

## Ca<sub>v</sub>1.3 L-type calcium channels increase vulnerability of substantia nigra dopaminergic neurons in MPTP mouse model of Parkinson's disease

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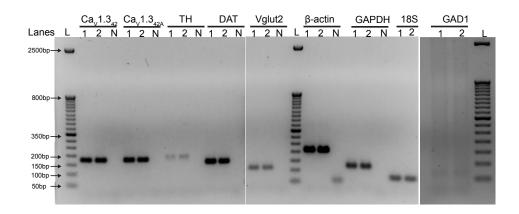
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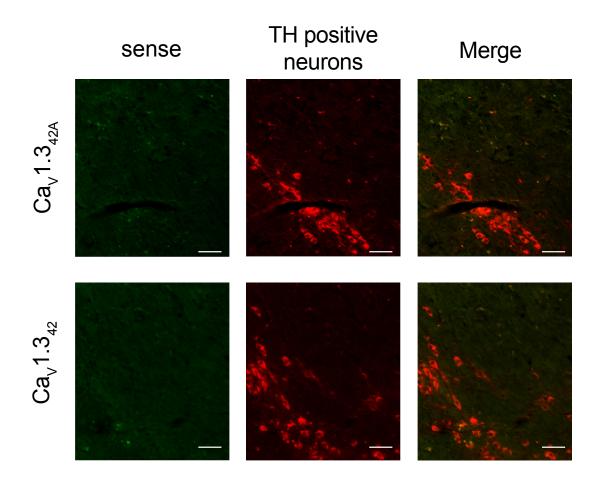
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## Supplementary Material



# Figure S1: Primer specificity for all the mouse qRT-PCR primers using agarose gel electrophoresis.

Single bands were observed at the desired lengths for the mouse qRT-PCR primers, viz.  $Ca_V 1.3_{42}$ ,  $Ca_V 1.3_{42A}$ , TH, DAT, Vglut2,  $\beta$ -actin, GAPDH, 18S and GAD1 using mouse cDNA derived from control tissue. Two SNpc cDNA samples (1 and 2) and a no template negative control (N) was run for each set of qRT-PCR primers. The qRT-PCR products were run on a 2% agarose gel at 100mV. Ladder used was Invitrogen TrackIt<sup>TM</sup> 50bp DNA ladder (ThermoFisher Scientific).



# Figure S2: Absence of signal in sections treated with sense *in situ* hybridization probes.

The use of sense probes against  $Ca_V 1.3_{42A}$  and  $Ca_V 1.3_{42}$  did not give fluorescent signal within TH positive neurons in the SNpc using fluorescence *in situ* hybridization and TH immunohistochemistry, respectively. Images were acquired using Zeiss Axio imager M2 fluorescence microscope with a 20X/0.8 objective. Scale bar = 50µm.

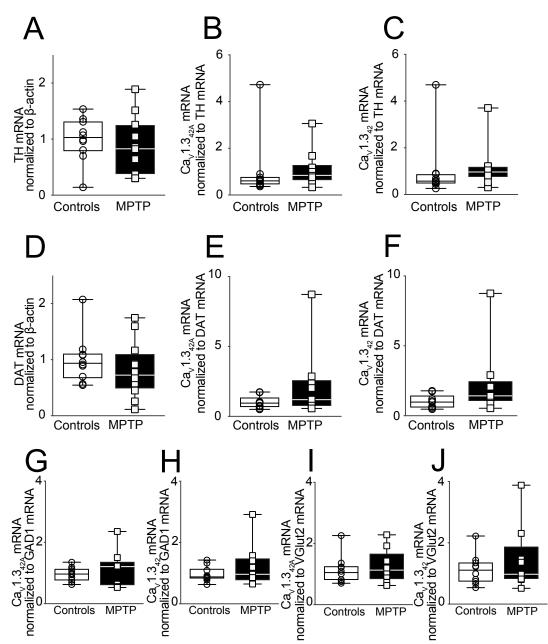


Figure S3: Ca<sub>V</sub>1.3<sub>42A</sub> and Ca<sub>V</sub>1.3<sub>42</sub> mRNA levels in the VTA of MPTP-treated mice.

