Supplementary Information

Supplementary Figure Legends

Figure S1. Gene ontology (GO), Kyoto encyclopedia of genes and genomes (KEGG) pathway and Hallmark analysis for genes dysregulated in colorectal tissue samples

(A, C, E) GO (A), KEGG (C) and Hallmark (E) analysis for genes up-regulated in colorectal tumor tissues as shown in Fig. 1D.

(B, D, F) GO (B), KEGG (D) and Hallmark (F) analysis for genes down-regulated in colorectal tumor tissues as shown in Fig. 1D.

Figure S2. Hundreds of lncRNAs were dysregulated in colorectal cancer

(A-D) UCSC genome browser views of RNA-seq as described in Fig. 1A for specific downregulated lncRNAs were shown as indicated.

(E) The expression of lncRNAs as shown in Fig. 2E in a cohort of clinical colorectal tumor (n =

647) and normal (n = 51) samples from TCGA (The Cancer Genome Atlas).

(F) The knockdown efficiency of siRNAs targeting lncRNAs as described in Fig. 2F was examined by RT-qPCR analysis (\pm s.e.m., **P < 0.01, ***P < 0.001).

Figure S3. LUCRC is a lncRNA with three exons and localized in the cytosol of cells

(A) UCSC genome browser view of RNA-seq as described in Fig. 1A for LUCRC was shown. The genomic location and the number of exons of LUCRC were depicted at the bottom.

(B) cDNA sequence of LUCRC (accession number: NR_135175.1).

(C, D) HCT116 cells were subjected to polysome profiling and the resultant fractions were subjected to RNA exaction and RT-qPCR analysis to examine the expression of LUCRC (C) and ACTIN (D). Fractions 1 to 4: free RNA (unbound RNA); Fractions 5 to 6: 40S; Fraction 7: 60S;

Fractions 8 to 10: monosome; Fractions 11 to 20: polysome.

(E) HCT116 cells were subjected to cellular fractionation followed by RNA extraction and RTqPCR analysis to quantify the amount of mRNA as indicated in both nucleus and cytosol of the cells. ACTIN and MALAT1 served as markers for cytosolic and nuclear fraction, respectively.

(F, G) RKO (F) and DLD1 (G) cells were transfected with control siRNA (siCTL) or siRNA specifically targeting LUCRC (siLUCRC) for duration as indicated followed by cell proliferation assay (\pm s.e.m., *P < 0.05, **P < 0.01).

(H) The expression of LUCRC in a cohort of clinical colorectal tumor samples at different stages (stage I, n=111; stage II, n=238; stage III, n=183; stage IV, n=90) and normal samples (n=51) from TCGA.

(I) Kaplan-Meier survival analyses for OS (overall survival) (n = 361) of colorectal cancer patients using LUCRC as input.

Figure S4. LUCRC was required for the expression of genes involved in ER stress response, including BIP

(A) Correlation of the effects of LUCRC on whole transcriptome based on RNA-seq between two biological repeats.

(B) MA plot shows the fold change (FC, siCTL/siLUCRC, log2) against the average of normalized counts for samples as described in Fig. 4A. Red dots represented genes with significant change in response to LUCRC knockdown (q <0.05), and blue line indicated fold change of 1.5.

(C) The expression of BIP in a cohort of clinical colorectal tumor samples at different stages (stage I, n=111; stage II, n=238; stage III, n=183; stage IV, n=90) and normal samples (n=51) from TCGA.
(D) Kaplan-Meier survival analyses for OS (overall survival) (n = 361) of colorectal cancer patients using BIP as input.

Supplementary Table Legend

Table S1. Sequence information for all qPCR primers used in this study. Sequence information of qPCR primers designed to detect the expression of *MYC*, *CCND1*, *FAM83H-AS1*, *MNX1-AS1*, *RNASEH1-AS1*, *LOC105370333*, *OLMALINC*, *SNHG8*, *LUCRC*, *VPS9D1-AS1*, *LOC101927811*, *GAS5*, *MALAT1* and *BIP*. F: forward; R: reverse.

