

## *Supplementary Material*

### **1 Supplementary Data**

#### Antibodies used for flow cytometry

For surface staining we used, allophycocyanin- or FITC-labeled F4/80; allophycocyanin-labeled anti-CD301 (MGL) and IgG2b mAbs; allophycocyanin-labeled anti-PDL-1 (Biolegend, San Diego, CA, USA) and anti-CD86 (eBioscience, San Diego, CA, USA) mAbs; PE-labeled anti-CD80 (eBioscience) and control hamster IgG (BD Biosciences, San Jose, CA, USA) mAbs; PE-labeled anti-CD206 (Biolegend) and anti-CD124 (BD Biosciences) mAbs. For intracellular staining, we employed the following mAbs: PE-labeled anti-IL-12p35 and control murine IgG1 mAbs; PE-labeled anti-NOS2 (iNOS) and control rat IgG2a; FITC-labeled anti-arginase-1 and control sheep IgG mAbs; allophycocyanin-labeled anti-IL-10 (BD Biosciences) mAb.

### **2 Supplementary Figures and tables**

**Supplementary Figure 1** - Inflammatory macrophages express a M2-like phenotype. Inflammatory macrophages from B6 mice were gated in F4/80<sup>+</sup> cells and analyzed for the expression of CD124 (IL-4R $\alpha$  subunit), arginase-1, IL-12p35, CD206 (MR), and the respective control mAbs. (A) Histograms correspond to CD124<sup>+</sup> macrophages and control IgG2a staining. (B) Plots depict F4/80<sup>+</sup> cells as evaluated for the expression of intracellular arginase-1 and IL-12p35, as well as staining with the control mAbs. (C) Histograms correspond to CD206<sup>+</sup> macrophages and control IgG2a staining, where CD206<sup>high</sup> and CD206<sup>low</sup> subsets were defined. (D) Plots depict CD206<sup>high</sup> and CD206<sup>low</sup> macrophage subsets, which were analyzed for CD206 and intracellular IL-10 expression.

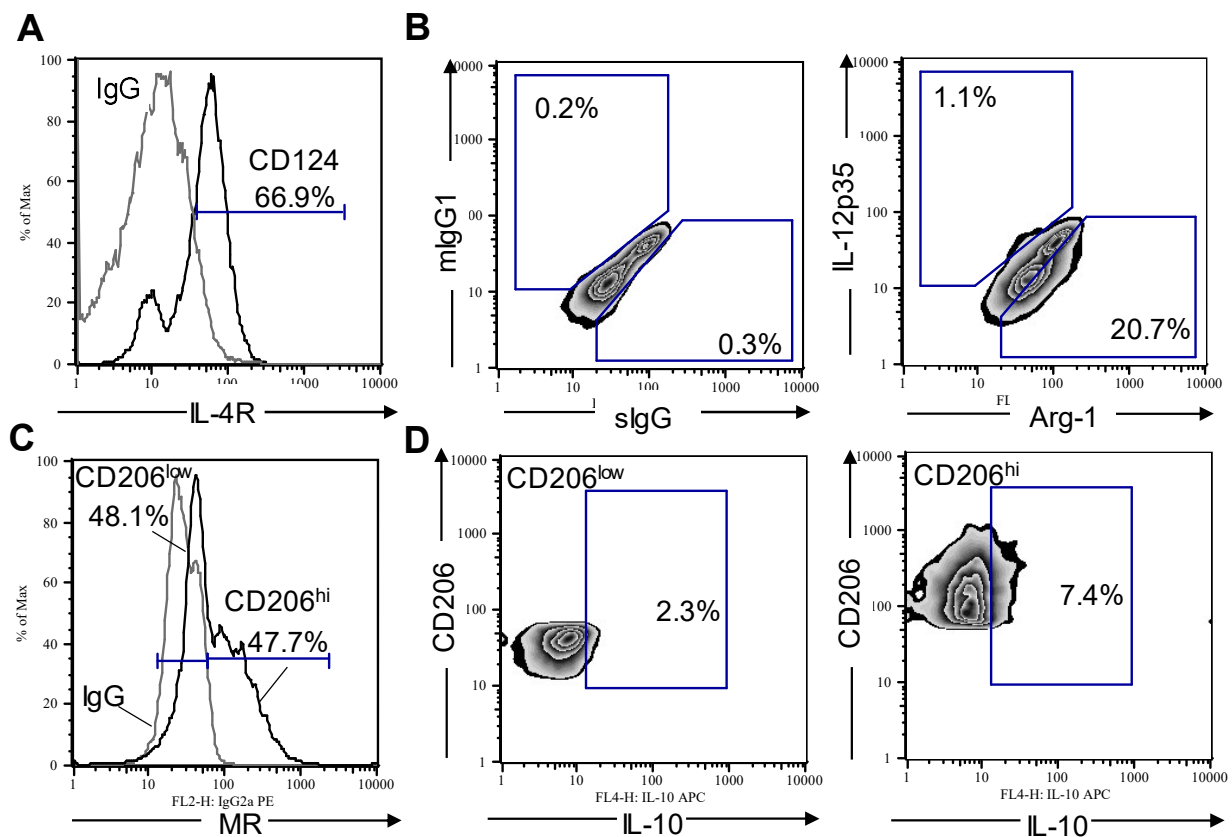
**Supplementary Figure 2** - Inflammatory macrophages express both costimulatory and coinhibitory molecules. Inflammatory (B6) macrophages were cultured in triplicates with medium or RANKL in the presence or absence of IFN- $\gamma$ . After 48 h, cells were harvested and stained with anti-F4/80, anti-CD80 (B7-1), anti-CD86 (B7-2) or anti-PDL-1. (A) F4/80<sup>+</sup> macrophages from unstimulated or RANKL-treated (upper panel only) cultures were analyzed for the expression of F4/80, CD80, CD86, and PDL-1. Staining with hIgG was used for the control of CD80 expression, otherwise, histograms represent unstained cells used for the control of CD86 and PDL-1 expression. (B) F4/80<sup>+</sup> cells were gated and evaluated for the expression of F4/80, CD80, CD86, and PDL-1, as mean (median) of fluorescence intensity (MFI). Results are expressed as means and S.E.M. Data were analyzed by one-way ANOVA followed by Dunnett post-test. Significant differences between unstimulated cultures and those treated with RANKL and/or IFN- $\gamma$  were indicated for P<0.01 (\*\*), P<0.001 (\*\*\*), and P<0.0001 (\*\*\*\*).

