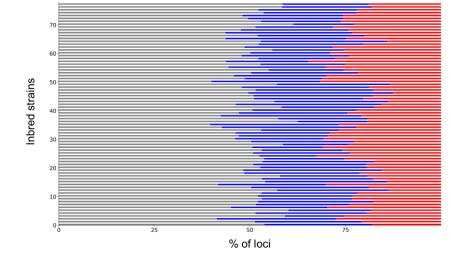
Figure S1, Sampaio et al.



MATα-ACE2 Haploids

		MAId-ACE2 Hapioids												
		T1	T2	T3	T4	T5	<b>T6</b>	<i>T</i> 7	T8	T9	T10	T11	T12	T13
	JAY#	1520	1525	1526	1531	1536	1539	1545	1546	1553	1557	290	293	328
T1	1521		1598	1600	1602	1604	1606	1608	1610	1612	1614	1616	1618	162
T2	1522			1622	1624	1626	1628	1630	1632	1634	1636	1638	1640	164
тз	1529				1644	1646	1648	1650	1652	1654	1656	1658	1660	166
T4	1533					1664	1666	1668	1670	1672	1674	1676	1678	168
Т5	1535						1682	1684	1686	1688	1690	1692	1694	169
<b>T</b> 6	1540							1698	1700	1702	1704	1706	1708	171
77	1543								1712	1714	1716	1718	1720	172
Т8	1549									1724	1726	1728	1730	173
Т9	1552										1734	1736	1738	174
T10	1554											1742	1744	174
T11	291												1748	175
T12	295													175
T13	326													

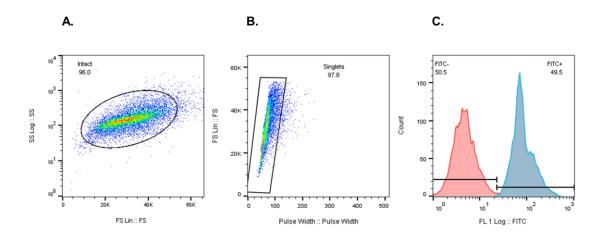


C.	78 strains in	Geno	Genotype Frequency (%)				
	inbred collection	M/P	P/P	M/M			
	Average	51.02	26.11	22.87			
	Median	51.49	25.98	22.68			
	Maximum	62.52	37.95	34.65			
	Minimum	39.64	14.72	12.57			

В.

# Incubation Temperature 30C 39C 0 0 0 0 000 000

Phenotype classification	Representative growth at 39C	Representative growth at 30C
Score 1 (JAY1664) Very small colonies, at the lower limit of unaided visualization.		
Score 2 (JAY1628) Small, yet easily visible, colonies.		
Score 3 (JAY270) Heterogeneous colony size, ranging from medium to small sizes.	$\bigcirc$	
Score 4 (JAY1658) Medium to large colonies, with most having medium size. Narrow variation in colony size.	(10) (10) (10) (10) (10) (10) (10) (10)	
Score 5 (JAY1610) Large colonies. Homogenous size.		





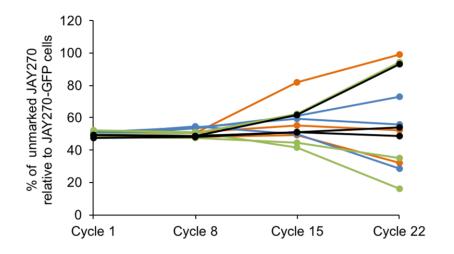
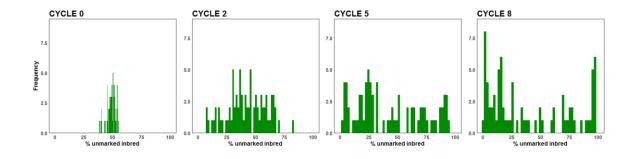


Figure S6, Sampaio et al.



