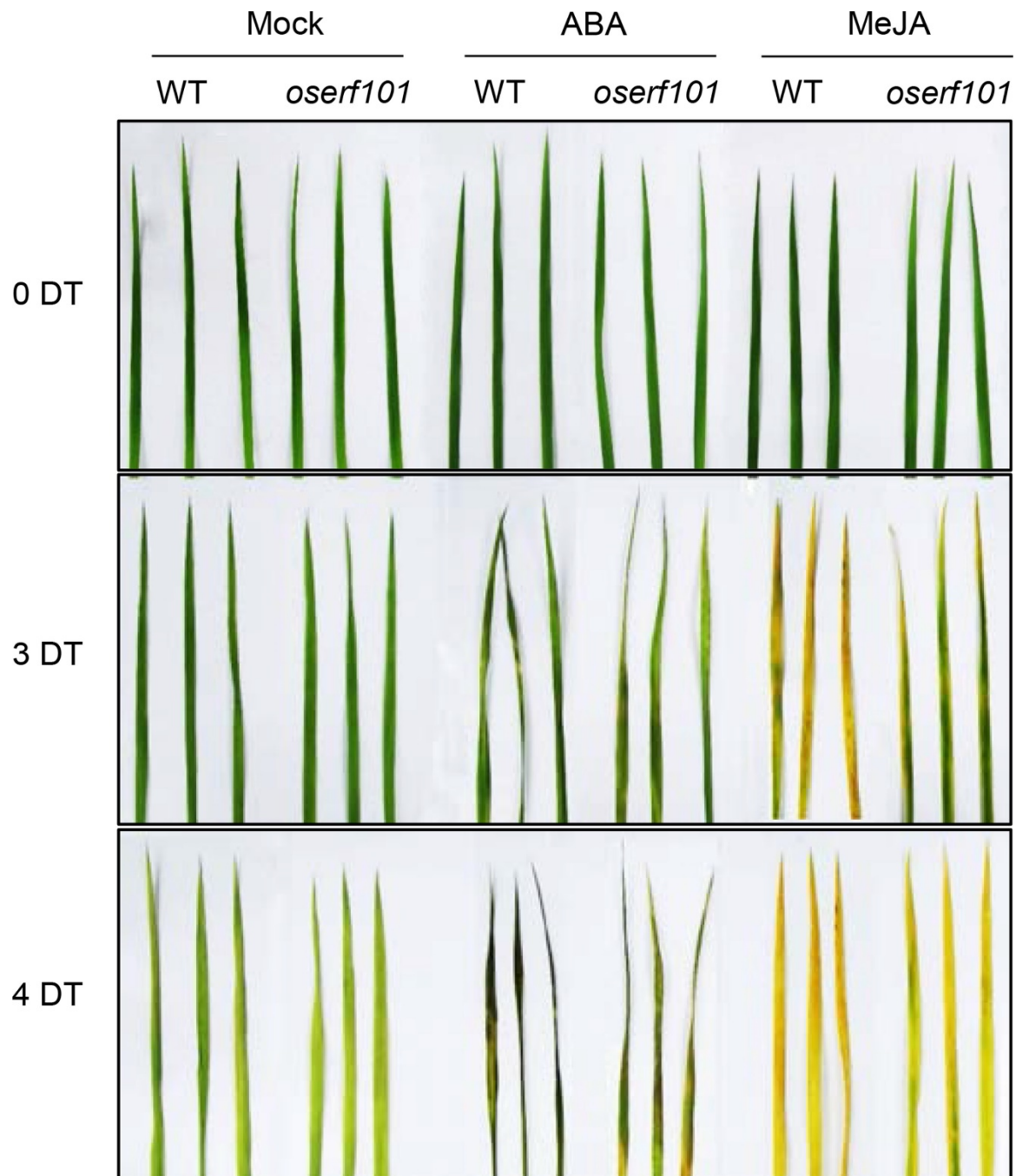


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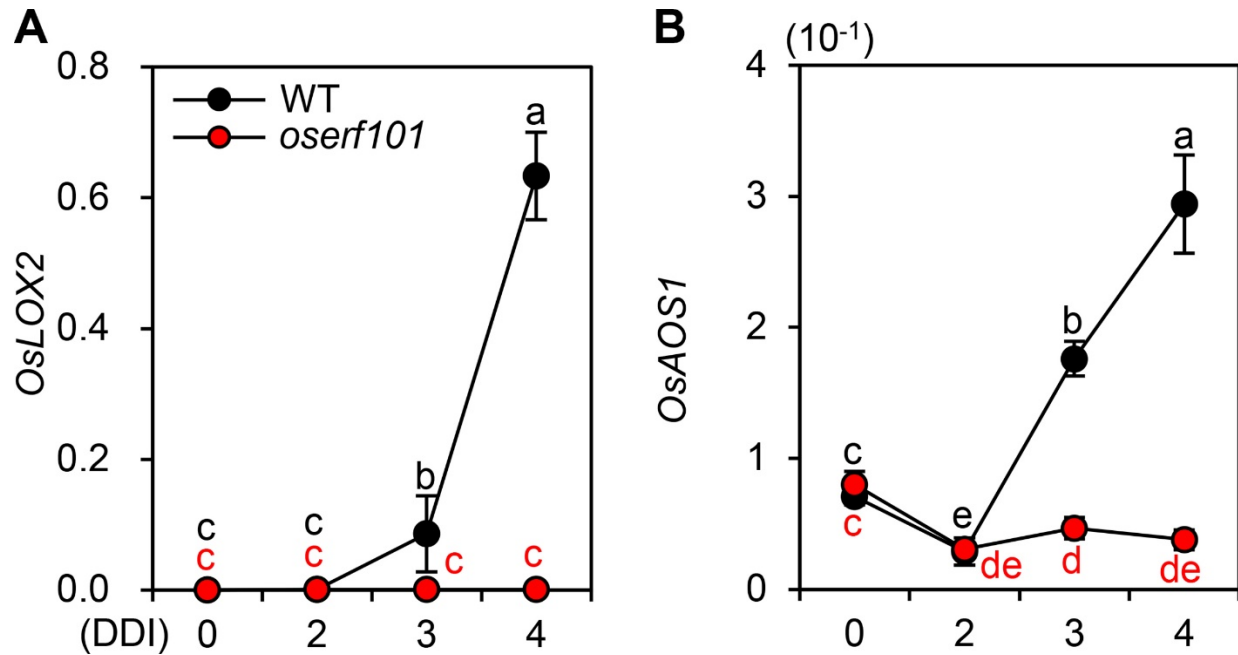
OsERF101	1	-----MVTALAHVIRAPDLHLPHH
Arabidopsis thaliana	1	-----MVSALSRVIENPTDPPV---
Brachypodium distachyon	1	-----MVSALSHVIRAPDGPD---
Sorghum bicolor	1	-----MVSALSHVIRAPDQ----
Hordeum vulgare	1	-----MVSALSHVIRAPDQ----
Glycine max	1	MQYGKLPSSQASEEKEGTVEFDAYNNYASWSSETDMSEMVSALSQVMGTTNRNPH--M
OsERF101	21	PSSASAAAAHPQASSFYPTAAAAASSP-----SD-----QLAAAAAAEEQGRRRHYRG
Arabidopsis thaliana	18	-----KQELDKS-----DQHQPDDQERRRHRYRG
Brachypodium distachyon	17	-----QQQGFYPTAAAVPGQQ-----QQQQQIGIAAAAAAAEEQGRKRHYRG
Sorghum bicolor	1	-----MLYAGAPGLRQGDQPQAAHHHPGGHVA AAAEEEQAGRHHRYRG
Hordeum vulgare	16	-----EPAYYPAGPAAVSRE-----QQH-----QHAAATAEEQGRKRHYRG
Glycine max	59	P-----QSNSEFSSSTSVVKDQPQPYP-----SQPPFDQVTKKPHYRG
OsERF101	70	VRQRPWGKWAAEIRDPKKAARVWLGTFTDAEDAAIAYDEAALRFKGTAKLNFPERVQGR
Arabidopsis thaliana	42	VRQRPWGKWAAEIRDPKKAARVWLGTFTETAEFAALAYDRAALKFKGTAKLNFPERVQGP