**Supplementary Figure 3** - RANKL and IFN- $\gamma$  downregulate the M2-like phenotype. Inflammatory (B6) macrophages were cultured in triplicates with medium or RANKL in the presence or absence of IFN- $\gamma$ . After 48 h, cells were harvested and stained with anti-F4/80, anti-CD206 (MR), anti-CD124 (IL-4R $\alpha$  subunit), and anti-arginase-1, as well as the respective control mAbs. Plots depict F4/80<sup>+</sup>

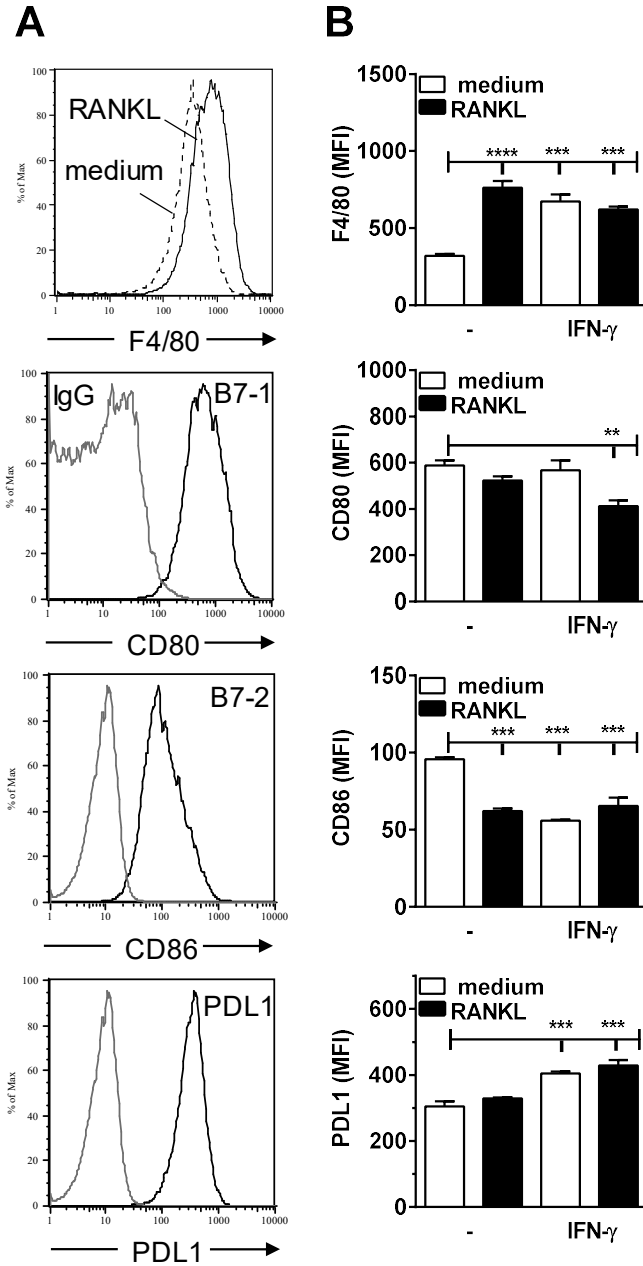
cells as evaluated for the expression of surface CD206 and CD124, as well as intracellular arginase-1. Staining with IgG2a was used for the control of CD206 and CD124 expression, whereas sIgG controlled intracellular arginase-1 expression.

