## Supplementary Information

## Supplementary Figure Legends

Figure S1. Gene ontology (GO), Kyoto encyclopedia of genes and genomes (KEGG) pathway and Hallmark analysis for genes dysregulated in colorectal tissue samples
(A, C, E) GO (A), KEGG (C) and Hallmark (E) analysis for genes up-regulated in colorectal tumor tissues as shown in Fig. 1D.
(B, D, F) GO (B), KEGG (D) and Hallmark (F) analysis for genes down-regulated in colorectal tumor tissues as shown in Fig. 1D.

Figure S2. Hundreds of IncRNAs were dysregulated in colorectal cancer (A-D) UCSC genome browser views of RNA-seq as described in Fig. 1A for specific downregulated lncRNAs were shown as indicated.
(E) The expression of lncRNAs as shown in Fig. 2E in a cohort of clinical colorectal tumor ( $\mathrm{n}=$ 647) and normal $(\mathrm{n}=51)$ samples from TCGA (The Cancer Genome Atlas).
(F) The knockdown efficiency of siRNAs targeting lncRNAs as described in Fig. 2F was examined by RT-qPCR analysis ( $\pm$ s.e.m., ${ }^{* *} \mathrm{P}<0.01, * * * \mathrm{P}<0.001$ ).

Figure S3. LUCRC is a lncRNA with three exons and localized in the cytosol of cells
(A) UCSC genome browser view of RNA-seq as described in Fig. 1 A for LUCRC was shown. The genomic location and the number of exons of LUCRC were depicted at the bottom.
(B) cDNA sequence of LUCRC (accession number: NR_135175.1).
(C, D) HCT116 cells were subjected to polysome profiling and the resultant fractions were subjected to RNA exaction and RT-qPCR analysis to examine the expression of LUCRC (C) and ACTIN (D). Fractions 1 to 4: free RNA (unbound RNA); Fractions 5 to 6: 40S; Fraction 7: 60S;

Fractions 8 to 10: monosome; Fractions 11 to 20: polysome.
(E) HCT116 cells were subjected to cellular fractionation followed by RNA extraction and RTqPCR analysis to quantify the amount of mRNA as indicated in both nucleus and cytosol of the cells. ACTIN and MALAT1 served as markers for cytosolic and nuclear fraction, respectively.
(F, G) RKO (F) and DLD1 (G) cells were transfected with control siRNA (siCTL) or siRNA specifically targeting LUCRC (siLUCRC) for duration as indicated followed by cell proliferation assay ( $\pm$ s.e.m., ${ }^{*} \mathrm{P}<0.05,{ }^{* *} \mathrm{P}<0.01$ ).
(H) The expression of LUCRC in a cohort of clinical colorectal tumor samples at different stages (stage I, $\mathrm{n}=111$; stage II, $\mathrm{n}=238$; stage III, $\mathrm{n}=183$; stage IV, $\mathrm{n}=90$ ) and normal samples $(\mathrm{n}=51)$ from TCGA.
(I) Kaplan-Meier survival analyses for OS (overall survival) $(\mathrm{n}=361)$ of colorectal cancer patients using LUCRC as input.

Figure S4. LUCRC was required for the expression of genes involved in ER stress response, including BIP
(A) Correlation of the effects of LUCRC on whole transcriptome based on RNA-seq between two biological repeats.
(B) MA plot shows the fold change (FC, siCTL/siLUCRC, $\log 2$ ) against the average of normalized counts for samples as described in Fig. 4A. Red dots represented genes with significant change in response to LUCRC knockdown ( $\mathrm{q}<0.05$ ), and blue line indicated fold change of 1.5 .
(C) The expression of BIP in a cohort of clinical colorectal tumor samples at different stages (stage I, $n=111$; stage II, $n=238$; stage III, $n=183$; stage IV, $n=90$ ) and normal samples ( $n=51$ ) from TCGA. (D) Kaplan-Meier survival analyses for OS (overall survival) $(\mathrm{n}=361)$ of colorectal cancer patients using BIP as input.