(A) TH mRNA expression in the VTA of mice treated subcutaneously with MPTP (30mg/kg body weight) for 14 days. MPTP treatment did not lead to significant reduction of TH mRNA levels in the VTA (p=0.6025, t=0.5296, df=19). qRT-PCR data were normalized to mRNA levels of  $\beta$ -actin. (B) Relative mRNA levels for Ca<sub>V</sub>1.3<sub>42A</sub> (Mann–Whitney U = 30, n<sub>1</sub> = 10, n<sub>2</sub> = 10, p=0.1431, two-tailed) and (C) Ca<sub>V</sub>1.3<sub>42</sub> (Mann–Whitney U = 28, n<sub>1</sub> = 10, n<sub>2</sub> = 10, p=0.1051, two-tailed) were found to be unchanged in the SNpc in response to MPTP treatment when the mRNA signal was normalized to TH. (D) mRNA levels of DAT were unchanged upon MPTP treatment when normalized to  $\beta$ -actin mRNA levels (Mann–Whitney U = 45, n<sub>1</sub> = 10, n<sub>2</sub> = 12, p=0.3463, two-tailed). (E) Relative mRNA levels for Ca<sub>V</sub>1.3<sub>42</sub> (Mann–Whitney U = 24, n<sub>1</sub> = 9, n<sub>2</sub> = 9, p=0.1615, two-tailed) and (F) Ca<sub>V</sub>1.3<sub>42</sub> (Mann–Whitney U = 22, n<sub>1</sub> = 9, n<sub>2</sub> = 9, p=0.1135, two-tailed) remained unaltered in the VTA in response to MPTP

Ca<sub>V</sub>1.3<sub>42A</sub> (Mann–Whitney U = 33,  $n_1 = 10$ ,  $n_2 = 9$ , p=0.3562, two-tailed) and **(H)** Ca<sub>V</sub>1.3<sub>42</sub> (Mann–Whitney U = 35,  $n_1 = 10$ ,  $n_2 = 9$ , p=0.4470, two-tailed) were found to be unchanged in the VTA of MPTP-treated mice upon normalization to GAD1. **(I)** mRNA levels for Ca<sub>V</sub>1.3<sub>42A</sub> (Mann–Whitney U = 40,  $n_1 = 10$ ,  $n_2 = 9$ , p=0.7197, two-tailed)and **(J)** Ca<sub>V</sub>1.3<sub>42</sub> (Mann–Whitney U = 38,  $n_1 = 10$ ,  $n_2 = 9$ , p=0.6038, two-tailed) were unchanged in the VTA of MPTP-treated mice upon normalization to Vglut2. Each point in the scatter represents an individual animal. For controls, n=10, for MPTP treated animals, n=9-10. Unpaired, two-tailed Student's t-test or two-tailed Mann Whitney U was performed for each of the pair-wise comparison. Data represented as box-whiskers plots where each box represents quartiles with the line indicating median. Whiskers show the absolute range.

#### Table S1: Sequences for mouse qRT-PCR primers

Gene	Primer	Sequence
Ca <sub>V</sub> 1.3 <sub>42</sub>	Forward primer	GTCCCTCCAGCTGGTGATGATGA
(NM_028981.2)		
	Reverse	GGCCCAATGTCATGCAGGGT
	primer	
Ca <sub>V</sub> 1.3 <sub>42A</sub>	Forward	GTCCCTCCAGCTGGTGATGATGA
	primer	
	Reverse	CAGGCAGAGAACTCTAAAGCATCCG
	primer	
β -actin	Forward	CCTTCTTGGGTATGGAATCCTGTGGC
(NM_007393.5)	primer	

	Reverse primer	GCGCTCAGGAGGAGCAATGATCTTG
18S rRNA	Forward primer	GAGGGAGCCTGAGAAACGG
	Reverse primer	GTCGGGAGTGGGTAATTTGC
GAPDH (NM_001289726.1)	Forward primer	GGCCTTCCGTGTTCCTAC
	Reverse primer	TGTCATCATACTTGGCAGGTT
TH (NM_009377.1)	Forward primer	GGAACGGTACTGTGGCTACC
	Reverse primer	GAGTGCATAGGTGAGGAGGC
DAT (NM_010020.3)	Forward primer	GGTTCTACGGTGTCCAGCAA

	Reverse primer	TAGTGTGGGGGTCTGAAGGT
Vglut2(NM_080853.3)	Forward primer	GCGGAGGCAAAGTTATCAAG
	Reverse primer	CCTGGAATCTGGGTGATGAT
GAD1(NM_008077.5)	Forward primer	GGGTGGTGGACTGCTCATGT
	Reverse primer	AATTGGCCCTTTCTATGCCG

### Table S2: qRT-PCR conditions

PCR step	Temperature	Time	Cycles
Initial Denaturation	95ºC	10min	1
Denaturation	95ºC	20s	40
Annealing	60ºC	30s	40

### Supplementary Material

Elongation	72ºC	40s	40