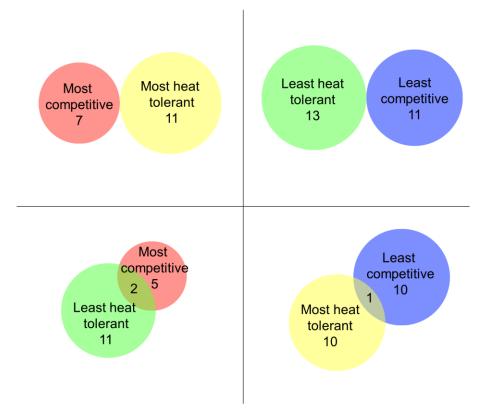
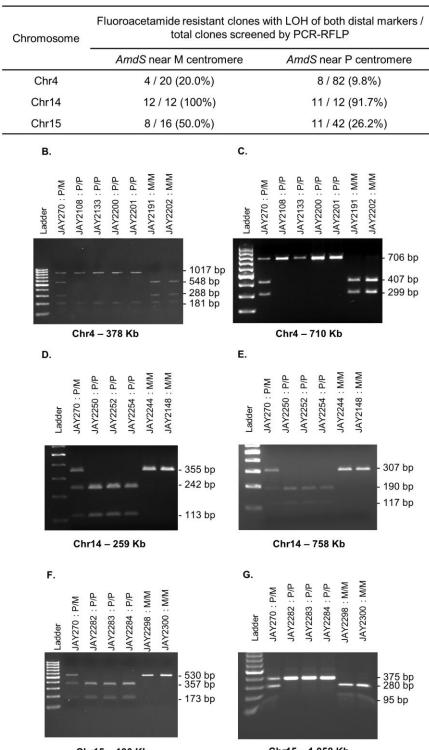


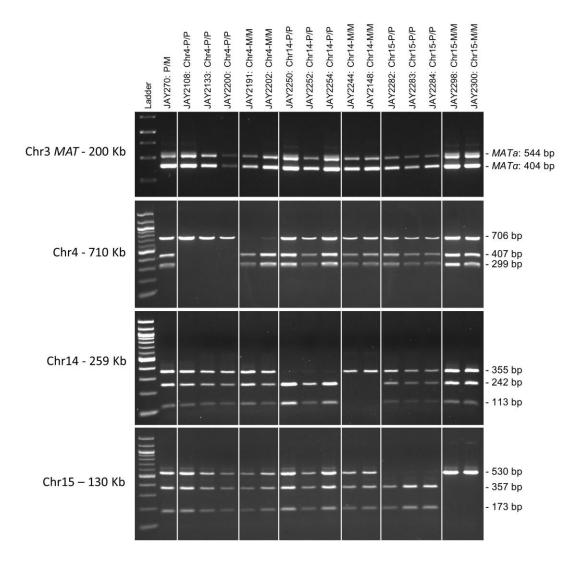
Figure S8, Sampaio et al.

A.



Chr15 - 130 Kb

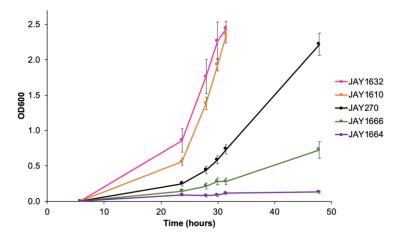
Chr15 - 1,058 Kb



		Heat tolerance phenotype scores (based on colony size)							
Strain	Description	Consensus	Ind	Independent biological replicates					
		score	А	В	С	D	E		
JAY1632	Heat tolerant inbred	5	4	5	5	5	4		
JAY1610	Heat tolerant inbred	5	5	4	5	5	4		
JAY270	Parental strain	3	3	2	3	-	_		
JAY1666	Heat sensitive inbred	1	2	2	1	1	1		
JAY1664	Heat sensitive inbred	1	1	1	1	1	1		