Brachypodium distachyon	60	VRQRPWGKWAAEIRDPKKAARVWLGTFTDAEDAAIAYDEAALRFKGTAKLNFPERVQGR
Sorghum bicolor	46	VRQRPWGKWAAEIRDPKKAARVWLGTFTDAEDAAIAYDEAALRFKGTAKLNFPERVQGR
Hordeum vulgare	52	VRQRPWGKWAAEIRDPKKAARVWLGTFTDAEDAAIAYDEAALRFKGTAKLNFPERVQGR
Glycine max	97	VRQRPWGKWAAEIRDPKKAARVWLGTFTETAEADAALAYDKAALKFKGTAKLNFPERLHQN
OsERF101	130	TDLGELVTRGIPPAAT-HGGGY-----YESSSP-----AAGACPP--PRQQQTVVVPYD
Arabidopsis thaliana	102	TTTTTISH-----APRCVSESMNSPPPRPGPPSTTTTSWPMTYNQD
Brachypodium distachyon	120	TDLGFVVTRGIPDRSSLLHHQQH-----YPC-----STAMRPP--QQQTVVVPYD
Sorghum bicolor	106	TDLGELVTRGIPADHH-----PAAVT-----LAAMPFRH-QHGHQTVVVPYD
Hordeum vulgare	112	TDLGFVVTRGIPDL--QQQHH-----YPAAVG-----APAMRPP--LHQQQAVVVPYD
Glycine max	157	VPYMQQHQQGSSNRNVFPFHATSSSTSSSATCSVSSLD---AVAPPSLSTH--DHQVFPN
OsERF101	176	LMRYAQLLQGGVG-----GSMYFPGGAATMSSS
Arabidopsis thaliana	143	ILQYAQLLTSNNEVDLSYYTSTLF-----SQP-----FS--TF----