**Supplementary Figure 4** - RANKL and IFN- $\gamma$  skew M2-like into M1 macrophages. Inflammatory (B6) macrophages were cultured in triplicates with medium or RANKL in the presence or absence of IFN- $\gamma$  during 48 h. (A, B) Cells were harvested and stained with anti-F4/80, anti-CD301 (MGL), iNOS or control IgG mAbs. Plots depict F4/80<sup>+</sup> cells as evaluated for the expression of surface CD301 and/or intracellular iNOS. (C) Culture supernatants were assayed for IL-12p70 and CCL17 by ELISA. Results are expressed as means and S.E.M. Data were analyzed by one-way ANOVA followed by Dunnett post-test. Significant differences between unstimulated and treated cultures were indicated for P<0.05 (\*), P<0.01 (\*\*), and P<0.001 (\*\*\*).

## 2.1. Supplementary Figures

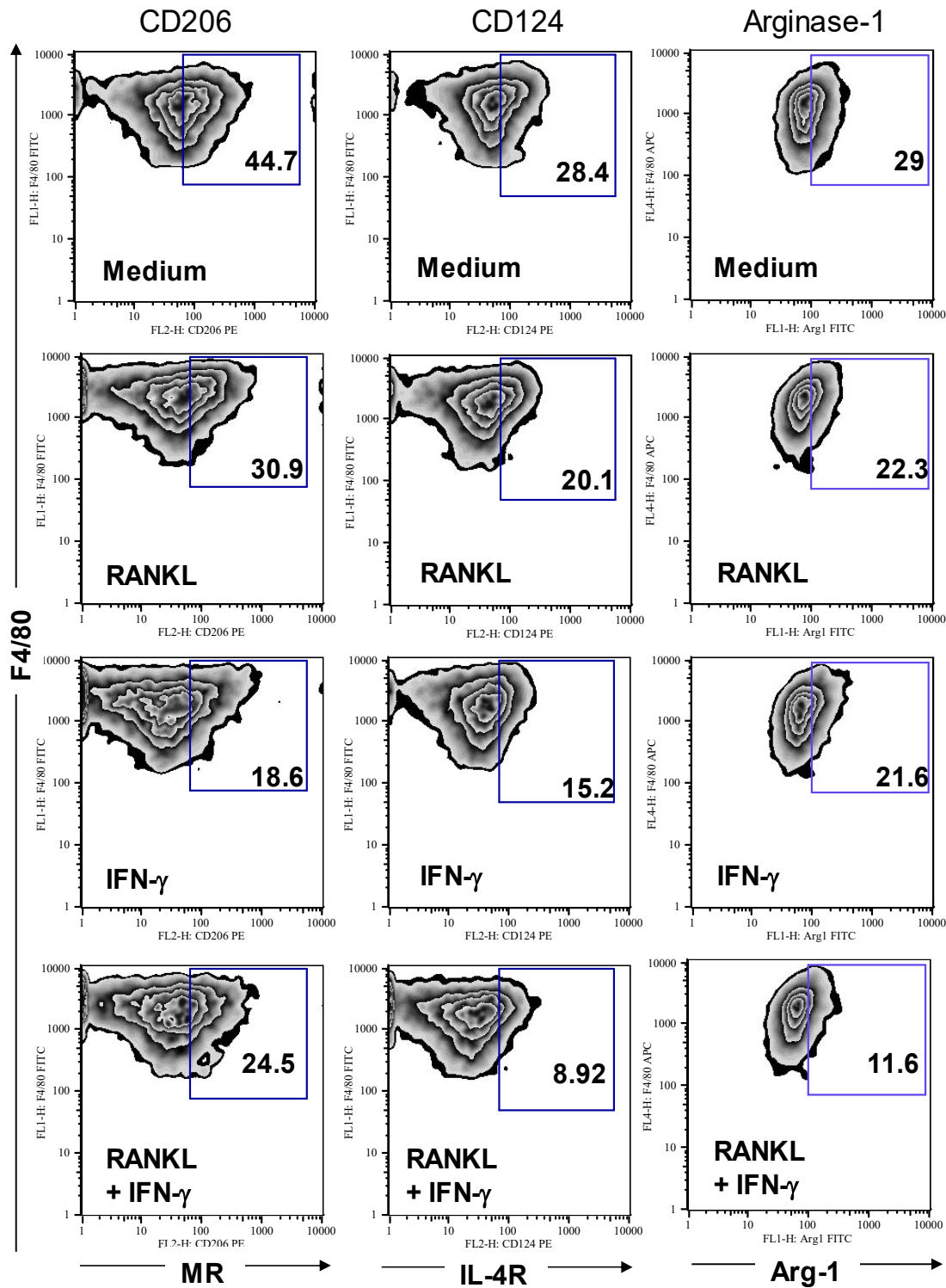


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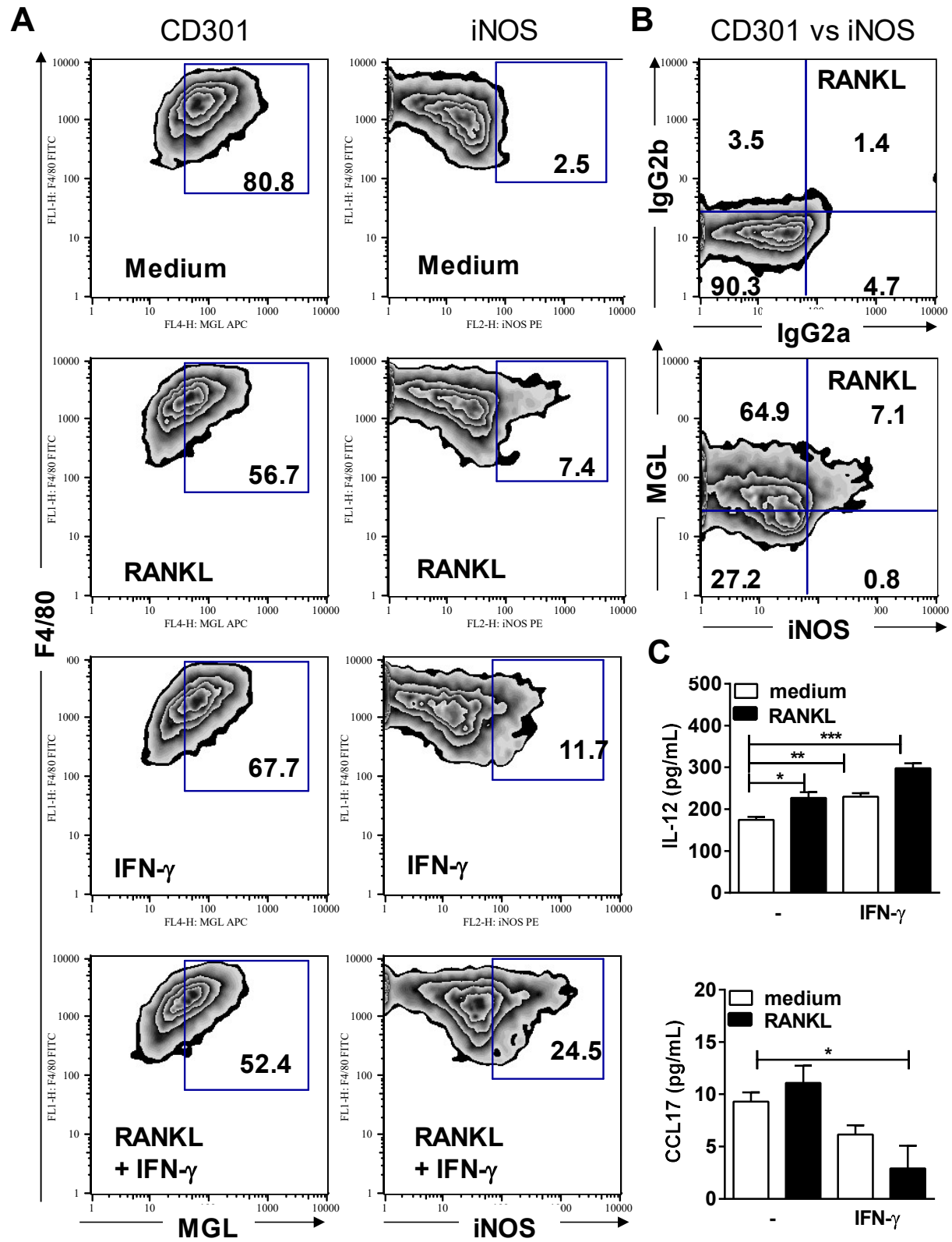


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and those treated with RANKL and/or IFN- $\gamma$  were indicated for  $P < 0.01$  (\*\*),  $P < 0.001$  (\*\*\*), and  $P < 0.0001$  (\*\*\*\*).



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