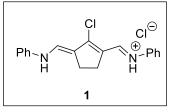


Supplementary Material

1 Experimental Procedures and NMR Data

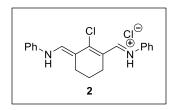
General Materials and Methods: Unless otherwise stated, all reactions were conducted in ovendried glassware under an atmosphere of argon using anhydrous solvents. All commercially obtained reagents were used as received. Flash chromatography was performed using Biotage SNAP Ultra C18 columns (12g and 30g) on the Biotage Isolera One instrument. LC-MS analyses were conducted on an Agilent 1260 Infinity ESI-MS using the following method: 30-100% MeCN (90% MeCN, 10% H₂O) in H₂O with 0.1% formic acid, 0.300 mL/min, 14 min. All NIR dye compounds were observed in the mass spectral data as the parent molecule without the Na⁺ counterion. NMR spectra was recorded on Varian 400, 500 or 600 MHz NMR spectrometers using the stated deuterated solvents.

1.1 Synthesis of NIR Cyanine Dyes



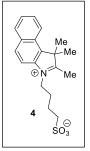
1: To a solution of N-methylformanilide (6.16 mL, 49.94 mmol) in chloroform (6.5 mL) was added phosphorous oxychloride (7.0 mL, 74.87 mmol) slowly at 0° C. After complete addition, the reaction mixture was stirred at room temperature for 1 hr. Cyclopentanone (1.77 mL, 19.97 mmol) was then added and the reaction was heated at 50° C for 4 hr. The mixture was cooled to room temperature, and the following reagents were

added: potassium carbonate (5.0 g), then aniline (4.12 mL, 45.10 mmol), 6N HCl (3.75 mL), H₂O (25 mL). After addition, the mixture was stirred at room temperature for 1 hr. DCM (25 mL) was added to the reaction mixture and let stand for a while. Finally, the solid was filtered, washed with acetone and H₂O, and dried overnight in a vacuum oven to afford the light purple solid (6.44 g, 93%). ¹H **NMR** ((CD₃)₂SO, 600 MHz): δ 8.31 (s, 1H), 7.62 (d, *J* = 8.0 Hz, 2H), 7.57 – 7.40 (m, 4H), 7.32 – 7.18 (m, 4H), 6.91 (s, 1H), 2.75 (s, 4H). **MW** (without Cl⁻ anion): 309.82 g/mol.



2: N,N'-Dimethylformamide (3.7mL, 48mmol) was placed in an oven dried 100mL round bottom flask, placed under Argon, and cooled to 0°C. The DMF was charged dropwise with phosphorus oxychloride (3.22mL, 34.5mmol) and stirred for 30 minutes at 0°C. Cyclohexanone (1.56mL, 15mmol) was added and the reaction mixture was refluxed for 1 hour. After cooling to room temperature, the flask was charged dropwise with

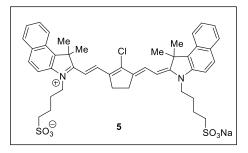
50mL of a 1:1 mixture of aniline and ethanol and stirred at room temperature for 30 minutes. The red solution was then poured into a flask with cold 2M HCl(aq) and was refrigerated for several hours. A purple solid was collected by vacuum filtration and washed with ice cold water. The solid was dried for several hours in a vacuum oven to yield a dark purple powdery solid (4.322g, 89%). ¹H



NMR (400 MHz, DMSO-d₆) δ 8.54 (s, 1H), 7.64 – 7.58 (m, 4H), 7.49 – 7.44 (m, 4H), 7.35 – 7.25 (m, 3H), 2.75 (t, *J* = 6.2 Hz, 4H), 1.86 (p, *J* = 6.1 Hz, 2H). **MW** (without Cl⁻ anion): 323.84 g/mol.

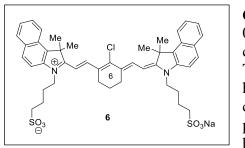
4: 1,1,2-trimethyl-1H-benzo[e]indole (1.01 g, 4.83 mmol) and 1,4-butane sultone (1.48 mL, 14.48 mmol) were added neat to a round-bottom flask at normal atmosphere, and stirred for 2 hr at 120° C. After 2 hr, the solid reaction mixture was washed thoroughly with acetone, filtered, and dried in a vacuum oven to afford the

grey-blue solid product (1.65 g, 99%). ¹**H NMR** ((CD₃)₂SO, 600 MHz): δ 8.36 (d, *J* =8.3 Hz, 1H), 8.28 (d, *J* =8.9 Hz, 1H), 8.23-8.19 (m, 2H), 7.80-7.76 (m, 1H), 7.74-7.70 (m, 1H), 4.61 (t, *J* = 7.9 Hz, 2H), 2.95 (s, 3H), 2.54-2.52 (m, 2H), 2.08-2.01 (m, 2H), 1.81-1.76 (m, 2H), 1.76 (s, 6H). **MW**: 345.46 g/mol.



5: Indolenium salt **4** (1.50 g, 4.34 mmol) and dianiline **1** (0.5 g, 1.45 mmol) were dissolved in isopropyl alcohol (7 mL) in a round-bottom flask. To the reaction mixture was added triethylamine (1.21 mL) then acetic anhydride (0.41 mL). The reaction was stirred at 50 °C for 2 hr, while monitored by LC-MS. Upon completion, the reaction was cooled to room temperature, added to diethyl ether, and cooled at 2° C to afford crystallization of the product. Product was purified by

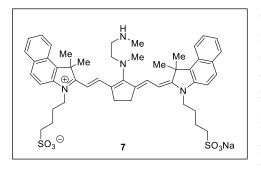
recrystallization from MeOH/Et₂O, followed by reverse-phase chromatography (30-100% ACN in H₂O) to afford **5** as a green solid (0.54 g, 45%). **LC-MS analysis**: $\lambda_{max} = 844$ nm, MS: [M+H] 813; [M-H] 811 (MW of **5**: 835.45 g/mol). ¹H NMR ((CD₃)₂SO, 600 MHz): δ 8.27 (d, *J* = 8.6 Hz, 2H), 8.09-8.05 (m, 4H), 7.86 (d, *J* = 12.9 Hz, 2H), 7.80 (d, *J* = 8.8 Hz, 2H), 7.67-7.62 (m, 2H), 7.54-7.47 (m, 2H), 6.25 (d, *J* = 14.2 Hz, 2H), 4.33 (t, *J* = 7.6 Hz, 4H), 3.15-3.05 (m, 4H), 3.01 (s, 4H), 1.94 (s, 12H), 1.89 (p, *J* = 7.8, 7.4 Hz, 4H), 1.82-1.74 (m, 4H).



6: Indolenium salt **4** (1.108g, 3.206mmol), dianiline **2** (0.259g 0.802mmol), and sodium acetate (0.1604g, 1.96mmol) were combined in an oven-dried 50mL flask and placed under Argon. The reagents were dissolved in 200 proof ethanol (10mL) and heated to reflux for 5 hours. The now blue-green solution was cooled to room temperature and concentrated under reduced pressure. The residue was dry loaded onto celite and purified by reverse phase column chromatography on a C18 column with

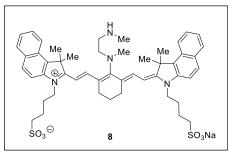
the product eluting in 60% acetonitrile in water. The relevant fractions were combined and concentrated under reduced pressure and transferred to an 8-dram vial with a minimal amount of DMF. The vial was filled with ethyl ether (25mL) and cooled in a refrigerator overnight. The vial was rewarmed to room temperature, and the top ether layer was removed with a pipette while leaving the crystallized solid at the bottom of the container. More fresh ether was added to the container with the green solid, and the solid was allowed to settle to the bottom. The ether layer was removed again once the solid settled to the bottom of the container. Subsequent washing with ether and removal of the ether was performed until the ether layer is clear. The solid was dried under a constant stream of dry nitrogen to afford IR-820 as a dark green powdery solid (0.396g, 60%). Drying the solid was only performed directly before usage of the compound. The product was stored in an 8-dram vial filled with ether to separate the solid at the bottom from the moisture and oxygen in the air. LC-MS analysis: λ_{max} (820 nm), observed [M+(-H)] = 827 [M-(-H)] = 825. ¹H NMR (500 MHz, DMSO- d_6) δ 8.41 – 8.34 (m, 2H), 8.30 (d, J = 8.7 Hz, 2H), 8.12 – 8.03 (m, 4H), 7.86 (d, J = 9.1 Hz, 2H), 7.68 – 7.62 (m, 2H), 7.52 (t, J = 8.4, 6.9 Hz, 2H), 6.56 (d, J = 14.2 Hz, 2H), 4.51 (t, J = 7.9 Hz, 4H), 2.79 (t, J = 6.3 Hz, 4H), 2.61 (t, J = 6.7 Hz, 4H), 2.44 – 2.37 (m, 2H), 2.13 – 2.03 (m, 4H), 1.96 (s, 12H), 1.91 – 1.72 (m, 4H).