Brachypodium distachyon	163	LMRYAQLLQAGGRPSGSGA-----DADAAARQMMMVGAAGSGVMNLPFGPMSPSSTM
Sorghum bicolor	147	LMQYAQLLQGGARGGGSGGDHAEAAAVQQAQQQLMMMGAGGRGGVTLPFSFSP----
Hordeum vulgare	157	LLRYAQLLQAGSA-----GCAVNLFPFGAMSPPSM--
Glycine max	211	LYQYAQILSSG--DEEFPPYTSHLF-----NQQQHQPPS--SQFST-
OsERF101	204	TVSSSSAPQILDFTSQQLIRAGPPSEMPSSGS-----GSATA---AASSTTSASS
Arabidopsis thaliana	174	-SSSSSSSQ---QTQQQLQQQQQREEEKN---Y-----G-----
Brachypodium distachyon	216	TSSSSSPQILDFTTQQLIREGPRPSSPAAAAAMSTGSG---GAATIS---SSTTTASPPP
Sorghum bicolor	202	SSSSSSATQILDFTSQQLIREGGLGPPSPAAAAAMSSSSAAAAAPSPSTTTTAAASSSPS
Hordeum vulgare	187	--SSSSSPHILDFTSQQLIRVSPASEAAATSGSAT---T---GPSI---SSSTTTASSP
Glycine max	249	TFSSSSSPSYFYNQQQYDQGHDSNH-----
OsERF101	251	PGAWPYGGSERKKKDS
Arabidopsis thaliana	204	YNYNYPRE-----
Brachypodium distachyon	270	ASAWPYGAEQHRSNKD--
Sorghum bicolor	262	GSAWPYGGHHNRNKDA--
Hordeum vulgare	235	GAAWPYTGQKNNKDS--

SUPPLEMENTARY FIGURE S1. Amino acid sequence alignment of ERF proteins.