## A. Colony formation in agar media at 39C heat tolerance scores

B. Liquid media 39C growth curves



## Supplemental Figures, Sampaio et al.

**Figure S1.** Draft map of heterozygosity in the JAY270 genome (Rodrigues-Prause et al., 2018), showing the distribution of 12,023 phased heterozygous single nucleotide polymorphisms (HetSNPs; File S1; double colored markers, arbitrarily defined as maternal (red) or paternal (blue) haplotypes). Each horizontal line represents a *S. cerevisiae* chromosome. Black circles indicate the positions of the centromeres. Plots were generated to scale in Python 2.7 using the matplotlib package and a custom script. For size reference, Chr1 is 230 Kb. Note that Chr1 is fully homozygous.

Figure S2. Inter-tetrad matings and genotype distribution in the inbred collection.

A) Schematic representation of the matings between recombinant haploids. Two haploid spores from each of 13 different tetrads (T1 – T13) were selected to generate a collection of diploid inbred strains resulting from intertetrad matings. The JAY number (*e.g.*, JAY1520, JAY1598, etc.) corresponding to the strain identifier is displayed for the parent haploids and for the inbreds, and matches the entries in Table S1.

B) The inbreds were expected to contain approximately 50% of the original heterozygosity present in the parental JAY270 genome, and be homozygous for 25% of the paternal alleles and 25% of the maternal alleles. The genotype classes in the inbred collection from panel A and Figure 1B are plotted as cumulative frequencies for all HetSNP markers. Heterozygous M/P (gray); homozygous P/P (blue); homozygous M/M (red). C) Summary of the frequency of genotype classes in the inbred collection.

**Figure S3.** Variation in tolerance to heat stress in the inbred collection. A plate spotting assay was used for phenotypic screening. The column to the left shows uniform growth among strains incubated for 24 hours at 30C. The column to the right shows variable growth at 39C after four days of incubation. Black circles highlight the control JAY270, which displays an intermediate level of heat tolerance. Rectangles highlight the growth of representative strains shown in Figures S4 and S8 with different levels of heat tolerance, from high to low: JAY1632 (pink), JAY1610 (orange), JAY1658 (yellow), JAY1628 (turquoise), JAY1666 (green), and JAY1664 (purple).

**Figure S4.** Inbred strains show phenotypic variation in tolerance to heat stress. A scoring system was used to assess the heat stress tolerance phenotype of each inbred strain. The criteria for each score used and the strains chosen as representative examples for each phenotypic category are described next to images of growth on Petri dishes. Incubation at 39C for 4 days and incubations at 30C for 2 days are depicted.

## Figure S5. Flow cytometry analysis.

A) False events corresponding to cell debris or other material were eliminated by gating intact cells in a side scatter by forward scatter plot.

B) Potential cell aggregates were subsequently removed from counting by gating single cells in a forward scatter by pulse width plot.

C) Final counts of GFP+ (JAY270 parent strain) and GFP- cells (inbred diploid strains) in each co-culture were determine by gating the FITC+ and FITC- population in a count by FITC signal plot.

D) Co-culture competition assays between four independently-generated GFP-marked JAY270 derivatives and the unmarked JAY270 parent over 22 passage cycles (24 hours per cycle). Each of the four GFP-marked controls (shown in orange, blue, green and black curves) were co-cultured with the original JAY270 parent in triplicates. A neutral and stable ratio of GFP- to GFP-tagged strains was observed in all cases through cycle 8 of competition. Beyond that point the ratios tended to diverge up or down between triplicates, presumably as a result of emergence of beneficial *de novo* mutations either in the GFP- or GFP+ competitors. These results indicated that the GFP marker itself did not confer an advantage or a burden to the cells carrying it, and that any departure from the starting 1:1 ratio detected before cycle 8 could be confidently attributed to the genotypes present in the starting inoculum, and not due to *de novo* mutations arising during the co-culture.

**Figure S6.** Distributions of the competitive growth phenotypes in the inbred strain collection in cumulative coculture over time. Histograms showing the change over time of the distributions of phenotypes in the competitive growth assay. By Cycle 8 the cumulative nature of this phenotype can be seen by the higher numbers of strains in the highest and lowest phenotypes.