1.2 Coupling of Linker to NIR Cyanine Dyes



7: To a solution of 5 (0.121 g, 0.145 mmol) in DMF (10 mL) in an 8-dram vial under normal atmosphere was added N,N'-diisopropylethylamine (0.076 mL, 0.434 mmol) and N,N'-dimethylethylenediamine (0.047 mL, 0.434 mmol). The reaction mixture was stirred at room temperature for one hour, and LC-MS analysis confirmed the complete conversion of 5 to 7. Reaction mixture was added to ether and cooled at 2° C for several hours to afford crystallization of the product. The product was purified by washing with 3 x 25 mL diethyl ether

and removing the top ether layer from the solid after each wash. After the third wash, the solid was dried under a flow of N₂ to afford **7** as a blue solid (0.112 g, 88%). **LC-MS analysis**: $\lambda_{max} = 738$ nm, MS: [M+H] 865; [M-H] 863 (MW of **7**: 887.14 g/mol). ¹H NMR ((CD₃)₂SO, 600 MHz): δ 8.15 (d, *J* = 8.7 Hz, 2H), 7.99 (d, *J* = 8.6 Hz, 4H), 7.88-7.84 (m, 2H), 7.63 (d, *J* = 8.8 Hz, 2H), 7.61-7.56 (m, 2H), 7.43-7.39 (m, 2H), 5.80 (d, *J* = 12.9 Hz, 2H), 4.18-4.12 (m, 4H), 3.93-3.88 (m, 3H), 3.07-3.01 (m, 4H), 2.86 (s, 2H), 2.81-2.76 (m, 4H), 2.56 (s, 2H), 2.43 (s, 3H), 1.88 (s, 12H), 1.77-1.72 (m, 8H).

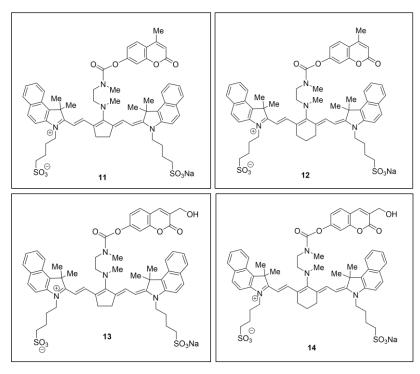


8: To a solution of **6** (3.3, 0.0473 g, 0.0560 mmol) in DMF (4 mL) in an 8-dram vial was added N,N'-Diisopropylethylamine (0.030 mL, 0.167 mmol) and N,N'-dimethylethylenediamine (0.018 mL, 0.167 mmol). The reaction mixture was stirred at room temperature for 80 minutes, and the completion of reaction was confirmed by LC/MS. The reaction mixture was charged with approximately 25mL of ethyl ether, filling the 8-dram vial, and was cooled to 4° C for several hours. The vial

was rewarmed to room temperature, and the top ether layer was removed with a pipette while leaving the crystallized solid at the bottom of the container. More fresh ether was added to the container with the blue solid, and the solid was allowed to settle to the bottom. The ether layer was removed again once the blue solid settled to the bottom of the container. Subsequent washing with ether and removal of the ether was performed until the ether layer is clear. The solid was dried under a constant stream of dry nitrogen to afford **8** as a dark blue solid (0.039 g, 91%, correcting for IR-820 because it contains 80% dye). Drying the solid was only performed directly before usage of the compound. The product was stored in an 8-dram vial filled with ether to separate the blue solid at the bottom from the moisture and oxygen in the air. **LC-MS analysis**: λ_{max} (770 nm), observed [M+(-H)] = 879 [M-(-H)] = 877. **MW** of **8**: 901.17 g/mol.

1.3 General Procedure for the Synthesis of Dyes 11-12, 13-14 and Gabapentin Conjugates 15-16

11-12, 13-14: To a solution of dye-linker (**7, 8**) (0.100 mmol) in acetonitrile (7 mL) was added N,N'diisopropylethylamine (0.052 mL, 0.301 mmol). Triphosgene in acetonitrile (0.015 g, 0.050 mmol) was added dropwise over several minutes, and reaction mixture was stirred at room temperature for 1.5 hours. LC-MS analysis confirmed the complete conversion of dye-linker to the activated acid chloride after 1.5 hours. To the reaction mixture containing the newly formed acid chloride was added N, N'-diisopropylethylamine (0.052 mL, 0.301 mmol), respective alcohol (0.301 mmol), and 4-dimethylaminopyridine (0.006 g, 0.050 mmol). The reaction was stirred at room temperature for 24-48 hr, until LC-MS analysis confirmed the disappearance of the acid chloride and product formation. Mixture was added to ether and cooled at 2° C to afford crystallization of the product. The product was purified by washing with 3 x 25 mL diethyl ether and removing the top ether layer from the solid after each wash. Compounds were subsequently purified by reverse-phase chromatography



(30-100% ACN in H_2O). Both products were dried under a flow of N_2 to afford **11-12** and **13-14** as blue solids (33-87%).

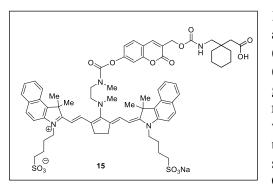
11: Synthesized with 7 and 4methylumbelliferone according to the general procedure outlined above to afford solid blue product (0.015 g, 42%). LC-MS Analysis: $\lambda_{max} = 715 \text{ nm}, \text{ MS: } [M+H] 1067;$ [M-H] 1065 (MW: 1089.3 g/mol). ¹**H NMR** (600 MHz, DMSO- d_6): δ 8.18-8.12 (m, 2H), 8.01-7.96 (m, 6H), 7.59 (t, J = 9.3 Hz, 3H), 7.53 (t, J = 7.7 Hz, 2H), 7.44-7.36 (m,4H), 6.52 (s, 1H), 5.81 (s, 2H), 4.15-4.05 (m, 4H), 3.64-3.56 (m, 4H), 3.17-3.16 (m, 6H), 2.99 (m, 2H), 2.82-2.78 (m, 5H), 2.73 (s, 4H), 1.89-1.87 (m, 8H), 1.76 (s, 12H).

12: Synthesized with **8** and 4methylumbelliferone according to the general procedure outline above

to afford solid blue product (0.010 g, 33%). **LC-MS Analysis**: $\lambda_{max} = 738$ nm, MS: [M+H] 1081; [M-H] 1079 (**MW**: 1103.3 g/mol). ¹**H NMR** (600 MHz, Methanol-*d*₄) δ 8.11 (d, *J* = 9.0 Hz, 2H), 7.97-7.90 (m, 4H), 7.81 (s, 2H), 7.69 (d, *J* = 8.6 Hz, 1H), 7.59-7.47 (m, 5H), 7.45-7.33 (m, 3H), 7.15-7.08 (m, 1H), 6.06 (s, 2H), 4.23-4.15 (m, 4H), 3.65 (s, 2H), 3.55 (s, 1H), 3.40 (d, *J* = 8.8 Hz, 2H), 3.17 (s, 1H), 3.10 (s, 3H), 2.90 (t, *J* = 7.3 Hz, 6H), 2.62 (s, 4H), 2.46 (s, 1H), 2.31 (s, 3H), 2.07-1.98 (m, 8H), 1.90 (s, 12H).

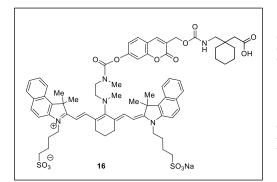
13: Synthesized with **7** and hydroxy-coumarin **26** according to the general procedure outlined above to afford solid blue product (0.087 g, 87%). **LC-MS Analysis**: $\lambda_{max} = 720$ nm, MS: [M+H] 1083; [M-H] 1081 (**MW**: 1105.3 g/mol).