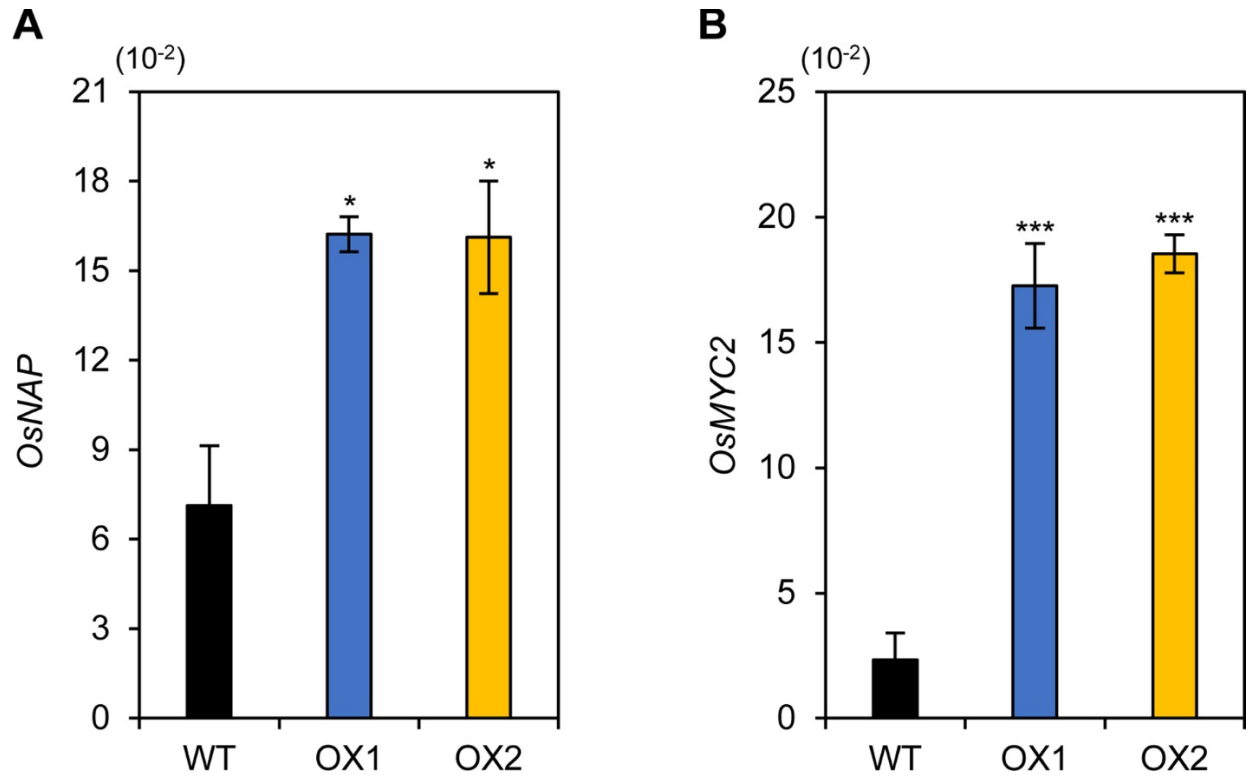
The amino acid sequences of ERF proteins from rice and other plant species homologous to OsERF101 were obtained from NCBI through BLAST analysis. Sequence alignment was performed using ClustalW with default parameters. The sequences are as follows: *Oryza sativa*, XP_015636603.1; *Arabidopsis thaliana*, NP_196837.1; *Brachypodium distachyon*, XP_010239814.1; *Sorghum bicolor*, XP_021318101.1; *Hordeum vulgare*, BAJ99457.1; *Glycine max*, XP_003530479.2; The black boxes represent the conserved AP2/ERF domains of ERF proteins. The red bar represents the AP2/ERF domain of ERF proteins.



SUPPLEMENTARY FIGURE S2. The *oserf101* mutant is less sensitive to MeJA than the WT. Whole leaves of WT and *oserf101* plants grown in paddy soil for three weeks were incubated in 3 mM MES buffer (pH 5.8) containing 50 μ M ABA or 50 μ M MeJA under continuous light conditions at 28°C. Whole leaves floated on 3 mM MES buffer (pH 5.8) without phytohormones were used as a mock control. The pictures were taken at 0, 3, and 4 days after treatment (DT).



SUPPLEMENTARY FIGURE S3. Expression profiles of JA biosynthesis genes in the *oserf101* mutant during DIS. Total RNA was isolated from detached leaves at 0, 2, 3, and 4 days of dark incubation (DDI) as shown in **Figure 3A**. The transcript levels of *OsLOX2* (**A**) and *OsAOS1* (**B**) were determined by qRT-PCR and normalized to that of *OsUBQ5* (Os01g22490). Mean and standard deviations were obtained from three biological repeats. Different letters indicate significantly different values according to a one-way ANOVA and Duncan's least significant range test ($p < 0.05$).