**Figure S7.** Venn diagrams displaying pairwise overlap comparisons of inbred strains that displayed extreme phenotypes in heat tolerance and competitive growth (at Cycle 5). Circles represent the groups of inbred strains that were most heat tolerant (yellow) and most competitive (red); and the groups that were least heat tolerant (green) and least competitive (purple). Diagrams were created using BioVenn web application (Hulsen et al., 2008)

#### Figure S8. Validation of targeted UPD strain constructions.

A) Table showing the frequency of concomitant loss of both (left and right arm) distal PCR-RFLP markers physically linked to the insertion of *AmdS* adjacent to the respective centromere (Figure 5A). A subset of the *AmdS* clones remained heterozygosis at one or both distal PCR-RFLP markers, suggesting that they were likely derived from mitotic recombination instead of the desired chromosome loss. All of the chromosome loss clones with bilateral LOH analyzed by tetrad dissection displayed full (4) spore viability, indicating that spontaneous endoduplication leading to UPD occurred very quickly after selection of the monosomic intermediates. B-G) Agarose gel electrophoresis of PCR-RFLP genotyping of HetSNP markers. Panels B, D and F show genotypes of RFLP markers located at the left arms of Chr4, Chr14 and Chr15, respectively. Panels C, E and G show genotypes of RFLP markers located at the right arms of Chr4, Chr14 and Chr15, respectively. The approximate coordinates of each RFLP marker are indicated below each image. The approximate positions of the relevant centromeres where *AmdS* was inserted are: *CEN4* – 450 Kb; *CEN14* – 629 Kb; *CEN15* – 327 Kb. The

corresponding genotype at each position is indicated for each UPD strain as P/P for homozygous paternal, M/M for homozygous maternal and P/M for heterozygous.

**Figure S9.** Validation of retention of heterozygosity at non-targeted loci in the UPD strains. PCR-RFLP genotyping of three non-targeted and one targeted HetSNP markers was performed for JAY270 and each of the 15 independently generated UPD strains constructed for this study. Panels show agarose gels used for genotyping of PCR-RFLP markers located at the Chr3 *MAT* locus (not targeted in any of the UPD strains), and at each of the targeted chromosomes. Note that all UPD strains remained heterozygous at the non-targeted chromosomes but were homozygous for the specific allele targeted. Vertical lines were used to facilitate the visualization of the groups of strains with the same targeted UPDs.

Figure S10. Comparison of the two assays used to characterize heat stress tolerance.

A) Representative results from colony size (agar media; 4 days incubation) scoring system described in Figure S4 and used in Figure 3A.

B) Pure culture (liquid media) growth curves and OD600 measurements over a period of 24-48 hours. Error bars shown in SD.

Strains used in the two heat tolerance assays are color coded similarly in A and B and Figure S3.

# **Table S1.** Yeast strains used in this study.

Strain	Mating type	Relevant genotype	Description	Source
Haploids:		0 11		
JAY1520	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1521	MATa	ace2-A7		
JAY1525	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1522	MATa	ace2-A7		
JAY1526	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1529	MATa	ace2-A7		
JAY1531	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1533	MATa	ace2-A7		
JAY1536	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1535	MATa	ace2-A7		
JAY1539	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1540	MATa	ace2-A7		
JAY1545	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1543	MATa	ace2-A7		
JAY1546	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1549	MATa	ace2-A7		
JAY1553	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1552	MATa	ace2-A7		_
JAY1557	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	(Rodrigues-Prause et al., 2018)
JAY1554	MATa	ace2-A7		
JAY290	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	Our strain collection
JAY291	MATa	ace2-A7		
JAY293	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	Our strain collection
JAY295	MATa	ace2-A7		
JAY328	ΜΑΤα	ACE2	Sibling spores from a JAY270 tetrad	Our strain collection
JAY326	MATa	ace2-A7		
Diploids:				
JAY270	MATa/MATa	ACE2/ace2-A7	Sugarcane bioethanol fermentation strain	Argueso et al., 2009
JAY2208 (JAY270-	MATa/MATa	ACE2/ace2-A7; CEN5::GFP-	Hemizygous GFP-KanMX insertion near Chr5	This study
GFP)		KanMX		
JAY1598	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1525	This study
JAY1600	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1526	This study
JAY1602	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1531	This study
JAY1604	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1536	This study
JAY1606	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1539	This study
JAY1608	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1545	This study
JAY1610	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1546	This study
JAY1612	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1553	This study
JAY1614	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY1557	This study
JAY1616	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY290	This study
JAY1618	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY293	This study