14: Synthesized with **8** and hydroxy-coumarin **26** according to the general procedure outline above to afford solid blue product (0.0929 g, 82%). **LC-MS Analysis**: $\lambda_{max} = 743$ nm, MS: [M+H] 1097; [M-H] 1095 (**MW**: 1119.3 g/mol).



15-16: To a solution of **13** or **14** (0.114 mmol) in acetonitrile (8 mL) was added N,N'-diisopropylethylamine (0.099 mL, 0.569 mmol) and 4-dimethylaminopyridine (0.014 g, 0.114 mmol). 4-nitrophenyl chloroformate (0.057 g, 0.285 mmol) in acetonitrile was added dropwise to reaction mixture at 0° C over 5-10 min. Reaction was warmed up to room temperature and stirred for 1.5-5 hr, until LC-MS analysis confirmed the disappearance of starting material and formation of the activated carbonate. Gabapentin (0.058 g, 0.342 mmol) in acetonitrile was then

added to the reaction mixture and stirred at room temperature for 12-24 hr until LC-MS analysis confirmed the disappearance of activated carbonate and formation of the desired product. The reaction mixture was filtered to remove excess Gabapentin and then filtrate added to ether and cooled at 2° C to afford crystallization of the product. The product was purified by washing with 3 x 25 mL diethyl ether and removing the top ether layer from the solid after each wash. Compounds were subsequently purified by reverse-phase chromatography (30-100% ACN in H₂O). Both products were dried under a flow of N₂ to afford **15-16** as blue solids (28-43%).

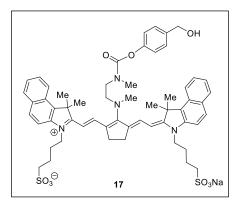


15: Synthesized with **13** according to the general procedure outline above to afford solid blue product (0.045 g, 43%). **LC-MS Analysis:** $\lambda_{max} = 715$ nm, MS: [M-H (+Na⁺)] 1301 (**MW**: 1302.5 g/mol).

16: Synthesized with 14 according to the general procedure outline above to afford solid blue product (0.042 g, 28%). LC-MS Analysis: $\lambda_{max} = 736$ nm, MS: [M+H (+Na⁺)] 1316; [M-H (+Na⁺)] 1314 (MW: 1316.6 g/mol).

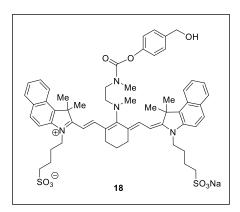
1.4 General Procedure for Synthesis of NIR Dyes 17-18 and Gabapentin Conjugates 19-20

17-18: To a solution of dye-linker (**7**, **8**) (0.100 mmol) in acetonitrile (7 mL) was added N,N'diisopropylethylamine (0.052 mL, 0.301 mmol). Triphosgene in acetonitrile (0.015 g, 0.050 mmol) was added dropwise over several minutes, and reaction mixture was stirred at room temperature for 1.5 hours. LC-MS analysis confirmed the complete conversion of dye-linker to the activated acid chloride after 1.5 hours. To the reaction mixture containing the newly formed acid chloride was added N, N'-diisopropylethylamine (0.052 mL, 0.301 mmol), 4-hydroxybenzyl alcohol (0.301 mmol), and 4-dimethylaminopyridine (0.006 g, 0.050 mmol). The reaction was stirred at room temperature for 24-48 hr, until LC-MS analysis confirmed the disappearance of the acid chloride and product formation. Mixture was added to ether and cooled at 2° C to afford crystallization of the product. The product was purified by washing with 3 x 25 mL diethyl ether and removing the top ether layer from the solid after each wash. Compounds were subsequently purified by reverse-phase chromatography (30-100% ACN in H₂O). Both products were dried under a flow of N₂ to afford **17-18** as blue solids (72-79%).



17: Synthesized with **7** according to the general procedure outlined above to afford solid blue product (0.075 g, 72%). **LC-MS Analysis:** $\lambda_{max} = 718$ nm, MS: [M+H] 1015; [M-H] 1013 (MW: 1037.3 g/mol). ¹H NMR (600 MHz, Methanol-*d*₄): δ 8.20 (d, J = 8.2 Hz, 1H), 8.01-7.94 (m, 5H), 7.77 (d, J = 8.8 Hz, 1H), 7.56 (d, J = 8.7 Hz, 3H), 7.47-7.44 (m, 3H), 7.38-7.35 (m, 3H), 7.25 (d, J = 8.7 Hz, 1H), 6.95-6.92 (m, 1H), 6.14 (d, J = 13.7 Hz, 2H), 4.26-4.23 (m, 2H), 4.10 (m, 4H), 3.86-3.82 (m, 2H), 3.69 (s, 2H), 3.57 (s, 2H), 3.14-3.10 (m, 2H), 3.05-2.99 (m, 3H), 2.81 (s, 3H), 2.77-2.74 (m, 2H), 2.61-2.55 (m, 2H), 1.95-1.91 (m, 8H), 1.88-1.78 (m, 12H).

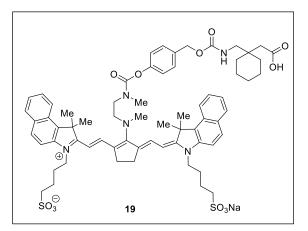
18: Synthesized with **8** according to the general procedure outlined above to afford solid blue product (0.093 g, 79%). **LC-MS Analysis**: $\lambda_{max} = 736$ nm, MS: [M+H] 1029; [M-H] 1027 (**MW**: 1051.3 g/mol). ¹**H NMR** (600 MHz, DMSO-*d*₆) δ 8.16 – 8.10 (m, 1H), 8.07 (d, *J* = 8.6 Hz, 1H), 8.02 – 7.97

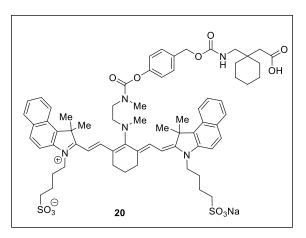


(m, 2H), 7.95 (s, 1H), 7.86 – 7.78 (m, 3H), 7.68 – 7.54 (m, 5H), 7.49 (t, J = 7.6 Hz, 2H), 7.45 – 7.39 (m, 1H), 7.35 (t, J = 7.5 Hz, 2H), 6.05 – 5.93 (m, 2H), 4.51 – 4.33 (m, 2H), 4.24 – 4.06 (m, 5H), 3.96 (s, 1H), 3.83 – 3.69 (m, 3H), 3.17 (s, 1H), 3.06 (s, 1H), 2.98 (s, 1H), 2.81 – 2.69 (m, 3H), 2.56 (s, 2H), 1.98 – 1.77 (m, 13H), 1.75 (s, 12H).

19-20: To a solution of **17** or **18** (0.114 mmol) in acetonitrile (8 mL) was added N,N'-diisopropylethylamine (0.099 mL, 0.569 mmol) and 4-dimethylaminopyridine (0.014 g, 0.114 mmol). 4-nitrophenyl chloroformate (0.057 g, 0.285 mmol) in acetonitrile

was added dropwise to reaction mixture at 0° C over 5-10 min. Reaction was warmed up to room temperature and stirred for 1.5-5 hr, until LC-MS analysis confirmed the disappearance of starting material and formation of the activated carbonate. Gabapentin (0.058 g, 0.342 mmol) in acetonitrile was then added to the reaction mixture and stirred at room temperature for 12-24 hr until LC-MS analysis confirmed the disappearance of activated carbonate and formation of the desired product. The reaction mixture was filtered to remove excess Gabapentin and then filtrate added to ether and cooled at 2° C to afford crystallization of the product. The product was purified by washing with 3 x 25 mL diethyl ether and removing the top ether layer from the solid after each wash. Compounds were subsequently purified by reverse-phase chromatography (30-100% ACN in H₂O). Both products were dried under a flow of N₂ to afford **19-20** as blue solids (36-47%).