SUPPLEMENTARY FIGURE S4. Expression profiles of *OsNAP* and *OsMYC2* in the *OsERF101*-OX plants (OX1, OX2) grown for one month in the paddy field under NLD conditions. The transcript levels of *OsNAP* (**A**) and *OsMYC2* (**B**) were determined by qRT-PCR and normalized to that of *OsUBQ5* (Os01g22490). Mean and standard deviations were obtained from three biological repeats. Asterisks indicate statistically significant differences between treated samples and the mock control, as determined by Student's *t*-test (* $p < 0.05$, *** $p < 0.001$).

SUPPLEMENTARY TABLE S1. Primers used in this study.

	Forward primers (5'→3')	Reverse primers (5'→3')
A. primers for verification of transgenic plants		
PFG_2D-00368	TTGCATGCATTTTCATCTCC	TTAACTCCCAAATCGTTGCC
pGA2772	TCCGAAACTATCAGTGTCTAGCT	TTGGGGTTTCTACAGGACGTAAC
B. Primers used for gene cloning		
<i>OsERF101</i>	ATGGTCACCGCGCTAGCCCA	TCACGACGACGAATCCTTC
<i>proNAP</i>	AAACTGCAGAAACGTCTCATTCACTAT	AAACCATGGGGCAGTCACCCA
<i>proMYC2</i>	AAACTGCAGGCCCTAACACAAATAC	AAACCATGGGGGTCCTATGGCTAA
C. Primers used for semi-quantitative RT-PCR		
<i>OsERF101</i>	ATGGTCACCGCGCTAGCCCA	TCACGACGACGAATCCTTC
<i>OsUBQ5</i>	ACCACTTCGACCGCCACTACT	ACGCCTAAGCCTGCTGGTT
D. Primers used for ChIP assays		
<i>pOsNAP-a</i>	GGAAACGTCTCATTCACTATTAGGTTT	GGGCATTTTGGAAGTGCAAGAATG
<i>pOsNAP-b</i>	CATTCTTGACACTTCCAAATGCCCTTA	GGTTACAGTGAACAGGAACACAGTTG
<i>pOsNAP-c</i>	CTGTGTTCTCTGTTCACTGTAACCAAATTC	GTGGGTTTGGTCCGTTATCCCT
<i>pOsNAP-d</i>	AAGCGCCGGAAACACAGAAAA	GGCAGTCACCCACACACAACA
<i>pOsMYC2-a</i>	GCCCTAACACAAATACATTAATGT	GATGATGCATACCAAGTTTGTTAC
<i>pOsMYC2-b</i>	GTAACAACTTGGTGTATGCATCATC	AAGAGTTACTCAAGCCTTAAGGCTAT
<i>pOsMYC2-c</i>	ATAGCCTTAAGGCTTGAGTAACTCTT	TAATCAGCCTCCGCTCTCGCTCGAT
<i>pOsMYC2-d</i>	ATCGAGCGAGAGCGGAGGCTGATTA	GTTTGCATGCATTTTGTATATCGTG
<i>pOsMYC2-e</i>	CACGATATACAAAATGCATGCAAC	GGGTCCCATGGCTAATCACTCAA
E. Primers used for RT-qPCR		
<i>OsERF101</i>	TGGTGCCATGTGTCTGAGATT	GCACGGGATGGGGATGATAC
<i>SGR</i>	AGGGGTGGTACAACAAGCTG	GCTCCTTGCGGAAGATGTAG
<i>NYC1</i>	CATGCAACACCAACAAAAGG	GACCATTCCAGGAGAAGCAG
<i>NYC3</i>	TGTCGTTGCCATGTGAAGAT	TTGGTCACGCCACAAATCTA
<i>OsNAP</i>	CAAGAAGCCGAACGGTTC	GTTAGAGTGGAGCAGCAT
<i>OsLOX2</i>	GAGCCCGACTCCGGAACCCCTC	TCGCGTGGCAGTAGATGTTCTCC
<i>OsAOS1</i>	CAATACGTGTACTGGTCAATGG	AAGGTGTCGTACCGGAGGAA
<i>OsMYC2</i>	CAGTGCCACAAGCGGAACCAC	GTTGAGCTGGTCTGCGAGTAG
<i>OsCOI1a</i>	GATGCCCTCCCTGAGATACA	AGTCAGACCTCCTTCCAGCA
<i>OsUBQ5</i>	ACCACTTCGACCGCCACTACT	ACGCCTAAGCCTGCTGGTT