAGTACGGCGACAG
NtClpR1_R-qPCR	GGCATGCCAGATAGACAAT
NtClpR2_F-qPCR	GGGATTGCAGCGTCTAATGT
NtClpR2_R-qPCR	CACGTTACGGTAAAGAGCA
NtClpR3_F-qPCR	GTGCCAGCAGTCACAGAGTT
NtClpR3_R-qPCR	CAGCTGCAAGCAAGAGACAC
NtClpR4_F-qPCR	CTCTTCGCCCTTCTCCTCT
NtClpR4_R-qPCR	TGGCTTTCCCTCATCCTCAT
NtClpS_F-qPCR	TCCCCATCAAATCTTCCAAC
NtClpS_R-qPCR	GAATTCAGATTACGGCCAG
NtClpT1_F-qPCR	TCAACTTCGACTTCTTACAA
NtClpT1_R-qPCR	GCCGACCATTGGGCTGTAT
NtClpT2_F-qPCR	TAGGACCAATTCTCGTCGTT
NtClpT2_R-qPCR	ATTCACCCATGGCAAATGAT
NtClpC_F-qPCR	GGCGTCGACCTTACACTGTT
NtClpC_R-qPCR	TCACACTGCTCCGACATTC
NtClpD_F-qPCR	AATGCTGCTGTGCAACTGTC
NtClpD_R-qPCR	TGCTTGCCAAGATCACTTCA
<b>Cloning in <i>Arabidopsis</i> mutant</b>	
AtclpP3-F	TCCCAATCCGAAACCATAGAATCC
AtclpP3-R	GGTACAAACACAGGAAATGCAACGT
pROK2-LBb1	GCGTGGACCGCTTGCTGCA

<sup>a</sup> Primers used for tobacco were taken from Moreno et al. *J. Exp. Bot.* **68**, 2199–2218(2017).