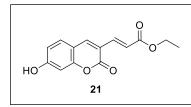




19: Synthesized with **17** according to the general procedure outline above to afford solid blue product (0.035 g, 36%). **LC-MS Analysis**: λ_{max} =715 nm, MS: [M+H] 1212, [(MW+2)/2)] 606; [M-H] 1210 (**MW**: 1234.5 g/mol). **:** ¹**H NMR** (600 MHz, Methanol-*d*₄): δ 8.08 (d, *J* = 7.2 Hz, 1H), 8.03 (d, *J* = 8.5 Hz, 1H), 7.94-7.82 (m, 6H), 7.57-7.51 (m, 3H), 7.48-7.42 (m, 3H), 7.35 (t, *J* = 7.4 Hz, 3H), 6.94 (d, *J* = 6.7 Hz, 1H), 5.74 (d, *J* = 12.0 Hz, 2H), 4.29 (s, 1H), 4.15-4.03 (m, 4H), 3.79 (s, 1H), 3.75-3.64 (m, 2H), 3.22 (s, 4H), 3.05-2.95 (m, 2H), 2.95-2.78 (m, 10H), 2.03-1.88 (m, 12H), 1.86-1.73 (m, 5H), 1.66 (s, 7H), 1.52-1.26 (m, 10H).

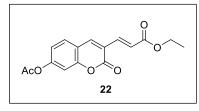
20: Synthesized with **18** according to the general procedure outline above to afford solid blue product (0.052 g, 47%). **LC-MS Analysis**: λ_{max} =733 nm, MS: [M-H] 1224 (**MW**: 1248.5 g/mol).

1.5 Synthesis of Hydroxy-Coumarin 26



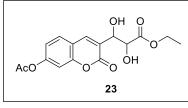
21: To a solution of 2,4-dihydroxybenzaldehyde (1.413g, 10.3mmol) in ethanol (30mL) in an oven-dried 100mL round bottom flask was added diethylglutaconate (1.9mL, 10.7mmol) and catalytic piperidine (3 drops). The reaction mixture was refluxed under Argon atmosphere for 24 hours. The now red solution was cooled to room temperature and then cooled further to -20°C for one hour. The resulting yellow

precipitate was isolated by vacuum filtration and dried to yield a powdery yellow solid (2.1342g, 80%). ¹**H NMR** (400 MHz, DMSO- d_6) δ 10.90 (s, 1H), 8.42 (s, 1H), 7.60 – 7.44 (m, 2H), 6.88 – 6.79 (m, 2H), 6.73 (dd, J = 2.2, 0.6 Hz, 1H), 4.16 (q, J = 7.1 Hz, 2H), 1.24 (t, J = 7.1 Hz, 3H). ¹³**C NMR** (101 MHz, DMSO- d_6) δ 166.70, 163.31, 159.66, 155.81, 145.95, 131.35, 120.19, 116.73, 114.48, 111.98, 102.41, 102.39, 60.52, 14.64.



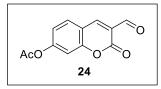
22: Phenol **21** (2.1178g, 8.2mmol) was dissolved in pyridine (15mL) in a 100mL round bottom flask and then charged dropwise with acetic anhydride (15mL) and stirred at room temperature for 30 minutes. The reaction mixture was then poured into an ice water mixture and stirred for 10 minutes. The resulting precipitate was collected by vacuum filtration and dried overnight in a vacuum oven to yield a

white solid (2.42g, 98%). ¹**H NMR** (400 MHz, DMSO-*d*₆) δ 8.56 (s, 1H), 7.77 (d, *J* = 8.5 Hz, 1H), 7.54 (dd, *J* = 16.0, 0.6 Hz, 1H), 7.35 – 7.30 (m, 1H), 7.20 (dd, *J* = 8.5, 2.2 Hz, 1H), 6.97 – 6.89 (m, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 2.30 (s, 3H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³**C NMR** (101 MHz, DMSO-*d*₆) δ 169.15, 166.41, 159.13, 154.28, 154.15, 144.67, 130.62, 122.23, 120.89, 117.27, 110.39, 109.99, 60.73, 25.24, 21.32, 14.58.



23: α , β -unsaturated ester **22** (0.930g, 3.08mmol) was dissolved in dichloromethane (15mL) and charged with a 1:1 w/w solution of N-methylmorpholine-N-oxide (0.721g, 6.16mmol) in water (0.8mL) and 3.0mL of 2.5wt% OsO₄ in t-BuOH. The solution was stirred at room temperature for 10 hours until completion was confirmed by TLC. The reaction was then quenched with water (50mL) and

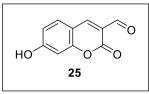
extracted with ethyl acetate (3 x 30mL). The combined organic layers were washed with brine (30mL), dried over Na₂SO₄, filtered, and concentrated under reduced pressure to yield a white solid (0.7486g, 72%). ¹**H NMR** (400 MHz, DMSO- d_6) δ 8.02 (s, 1H), 7.83 (d, J = 8.5 Hz, 1H), 7.30 (d, 1H), 7.17 (dd, J = 8.4, 2.2 Hz, 1H), 5.72 (d, J = 6.7 Hz, 1H), 5.18 (d, J = 7.1 Hz, 1H), 4.99 – 4.93 (m, 1H), 4.39 (dd, J = 7.0, 2.6 Hz, 1H), 4.03 (q, J = 7.1 Hz, 2H), 2.31 (s, 3H), 1.17 (t, J = 7.1 Hz, 3H). ¹³**C NMR** (101 MHz, DMSO- d_6) δ 171.78, 168.84, 159.53, 153.05, 152.38, 139.36, 129.14, 128.00, 116.93, 110.31, 109.81, 71.36, 60.30, 20.91, 20.75, 14.18.



24: Diol **23** (0.7486g 2.23mmol) was added to a slurry of silica gel (5g) in dichloromethane (10mL). While stirring vigorously, the reaction mixture was charged dropwise with a solution of sodium periodate (NaIO₄, 0.6333g, 2.96mmol) over 5 minutes and then stirred at room temperature for 2 hours. The reaction mixture was filtered through a cake of celite and washed with

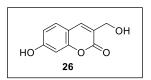
copious amounts of ethyl acetate. The ethyl acetate was washed with water (30mL) and brine (30mL), dried over Na₂SO₄, filtered, and dry-loaded onto celite. The crude reaction mixture was purified by silica gel chromatography using a 5-50% ethyl acetate in hexane, and the product eluted in 45%. The relevant fractions were combined and concentrated under reduced pressure to yield a

white solid (0.396g, 76%). ¹**H NMR** (400 MHz, DMSO-*d*₆) δ 10.03 (s, 1H), 8.71 – 8.68 (m, 1H), 8.07 – 8.01 (m, 1H), 7.41 – 7.37 (m, 1H), 7.27 (dd, *J* = 8.5, 2.2 Hz, 1H), 2.33 (s, 3H).



25: Aldehyde **24** (0.395g, 1.7mmol) was dissolved in 30% w/w NH₄OH in water (5mL) and charged with acetonitrile (5mL) slowly. The solution was then stirred 10 minutes at room temperature before being quenched with ethyl acetate (30mL). The organic layer was then washed with 1M HCl (2 x 20mL), dried over Na₂SO₄, filtered, and concentrated under reduced

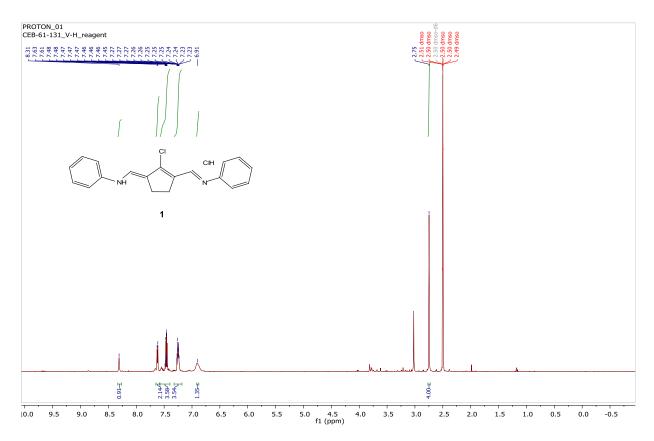
pressure to yield a yellow solid (0.2605g, 81%). ¹**H NMR** (400 MHz, DMSO- d_6) δ 9.96 (s, 1H), 8.61 – 8.56 (m, 1H), 7.82 (d, J = 8.6 Hz, 1H), 6.88 (dd, J = 8.6, 2.3 Hz, 1H), 6.80 – 6.76 (m, 1H).



