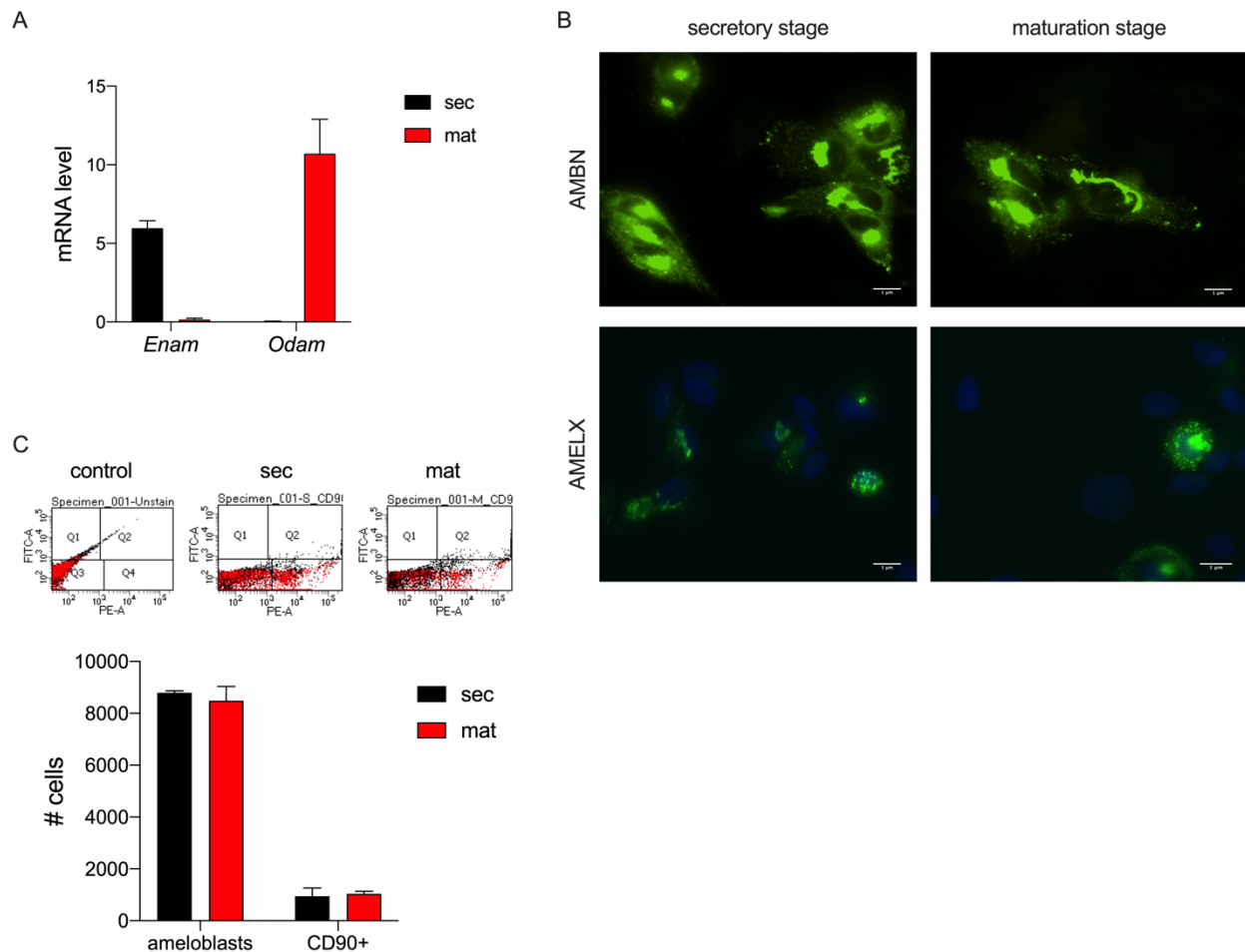
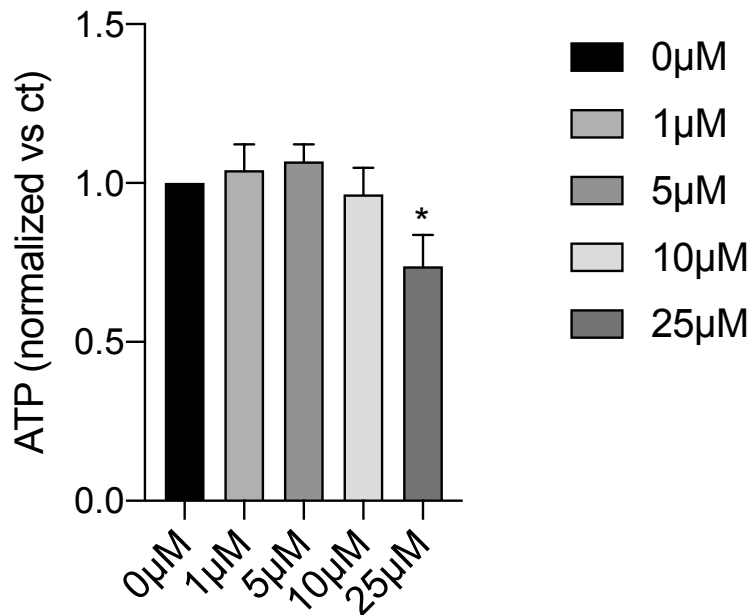