# Table S1 (continued). Yeast strains used in this study.

Strain	Mating type	Relevant genotype	Description	Source
JAY1620	ΜΑΤα/ΜΑΤα	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1521 x JAY328	This study
JAY1622	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1526	This study
JAY1624	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1531	This study
JAY1626	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1536	This study
JAY1628	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1539	This study
JAY1630	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1545	This study
JAY1632	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1546	This study
JAY1634	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1553	This study
JAY1636	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY1557	This study
JAY1638	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY290	This study
JAY1640	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY293	This study
JAY1642	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1522 x JAY328	This study
JAY1644	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1531	This study
JAY1646	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1536	This study
JAY1648	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1539	This study
JAY1650	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1545	This study
JAY1652	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1546	This study
JAY1654	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1553	This study
JAY1656	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY1557	This study
JAY1658	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY290	This study
JAY1660	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY293	This study
JAY1662	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1529 x JAY328	This study
JAY1664	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY1536	This study
JAY1666	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY1539	This study
JAY1668	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY1545	This study
JAY1670	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY1546	This study
JAY1672	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY1553	This study
JAY1674	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY1557	This study
JAY1676	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY290	This study
JAY1678	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY293	This study
JAY1680	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1533 x JAY328	This study
JAY1682	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY1539	This study
JAY1684	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY1545	This study
JAY1686	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY1546	This study
JAY1688	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY1553	This study
JAY1690	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY1557	This study
JAY1692	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY290	This study
JAY1694	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY293	This study
JAY1696	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1535 x JAY328	This study
JAY1698	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY1545	This study
JAY1700	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY1546	This study
JAY1702	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY1553	This study
JAY1704	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY1557	This study