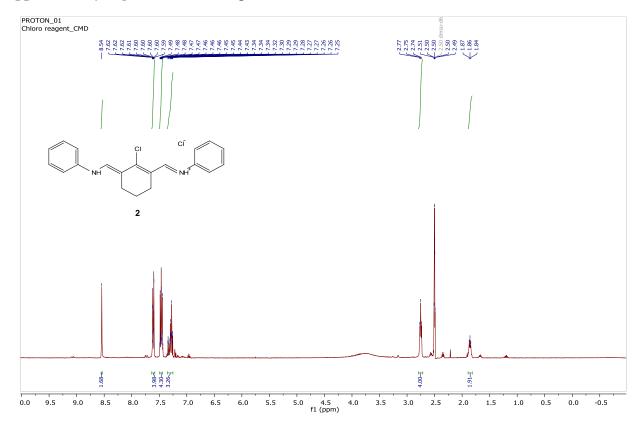
26: Phenol **25** (0.0541g, 0.28mmol) was dissolved in methanol (5mL) and cooled to 0°C. Solid NaBH₄ (0.013g, 0.34mmol) was added slowly, and the reaction mixture was stirred at 0°C for 30 minutes. The mixture was then diluted with ethyl acetate (20mL) and washed with saturated NH₄Cl(aq)

solution (20mL), dried over Na2SO4, filtered, and concentrated under reduced pressure to yield a yellow solid (0.0465g, 85%).

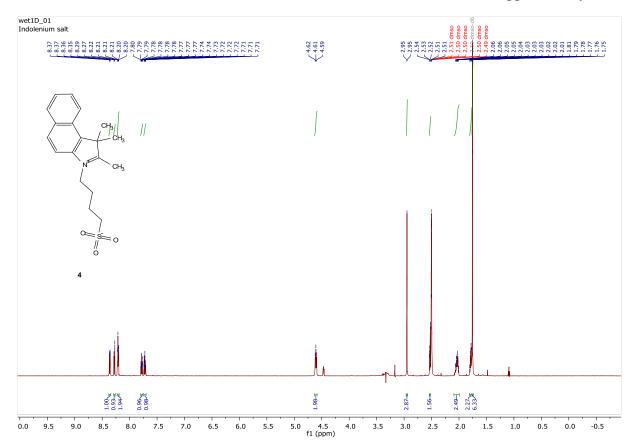
2 NMR and LC-MS Spectra



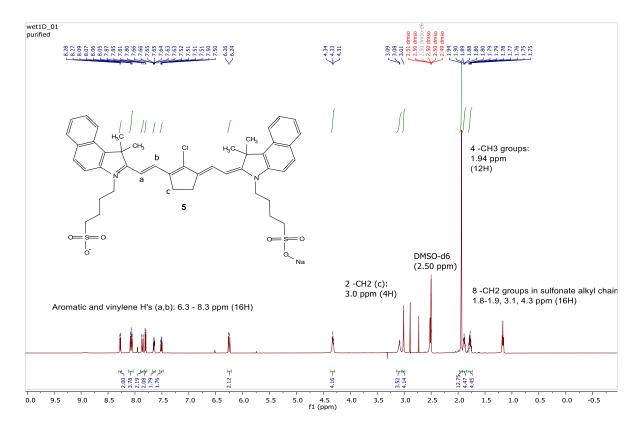
Supplementary Figure 1. ¹H NMR spectrum of 1.



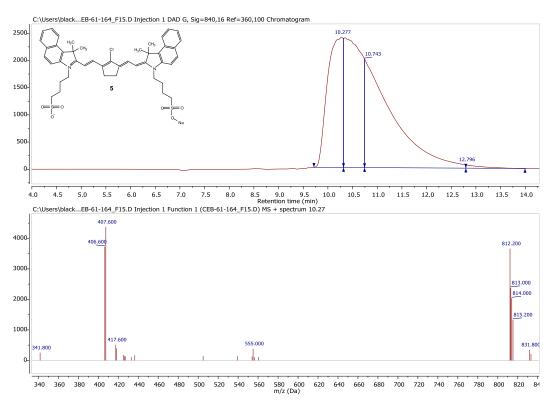
Supplementary Figure 2. ¹H NMR spectrum of 2.



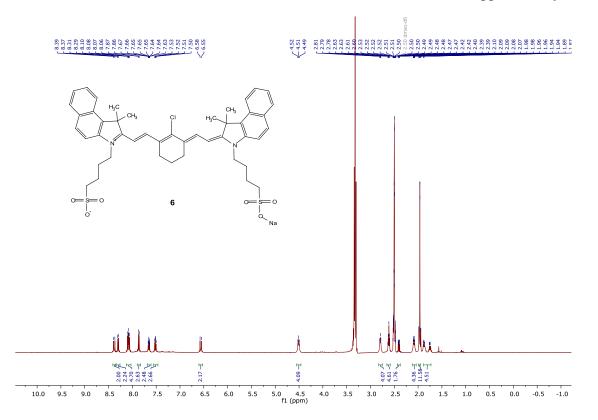
Supplementary Figure 3. ¹H NMR spectrum of 4.



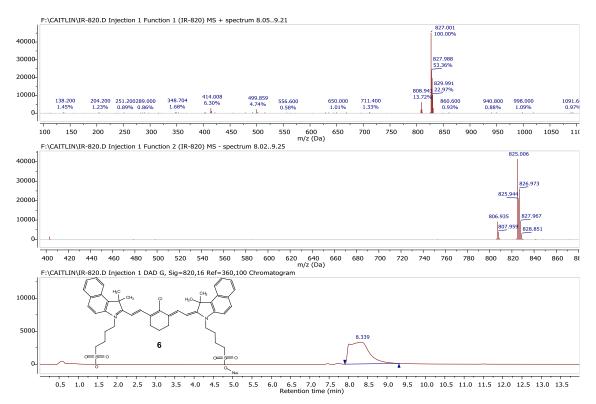
Supplementary Figure 4. ¹H NMR spectrum of 5.



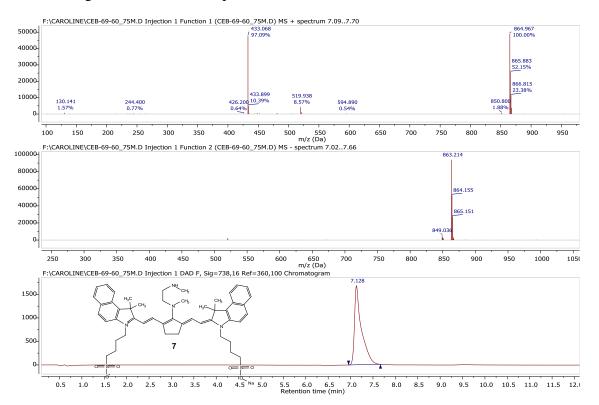
Supplementary Figure 5. LC-MS data of **5**, $\lambda_{max} = 844$ nm. *Top:* Chromatogram at 840 nm with pure **5** at T_{ret} = 10.3 minutes. *Bottom:* MS in positive mode with **5** seen at 812.2 [MW without Na⁺ ion: 811.26 g/mol].



Supplementary Figure 6. ¹H NMR spectrum of 6.

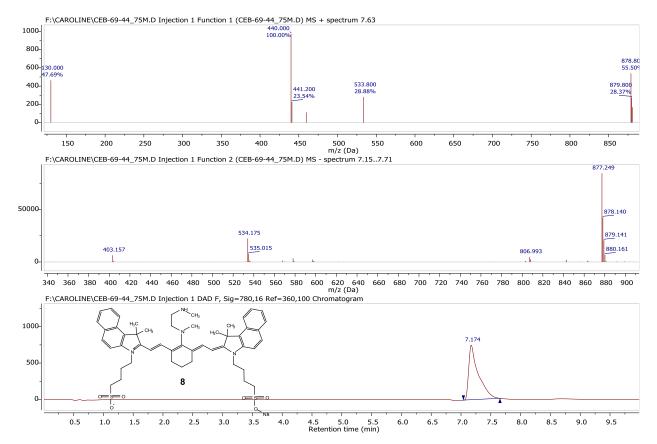


Supplementary Figure 7. LC-MS data of **6**, $\lambda_{max} = 820$ nm. *Top:* MS in positive mode with **6** seen at 827.0 [MW without Na⁺ ion: 826.5 g/mol]. *Middle:* MS in negative mode with **6** seen at 825.0. *Bottom:* Chromatogram at 820 nm with pure **6** at T_{ret} = 8.0 – 8.9 minutes.

