# Supplementary Material

## Supplemental Table 1

Demographic and clinical data of influenza-induced ARDS patients

| Characteristic                                   | Flu-induced ARDS patients<br>(n=20) |  |
|--|-------------------------------------|--|
| Age (mean±SD, year)                              | 53.2±14.5                           |  |
| Male (n, %)                                      | 13/20                               |  |
| Flu virus strains                                |                                     |  |
| Influenza type A                                 | 16/20                               |  |
| Influenza type B                                 | 4/20                                |  |
| Comorbidities                                    |                                     |  |
| Pneumonia disease                                | 20/20                               |  |
| COPD   | 0/20                                |  |
| Asthma   | 0/20                                |  |
| Bronchiectasis                                   | 1/20                                |  |
| Diabetes   | 5/20                                |  |
| Kidney disease                                   | 10/20                               |  |
| Skin disease                                     | 0/20                                |  |
| Clinical features                                |                                     |  |
| Fever  | 20/20                               |  |
| Cough  | 20/20                               |  |
| Sputum   | 12/20                               |  |
| Hemoptysis                                       | 1/20                                |  |
| Consciousness Disorders                          | 1/20                                |  |
| Laboratory tests                                 |                                     |  |
| Total leukocytes (×10 <sup>9</sup> /L)           | 9.8±5.5                             |  |
| Neutrophil (×10 <sup>9</sup> /L)                 | 8.6±5.2                             |  |
| Lymphocytes (×10 <sup>9</sup> /L)                | $0.8 \pm 0.4$                       |  |
| Hemoglobin (g/L)                                 | 122±24.8                            |  |
| Platelet ( $\times 10^{9}/L$ )                   | 178.4±88.6                          |  |
| Creatine kinase (U/L)                            | 519.1±998.5                         |  |
| LDH (U/L)  | 611.1±313.9                         |  |
| PaO <sub>2</sub> /FiO <sub>2</sub> grade (mm Hg) |                                     |  |
| ≥200 to <300                                     | 4/20                                |  |
| ≥100 to <200                                     | 10/20                               |  |
| <100   | 6/20                                |  |
| ICU admission                                    | 20/20                               |  |
| NAI treatment                                    | 20/20                               |  |
| Use of invasive ventilation                      | 16/20                               |  |
| ЕСМО   | 10/20                               |  |

Note: ECMO = Extracorporeal membrane oxygenation

## Supplemental Table 2

| Primer   | Sequence (5'-3')             | Use For                   |
|----------|------------------------------|---------------------------|
| наса     | F: GACCAGACGCTCATAGCAG       | Q-PCR for detecting human |
| 1L-36A   | R: CTTTAGCACACATCAGGCAG      | IL-36A gene               |
| IL-36B   | F: ACAAAAAGCCTTTCTGTTCTATCAT | Q-PCR for detecting human |
|          | R: CCATGTTGGATTTACTTCTCAGACT | IL-36B gene               |
| IL-36G   | F: GGTCAGAACCTTGTGGCAGT      | Q-PCR for detecting human |
|          | R: CAAGAGCCTCTGGATACTTGC     | IL-36G gene               |
| IL-18    | F: AAGACTCTTGCGTCAACTTCAAGGA | Q-PCR for detecting human |
|          | R: AGTCGGCCAAAGTTGTCTGATTC   | IL-18 gene                |
| G-CSF    | F: TCCCCATCCCATGTATTTATCT    | Q-PCR for detecting human |
|          | R: AACTCAGAAATGCAGGGAAGGA    | G-CSF gene                |
| CCL20    | F: TCTGTGTGCGCAAATCCAA       | Q-PCR for detecting human |
|          | R: CCATTCCAGAAAAGCCACAG      | CCL20 gene                |
| CXCL3    | F: CGCCCAAACCGAAGTCAT        | Q-PCR for detecting human |
|          | R: GTGCTCCCCTTGTTCAGTATC     | CXCL3 gene                |
| н        | F: TAATGGGCATTCCTTCTTCT      | Q-PCR for detecting human |
| 1L-6     | R: TGTCCTAACGCTCATACTTTT     | IL-6 gene                 |
| CXCL8    | F: AGGGTTGCCAGATGCAATAC      | Q-PCR for detecting human |
|          | R: GCAAACCCATTCAATTCCTG      | CXCL8 gene                |
| IL-12    | F: TGGAGTGCCAGGAGGACAGT      | Q-PCR for detecting human |
|          | R: GTTGATGTCCCTGATGAAGAAGC   | IL-12 gene                |
| IENC     | F: GCAGAGCCAAATTGTCTCCT      | Q-PCR for detecting human |
| IFNG     | R: ATGCTCTTCGACCTCGAAAC      | IFNG gene                 |
| н 26р    | F: GCTGGAGTGTCCACAGCATA      | Q-PCR for detecting human |
| IL-36R   | R: GCGATAAGCCCTCCTATCAA      | IL-36R gene               |
|          | F: AAGACAGCTGTTTCAATTCC      | Q-PCR for detecting human |
| IL-IRACP | R: TTGAAATTAAGGCAATGAGG      | <i>IL-1Acp</i> gene       |
| COX-2    | F: GTCCTGGCGCTCAGCCATACAG    | Q-PCR for detecting human |
|          | R: TCCTGTCCGGGTACAATCGCAC    | COX-2 gene                |
| 0.1.7    | F: AGAAGGCAGCTCACGAAACC      | Q-PCR for detecting human |
| UAS      | R: CCACCACCCAAGTTTCCTGTA     | OSA gene                  |
| PKR      | F: AGAGTAACCGTTGGTGACATAACCT | Q-PCR for detecting human |
|          | R: GCAGCCTCTGCAGCTCTATGTT    | PKR gene                  |
| Mx1      | F: GAGAGGAAACTGTAGGGGA       | Q-PCR for detecting human |
|          | R: GGAAACATCTGTGAAAGCAA      | Mx1 gene                  |
| IAV-NP   | F: TCAGTGATTATGATGGACGACTA   | Q-PCR for detecting       |
|          | R: GCACTGGGATGCTCTTCTAGGTA   | IAV-NP                    |
| NP-mRNA  | Oligo DT                     | For reverse transcription |
| NP-vRNA  | AGCAAAAGCAGG                 | For reverse transcription |