Table S2. Targeting sequence for all siRNAs used in this study. Targeting sequences of siRNAsdesigned to knock down FAM83H-AS1, MNX1-AS1, RNASEH1-AS1, LOC105370333,OLMALINC, SNHG8, LUCRC, VPS9D1-AS1, LOC101927811 or GAS5 were shown.

Table S3. Genes regulated by LUCRC in HCT116 cells as detected by RNA-seq analysis. HCT116 cells transfected with control siRNA (siCTL) and siRNA specifically targeting LUCRC (siLUCRC) for three days were subjected to RNA-seq analysis, and two biological repeats were analyzed. Genes positively- and negatively-regulated by LUCRC were highlighted in red and blue, respectively (Fold change (FC) > 1.5, q<0.05). Α

GO enrichment analysis for up-regulated genes in tumors

rik	osome	biogen	esis						
ce	II divisi	on							
D	VA repli	cation							
m	mitotic cell cycle phase transition								
DNA conformation change									
peptide biosynthetic process									
ribosomal large subunit biogenesis									
D	VA repa	ir							
te	telomere organization								
mitochondrial gene expression									
chromatin organization									
signal transduction by n53 class mediator									
PNA motobolic process									
RNA localization									
ribesemal small suburit biogenesis									
riposomai small subunit biogenesis									
spinole organization									
Dr	DINA replication initiation								
meiotic cell cycle									
cellular amino acid metabolic process									
attachment of spindle microtubules to kinetochore									
_	40	20	20				70		
J	10	20	30	40	50	60	70		
Log10(P)									



KEGG pathway analysis for up-regulated genes in tumors







E2F	TARGETS								
MYC	C TARGETS	V1							
G2N	1 CHECKPC	DINT							
MYC	TARGETS	V2							
MTC	DRC1 SIGN	ALING							
EPITHELIAL MESENCHYMAL TRANSITION									
UNFOLDED PROTEIN RESPONSE									
DNA REPAIR									
GLY	COLYSIS								
ESTROGEN RESPONSE LATE									
UV RESPONSE UP									
ANGIOGENESIS									
CHOLESTEROL HOMEOSTASIS									
WN	WNT BETA CATENIN SIGNALING								
SPE	SPERMATOGENESIS								
P53	PATHWA	,							
0	20	40	60	80					
	Log10(P)								
	- 3(- /								



GO enrichment analysis for down-regulated genes in tumors





D KEGG pathway analysis for down-regulated genes in tumors





Hallmark gene sets analysis for down-regulated genes in tumors





GGCAGCGAGA GGCCCGCGCT GAGGCGGCGC AGCGCAAAGG GGCGGACGCT 50 GAGGCGGGCC AGGGCGCGGC CGGCCGCCGG GAACTCGAGG CGCAGCGTGA 100 CGGCGGCGTG CAGCCCCACG GCCGGGCTGT AGCGCGTGAG CTCCAGGAAC 150 ACAGCGCGGC TCCTGCGCAG AGGGTGCGGG GTCTGGCTGG ACTAAAGGCA 200 AAACTAAAGC CCAGAAGACA GACCAGTGCA CCGGATGCCC GTACCGCGTG 250 ATGGCCAGGA AGGCCCGGCT GTGCAGCTCC TGCTTGATGG CGCTTTGCAG 300 ACGGAGCCAG TGACCACCGA GGCTGTGCCA CTGCATCGGG CCACCATGCT 350 GATATGCCCG GTCCCAGAGC TGCTAGAGAA GAGGTACAGA GGCAGCGAAG 400 ACACGTTGAG GGGGAGGACG AGACCAACTG CGAGACGCCG AGTCCCGGGC 450 TCTCAGGACG CTCTCCCGTA CCTGCGCCCT CGTCAGCCCA TCACCGAAAG 500 AACGGCCTGA CCCAGGGTCA CACTGCACTG AATGCTTCCT GTTTTGCTGT 550

ACTIN



p=0.46

150

100

Months

50

n

Figure S3







D