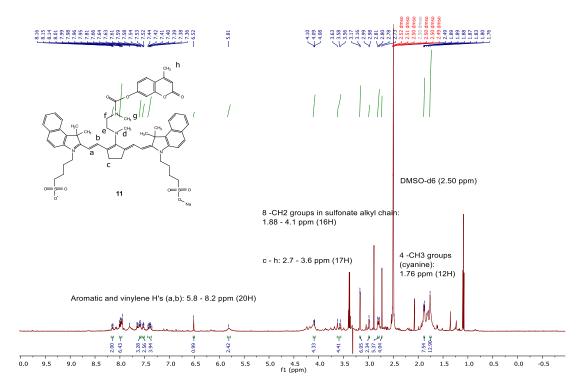


Supplementary Figure 8. LC-MS data of **7**, $\lambda_{max} = 738$ nm. *Top*: MS in positive mode with **7** at 865.0 and 433.1 ((MW+2)/2) [MW without Na⁺ ion: 864.2 g/mol]. *Middle*: MS in negative mode with **7** at 863.2. *Bottom*: Chromatogram at 738 nm with **7** at T_{ret} = 7.1 minutes.

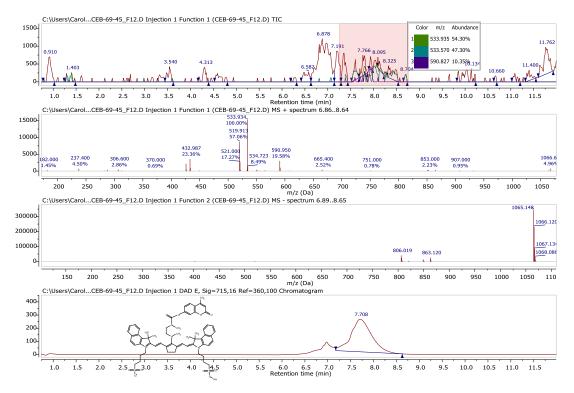
Supplementary Material



Supplementary Figure 9. LC-MS data of **8**, $\lambda_{max} = 778$ nm. *Top*: MS in positive mode with **8** at 878.8 and 440.0 ((MW+2)/2) [MW without Na⁺ ion: 878.2 g/mol]. *Middle:* MS in negative mode with **8** at 877.2. *Bottom:* Chromatogram at 780 nm with **8** at T_{ret} = 7.2 minutes.

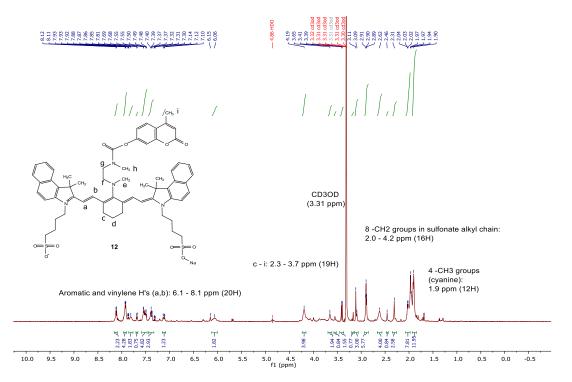


Supplementary Figure 10. ¹H NMR spectrum of 11.

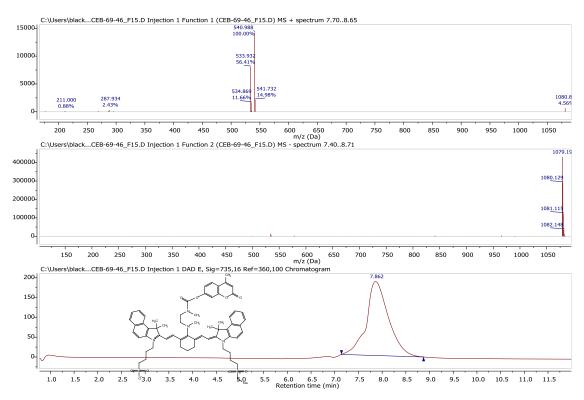


Supplementary Figure 11. LC-MS data of **11**, $\lambda_{max} = 715$ nm. *Top:* TIC showing the peak purity of the major LC peak (7.2-8.5 min) to be majority product masses [533.9/533.5 (MW+2)/2]. *Upper middle*: MS in positive mode with **11** at 1066.7 and 533.9 ((MW+2)/2) [MW without Na⁺ ion: 1066.3

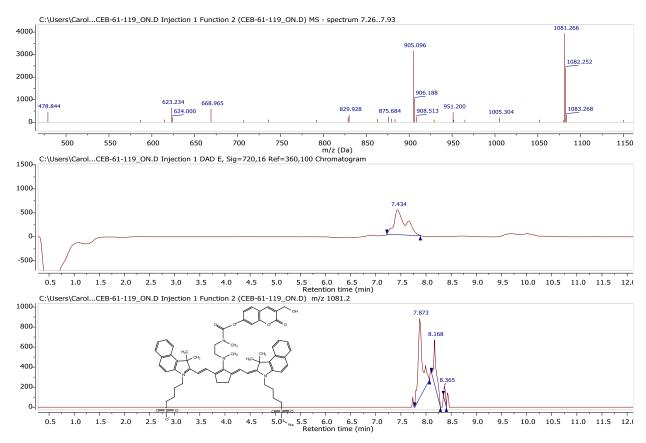
g/mol]. *Lower middle:* MS in negative mode with **11** at 1065.1. *Bottom:* Chromatogram at 715 nm with **11** at $T_{ret} = 7.2-8.5$ minutes.



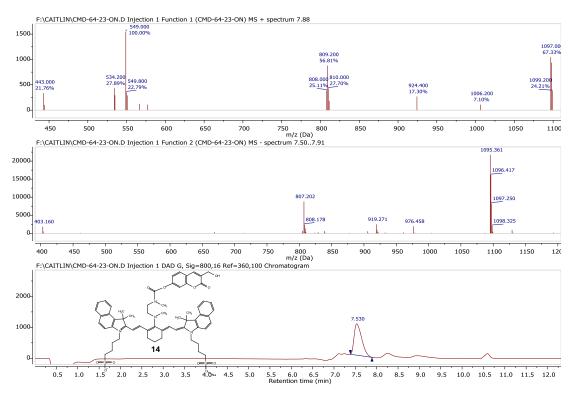
Supplementary Figure 12. ¹H NMR spectrum of 12.



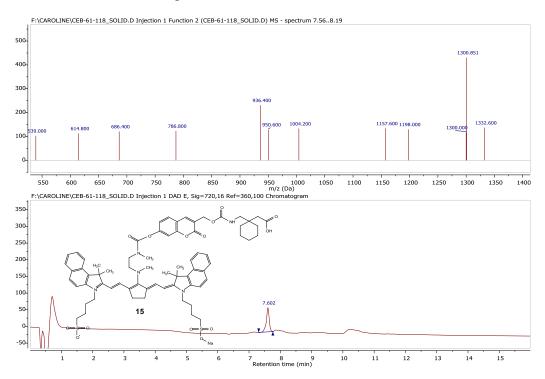
Supplementary Figure 13. LC-MS data of **12**, $\lambda_{max} = 735$ nm. *Top:* MS in positive mode with **12** at 1080.8 and 541.0 ((MW+2)/2) [MW without Na⁺ ion: 1080.4 g/mol]. *Middle:* MS in negative mode with **12** at 1079.2. *Bottom:* Chromatogram at 735 nm with **12** at T_{ret} = 7.9 minutes.