SUPPLEMENTARY FIGURE CAPTIONS

Supplementary Figure 1. Secretory and maturation stage sample purity was assessed upon different techniques. (A) *Enam* (marker for secretory stage) and *Odam* (Odam) (marker for maturation stage) mRNA levels were quantified by RT-qPCR, confirming the purity of isolation of both stages. (B) Immunofluorescence staining was used to verify ameloblastin (Ambn) and amelogenin (AmelX) expression in secretory and maturation cells, which are specific ameloblast proteins (scalebar = 1 μ m). (C) The proportion of positive cells for CD90 fibroblast marker was evaluated by cytofluorimetric analyses: our cell isolation procedure yielded 90% and 92% pure cultures (in secretory and maturation cell isolation respectively). Data represent mean $\pm$ SEM, from a minimum of 3 independent experiments. (* $P < 0.05$, ** $P < 0.005$, *** $P < 0.001$, 2 - tailed unpaired Student's t test).

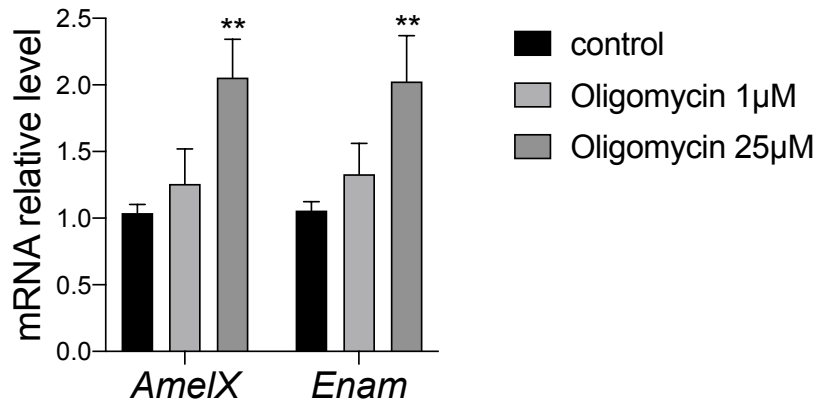


Supplementary Figure 2. Oligomycin decreases ATP levels in LS8 cells at high concentration. ATP content was decreased only when LS8 cells were exposed 15 min to 25 μ M of oligomycin. Data represent mean $\pm$ SEM of 4 independent experiments. (* $P < 0.05$, 2 - tailed unpaired Student's t test).



Supplementary Figure 3. Modulation of ATP by oligomycin alters enamel gene expression.

LS8 cells were exposed for 15 min to oligomycin (1 and 25 μ M) to detect changes in EMPs mRNA level. Only oligomycin (25 μ M) elicited a significant increase in *AmelX* and *Enam* mRNA levels. Data represent mean $\pm$ SEM of 4 independent experiments. (* $P < 0.05$, ** $P < 0.005$, 2 - tailed unpaired Student's t test).



Supplementary Table 1. Primer sequences used for RT-qPCR.

<i>Gene</i>	Forward Sequence	Reverse Sequence	Animal specificity
<i>Gapdh</i>	CTGGAGAAACCTGCCAAGTA	TGTTGCTGTAGCCGTATTCA	mouse
<i>AmelX</i>	GCCGTATCCTTCCTATGGTT	GATGTTTGGCTGATGGTGTT	mouse
<i>Enam</i>	TATGGTCTTCCACCAAGGAA	TAGGCACACCATCTCCAAAT	mouse
<i>Actin β</i>	CACACTGTGCCCATCTATGA	CCGATAGTGATGACCTGACC	rat
<i>Amelx</i>	GCCGTATCCTTCCTATGGTT	GATGTTTGGCTGATGGTGTT	rat
<i>Atp5f1a</i>	TAATGGCAAGCACGCTCTGA	TCAGCTTTGCATCCGACTGT	rat
<i>Cyt C</i>	GGTGTCGCCTCAAACCTATT	CGGGTGAGTCTTCTTGTTTCT	rat
<i>Drp1</i>	GGTGGAATTGGAGATGGTGGTCGA	TTCGTGCAACTGGAAGTGGCACA	rat
<i>Fis1</i>	GTGCCTGGTTTCGAAGCAAATAC	CATAATCCCGCTGCTCCTCTT	rat
<i>Mfn1</i>	ATCTTCGGCCAGTTACTGGAGTT	AGATCATCCTCGGTTGCTATCC	rat
<i>Mfn2</i>	CCTTGAAGACACCCACAGGAATA	CGCTGATTCCCCTGACCTT	rat
<i>Ndufa2</i>	AGGTACTGCGTGAGATTCGC	AAAAGCATAACGGGCCCAGA	rat
<i>Odam</i>	TTGACAGCTTTGTAGGCACA	TGACCTTCTGTTCTGGAAGC	rat
<i>Sdha</i>	AGCCTCAAGTTCGGGAAAGG	CAAGGTAAACCAGCCCCAGT	rat
<i>Uqcrrfs</i>	G TTCCTGCTTCTGTTTCGTTATTC	CTAGCCTCACTGCTCTCTTTAG	rat
mtDNA/nDNA			
<i>Gapdh</i>	GGAAAGACAGGTGTTTTGCA	AGGTCAGAGTGAGCAGGACA	rat
<i>Rnr2</i>	AGCTATTAATGGTTCGTTTGT	AGGAGGCTCCATTTCTCTTGT	rat