The primer used in the RT-PCR



#### **Supplemental Figure 1.**

(A) The relative mRNA expression levels of *IL-36R* and *IL-1RAcp* in PBMCs of Influenza A virus (IAV)-induced ARDS patients (n = 12) and healthy individuals (n = 12) were detected by RT-PCR. (**B**, **C**) The gating strategy and representative flow plots. IL-36R expression on peripheral blood mononuclear cells (PBMC) isolated from whole bloods were detected. (**D**) Representative flow cytometry dot plots of CD19<sup>+</sup>IL-36R<sup>+</sup> lymphocytes gated from CD19<sup>+</sup> B cells in healthy control and IAV-induced ARDS patient. (**E**) IL-36R<sup>+</sup> B cells in total B cells and expression levels of IL-36R on B cells from IAV-induced ARDS patients (n = 13) and healthy individuals (n = 12) were detected using flow cytometry. Results are presented as scatter plots with median of the proportion or mean fluorescence intensity (MFI) subtracting corresponding isotypic controls. Statistical significance is determined by Student's t test. \*P < 0.05, \*\*\*P < 0.001, *ns*, not significant.



### Supplemental Figure 2.

(A) 16HBE cells were infected for the indicated timeframes with IAV (MOI = 1), after this, treated with or without ATP (1 mM, 2 h), intracellular protein level of IL-36 $\gamma$  was examined and analyzed by Western Blotting and Image J. (B) 16HBE cells were treated with IL-36 $\gamma$  (100 ng/ml), or left untreated 24 h prior to infection with IAV-Ca07 (MOI = 1). Levels of IFN- $\lambda$ 1, IFN- $\alpha$  and IFN- $\beta$  in supernatant medium were measured by ELISA in indicated timeframes. (C) Apoptotic 16HBE cells were determined by flow cytometry after infected with IAV-Ca07 (MOI = 1) or joined IL-36 $\gamma$  (100 ng/ml) together for 24 h. The early apoptosis ratio and total apoptosis ratio (early apoptosis and late apoptosis) was quantified by Flow J software. The data shown were the mean  $\pm$  SD of three independent experiments. (D) 16HBE cells were infected with or without IAV-Ca07 and treatment with or without IL-36 $\gamma$  for 12 h. Cells were then collected for TEM analysis of the autophagic structures. Representative autophagosomes or autolysosomes (black arrows) are highlighted in the enlarged images (right). Statistical significance is determined by Student's t test. \*P < 0.05, \*\*P < 0.01, *ns*, not significant.



## **Supplemental Figure 3.**

(A) Flow cytometry to reveal IL-36R expression on the surface of Jurkat T cells after treated with IL-36 $\gamma$  (100 ng/ml) or infected with IAV (MOI = 1) for 24 h. (B) Relative mRNA expression levels of *IL-36R* and *IL-1RAcP* in A549,16HBE as determined by RT-PCR, and Jurkat T cells served as a control. (C) mRNA levels of *IL-36R* and *IL-1RAcp* in human BALF and in PBMCs from influenza-induced ARDS patients (n = 3) were detected using RT-PCR. Statistical significance is determined by Student's t test. \*\*P < 0.01, *ns*, not significant.



### **Supplemental Figure 4.**

(A) A549 cells were pre-treated with recombinant IL-36 $\gamma$  (100 ng/ml) for 12 h before IAV-Ca07 infection (MOI = 1). Cells were harvested for the indicated timeframes. Total RNA was isolated for detection of NP-specific mRNA and vRNA, and total protein was extracted for western blotting to detection of IAV-NP and M1. (B) A549 cells were pre-treated with IL-36 $\gamma$  (100 ng/ml) and (or) IL-36Ra (100 ng/ml) prior to IAV-Ca07 challenge. After 24 h, ELISA was performed to measure the IFN- $\alpha$ , IFN- $\beta$  and IFN- $\lambda$ 1 concentrations in the culture supernatant. Statistical significance is determined by Student's t test. \*P < 0.05, \*\*P < 0.01.