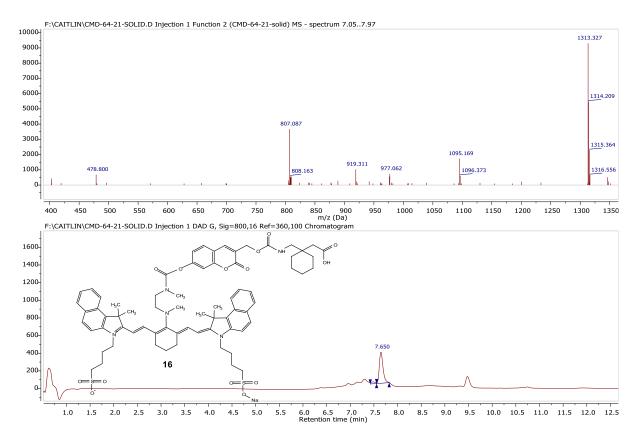
Supplementary Figure 14. LC-MS data of **13**, $\lambda_{max} = 720$ nm. *Top:* MS in negative mode with **13** at 1081.3 [MW without Na⁺ ion: 1082.3 g/mol]. *Middle:* Chromatogram at 720 nm with **13** at T_{ret} = 7.5 minutes. *Bottom:* EIC of product m/z 1081.2 in MS negative mode.



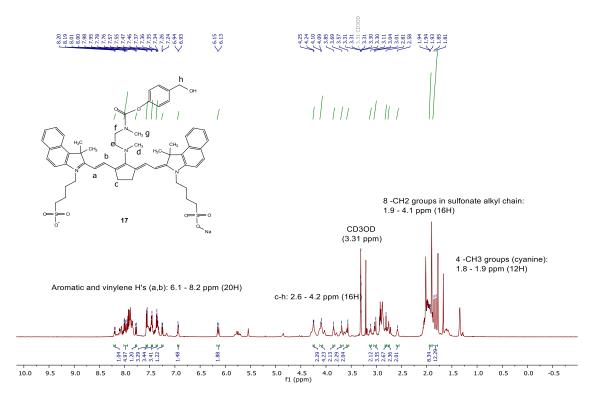
Supplementary Figure 15. LC-MS data of **14**, $\lambda_{max} = 743$ nm. *Top:* MS in positive mode with **14** at 1097.0 and 549.0 ((MW+2)/2) [MW without Na⁺ ion: 1096.3 g/mol]. *Middle:* MS in negative mode with **14** at 1095.4. *Bottom:* Chromatogram at 800 nm with **14** at T_{ret} = 7.5 minutes.



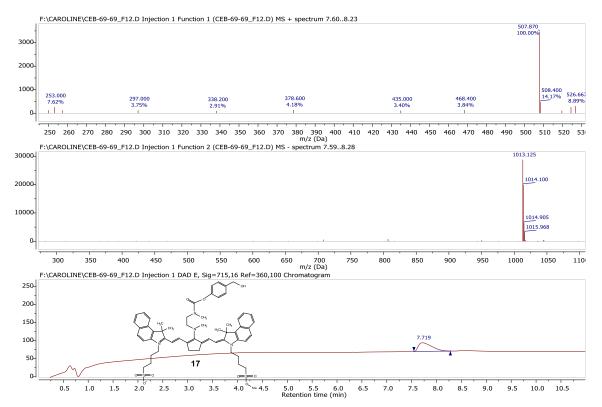
Supplementary Figure 16. LC-MS data of **15**, $\lambda_{max} = 715$ nm. *Top:* MS in negative mode with **15** at 1300.9 [MW: 1301.5 g/mol]. *Bottom*: Chromatogram at 720 nm with **15** at T_{ret} = 7.6 – 8 minutes.



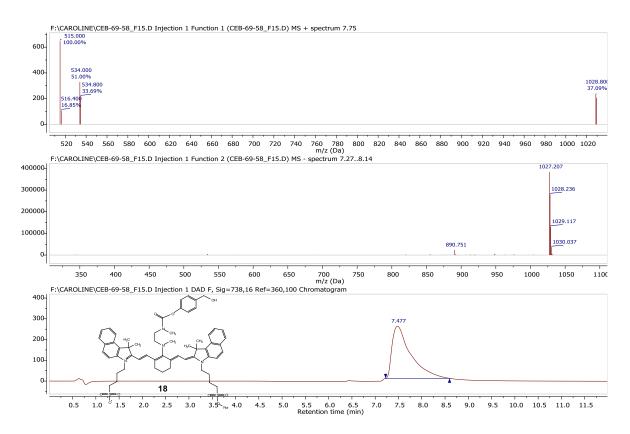
Supplementary Figure 17. LC-MS data of **16**, $\lambda_{max} = 735$ nm. *Top:* MS in negative mode with **16** at 1313.3/1314.2 [MW: 1315.5 g/mol]. *Bottom*: Chromatogram at 800 nm with **16** at T_{ret} = 7. - 8 minutes.



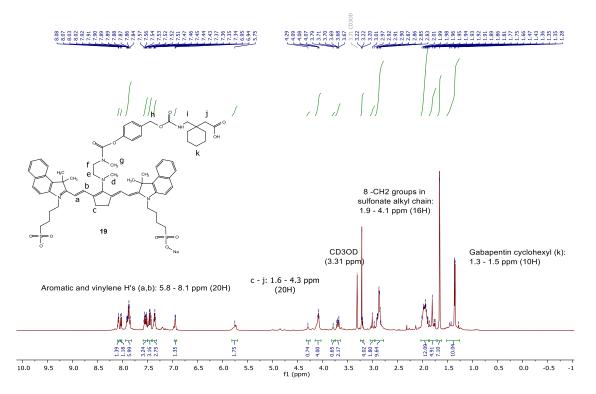
Supplementary Figure 18. ¹H NMR spectrum of 17.



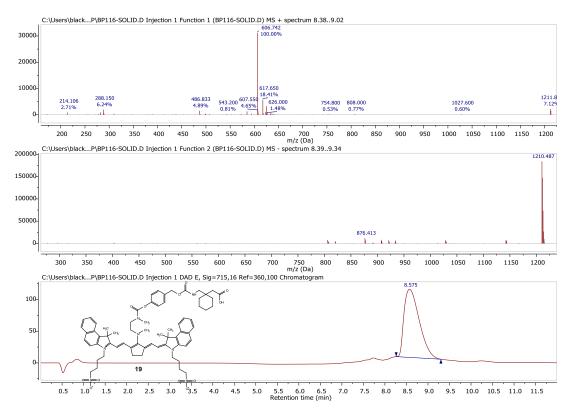
Supplementary Figure 19. LC-MS data of **17**, $\lambda_{max} = 718$ nm. *Top*: MS in positive mode with **17** at 507.9 ((MW+2)/2) [MW without Na⁺ ion: 1014.3 g/mol]. *Middle*: MS in negative mode with **17** at 1013.1. *Bottom*: Chromatogram at 715 nm with **17** at T_{ret} = 7.7 minutes.



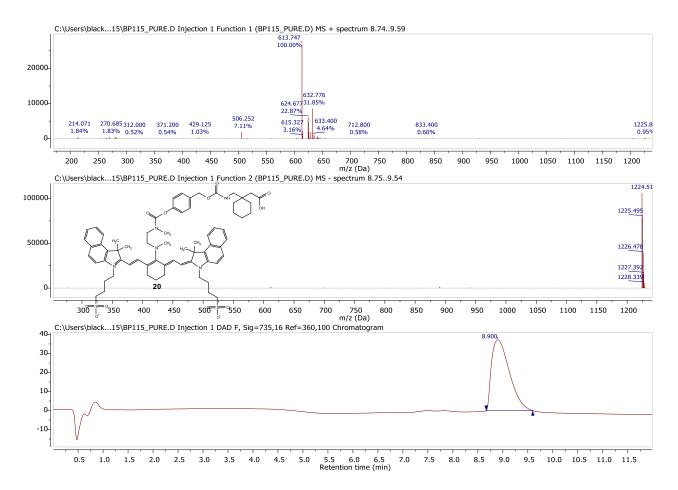
Supplementary Figure 20. LC-MS data of **18**, $\lambda_{max} = 738$ nm. *Top*: MS in positive mode with **18** at 1028.8 and 515.0 ((MW+2)/2) [MW without Na⁺ ion: 1028.3 g/mol]. *Middle:* MS in negative mode with **18** at 1027.2. *Bottom:* Chromatogram at 738 nm with **18** at T_{ret} = 7.5 minutes.