## Supplementary Table Legend

Table S1. Sequence information for all qPCR primers used in this study. Sequence information of qPCR primers designed to detect the expression of $M Y C, C C N D 1$, FAM83H-AS1, MNX1-AS1, RNASEH1-AS1, LOC105370333, OLMALINC, SNHG8, LUCRC, VPS9D1-AS1, LOC101927811, GAS5, MALAT1 and BIP. F: forward; R: reverse.

Table S2. Targeting sequence for all siRNAs used in this study. Targeting sequences of siRNAs designed to knock down FAM83H-AS1, MNX1-AS1, RNASEH1-AS1, LOC105370333, OLMALINC, SNHG8, LUCRC, VPS9D1-AS1, LOC101927811 or GAS5 were shown.

Table S3. Genes regulated by LUCRC in HCT116 cells as detected by RNA-seq analysis. HCT116 cells transfected with control siRNA (siCTL) and siRNA specifically targeting LUCRC (siLUCRC) for three days were subjected to RNA-seq analysis, and two biological repeats were analyzed. Genes positively- and negatively-regulated by LUCRC were highlighted in red and blue, respectively (Fold change (FC) $>1.5, \mathrm{q}<0.05$ ).

A
GO enrichment analysis for up-regulated genes in tumors

| ribosome biogenesis <br> cell division <br> DNA replication <br> mitotic cell cycle phase transition <br> DNA conformation change <br> peptide biosynthetic process <br> ribosomal large subunit biogenesis <br> DNA repair <br> telomere organization <br> mitochondrial gene expression <br> chromatin organization <br> signal transduction by p53 class mediator <br> tRNA metabolic process <br> RNA localization <br> ribosomal small subunit biogenesis <br> spindle organization <br> DNA replication initiation <br> meiotic cell cycle <br> cellular amino acid metabolic process <br> attachment of spindle microtubules to kinetochore |
| :--- |

C
KEGG pathway analysis for up-regulated genes in tumors


E
Hallmark gene sets analysis for up-regulated genes in tumors

|  | E2F TARGETS |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | MYC TARGETS V1 |  |  |  |
|  | G2M CHECKPOINT |  |  |  |
|  | MYC TARGETS V2 |  |  |  |
|  | MTORC1 SIGNALING |  |  |  |
|  | EPITHELIAL MESENCHYMAL TRANSITION UNFOLDED PROTEIN RESPONSE |  |  |  |
|  |  |  |  |  |
|  | DNA REPAIR |  |  |  |
|  | GLYCOLYSIS |  |  |  |
|  | ESTROGEN RESPONSE LATE |  |  |  |
|  | UV RESPONSE UP |  |  |  |
|  | ANGIOGENESIS |  |  |  |
|  | CHOLESTEROL HOMEOSTASIS |  |  |  |
|  | WNT BETA CATENIN SIGNALING |  |  |  |
|  | SPERMATOGENESISP53 PATHWAY |  |  |  |
|  |  |  |  |  |

B
GO enrichment analysis for down-regulated genes in tumors

| $\begin{array}{l}\text { lymphocyte activation } \\ \text { inorganic ion homeostasis } \\ \text { cytokine production } \\ \text { activation of immune response } \\ \text { taxis } \\ \text { negative regulation of immune system process } \\ \text { adaptive immune response }\end{array}$ <br> leukocyte migration <br> second-messenger-mediated signaling <br> cytokine-mediated signaling pathway <br> actin filament-based process <br> myeloid leukocyte activation <br> positive regulation of defense response <br> response to bacterium <br> regulation of seretion <br> inositol lipid-mediated signaling <br> regulation of ion transport <br> peptidyl-tyrosine phosphorylation <br> interferon-gamma-mediated signaling pathway <br> lymphocyte migration |
| :--- |
| $\begin{array}{l}\text { 10 20 } \quad \text { Log10(P) } \\ \text { 10 }\end{array}$ |

D
KEGG pathway analysis for down-regulated genes in tumors


F
Hallmark gene sets analysis for down-regulated genes in tumors




C


D