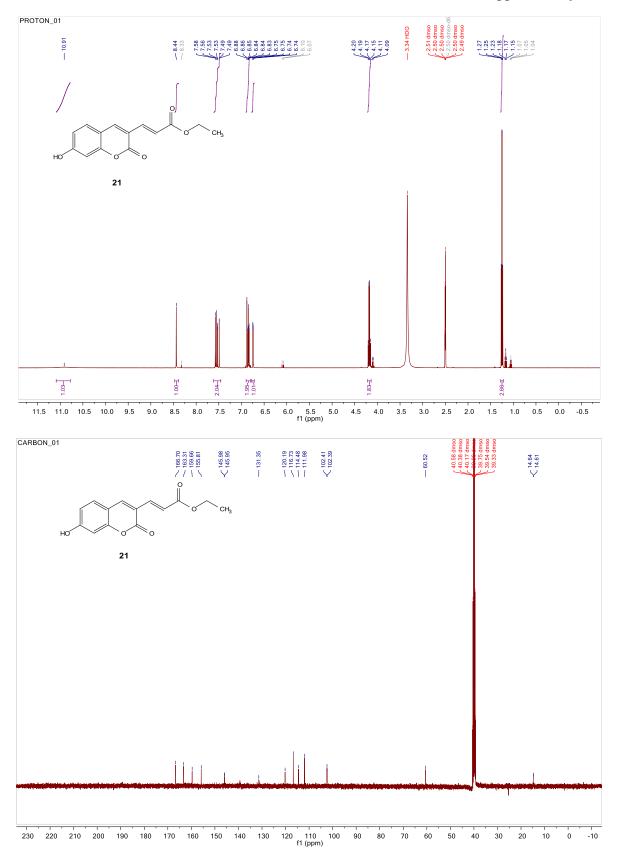
Supplementary Figure 21. ¹H NMR spectrum of 19.



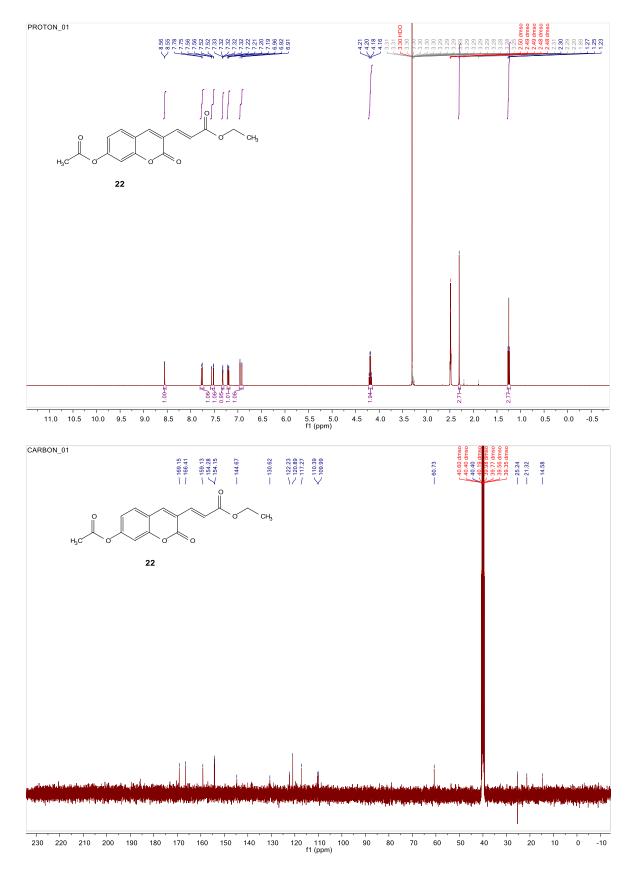
Supplementary Figure 22. LC-MS data of **19**, $\lambda_{max} = 715$ nm. *Top:* MS in positive mode with **19** at 1211.9 and 606.7 ((MW+2)/2) [MW without Na⁺ ion: 1211.5 g/mol]. *Middle:* MS in negative mode with **19** at 1210.5. *Bottom:* Chromatogram at 715 nm with **19** at T_{ret} = 8.6 minutes.



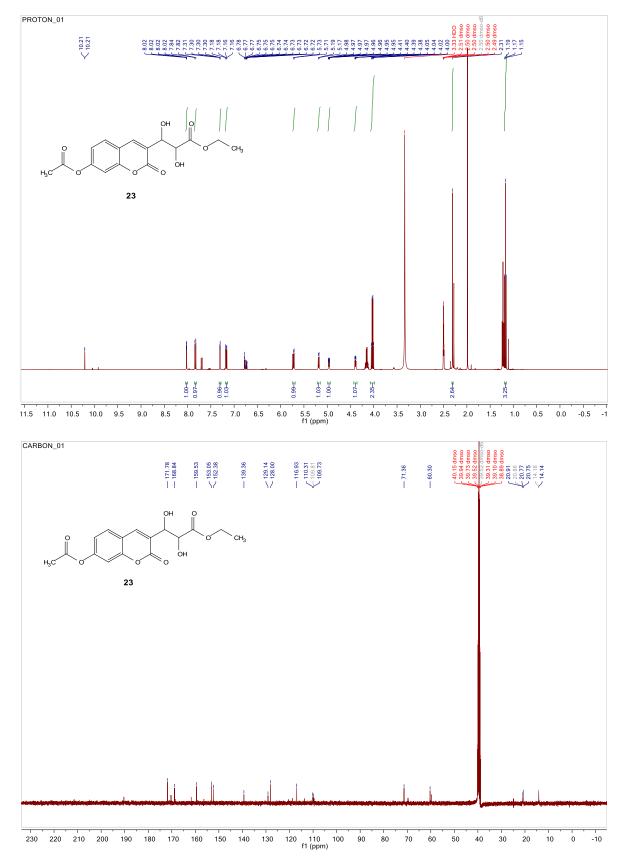
Supplementary Figure 23. LC-MS data of **20**, $\lambda_{max} = 733$ nm. Top: MS in positive mode with **20** at 1225.8 and 613.7 ((MW+2)/2) [MW without Na⁺ ion: 1225.6 g/mol]. Middle: MS in negative mode with **20** at 1224.5. Bottom: Chromatogram at 735 nm with **20** at T_{ret} = 8.9 minutes.



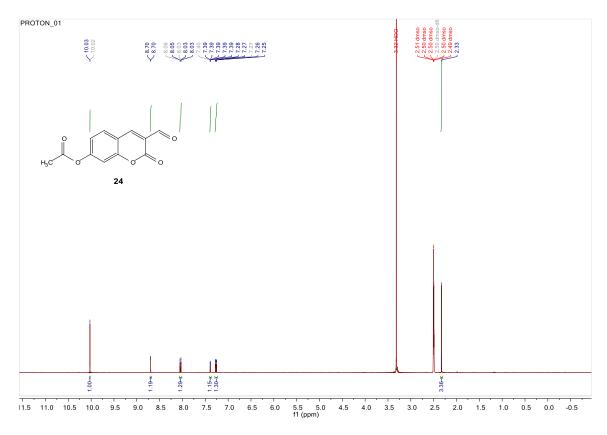
Supplementary Figure 24. ¹H and ¹³C NMR spectra of 21.



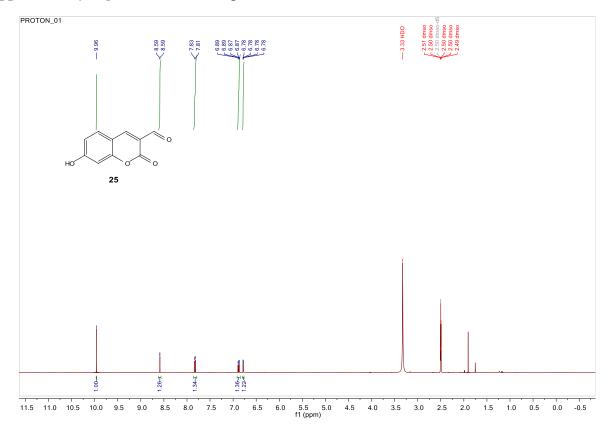
Supplementary Figure 25. ¹H and ¹³C NMR spectra of 22.



Supplementary Figure 26. ¹H and ¹³C NMR spectra of 23.



Supplementary Figure 27. ¹H NMR spectrum of 24.



Supplementary Figure 28. ¹H NMR spectrum of 25.