# Table S1 (continued). Yeast strains used in this study.

Strain	Mating type	Relevant genotype	Description	Source
JAY1706	ΜΑΤα/ΜΑΤα	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY290	This study
JAY1708	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY293	This study
JAY1710	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1540 x JAY328	This study
JAY1712	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1543 x JAY1546	This study
JAY1714	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1543 x JAY1553	This study
JAY1716	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1543 x JAY1557	This study
JAY1718	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1543 x JAY290	This study
JAY1720	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1543 x JAY293	This study
JAY1722	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1543 x JAY328	This study
JAY1724	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1549 x JAY1553	This study
JAY1726	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1549 x JAY1557	This study
JAY1728	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1549 x JAY290	This study
JAY1730	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1549 x JAY293	This study
JAY1732	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1549 x JAY328	This study
JAY1734	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1552 x JAY1557	This study
JAY1736	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1552 x JAY290	This study
JAY1738	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1552 x JAY293	This study
JAY1740	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1552 x JAY328	This study
JAY1742	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1554 x JAY290	This study
JAY1744	MATa/ MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1554 x JAY293	This study
JAY1746	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY1554 x JAY328	This study
JAY1748	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY291 x JAY293	This study
JAY1750	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY291 x JAY328	This study
JAY1752	MATa/MATa	ACE2/ace2-A7	Inbred diploid resulting from crossing between JAY295 x JAY328	This study
JAY2108	MATa/MATa	ACE2/ace2-A7; Chr4-UPD P/P	JAY270 derivative containing uniparental disomy for Chr4 (P/P)	This study
JAY2133	MATa/MATa	ACE2/ace2-A7; Chr4-UPD P/P	JAY270 derivative containing uniparental disomy for Chr4 (P/P)	This study
JAY2191	MATa/MATa	ACE2/ace2-A7; Chr4-UPD M/M	JAY270 derivative containing uniparental disomy for Chr4 (P/P)	This study
JAY2200	MATa/MATa	ACE2/ace2-A7; Chr4-UPD P/P	JAY270 derivative containing uniparental disomy for Chr4 (P/P)	This study
JAY2201	MATa/MATa	ACE2/ace2-A7; Chr4-UPD M/M	JAY270 derivative containing uniparental disomy for Chr4 (M/M)	This study
JAY2202	MATa/MATa	ACE2/ace2-A7; Chr4-UPD M/M	JAY270 derivative containing uniparental disomy for Chr4 (M/M)	This study
JAY2250	MATa/MATa	ACE2/ace2-A7; Chr14-UPD P/P	JAY270 derivative containing uniparental disomy for Chr14 (P/P)	This study
JAY2252	MATa/MATa	ACE2/ace2-A7; Chr14-UPD P/P	JAY270 derivative containing uniparental disomy for Chr14 (P/P)	This study
JAY2254	MATa/MATa	ACE2/ace2-A7; Chr14-UPD P/P	JAY270 derivative containing uniparental disomy for Chr14 (P/P)	This study
JAY2244	MATa/MATa	ACE2/ace2-A7; Chr14-UPD M/M	JAY270 derivative containing uniparental disomy for Chr14 (M/M)	This study
JAY2248	MATa/MATa	ACE2/ace2-A7; Chr14-UPD M/M	JAY270 derivative containing uniparental disomy for Chr14 (M/M)	This study
JAY2282	MATa/MATa	ACE2/ace2-A7; Chr15-UPD P/P	JAY270 derivative containing uniparental disomy for Chr15 (P/P)	This study
JAY2283	MATa/MATa	ACE2/ace2-A7; Chr15-UPD P/P	JAY270 derivative containing uniparental disomy for Chr15 (P/P)	This study
JAY2284	MATa/MATa	ACE2/ace2-A7; Chr15-UPD P/P	JAY270 derivative containing uniparental disomy for Chr15 (P/P)	This study
JAY2298	MATa/MATa	ACE2/ace2-A7; Chr15-UPD M/M	JAY270 derivative containing uniparental disomy for Chr15 (M/M)	This study
JAY2300	MATa/MATa	ACE2/ace2-A7; Chr15-UPD M/M	JAY270 derivative containing uniparental disomy for Chr15 (M/M)	This study

 Table S2. Oligonucleotides used in this study.

Name	5' – 3' sequence <sup>a</sup>	Description
JAO14	AGGAGGGTATTCTGGGCCTCCATG	Inside MX4 region. To confirm insertion of MX4 cassettes; Rev.
JAO15	ATGCGAAGTTAAGTGCGCAGAAAG	Inside MX4 region. To confirm insertion of MX4 cassettes; Fwd.
JAO1385	GATTACACCGTACTTCTTTTCAATGCGTAAACAACTAGAGTTGACAAA CTGCTGTCGATTCGAT	To amplify GFP cassette; Fwd. Homology tail to CEN5 region.
JAO1386	CACGTCAAGACTGTCAAGGAGGGTATTCTGGGCCTCCATGTC GTCGATGAATTCGAGCTC	To amplify GFP cassette; Rev. Homology tail to KanMX4 cassette.
JAO1387	GACATGGAGGCCCAGAATAC	To amplify <i>KanMX4 cassette</i> ; Fwd.
JAO466	TACGAAGTGCTAGGAGGTATATTATTAATATGATAGTTTTTAGTTTATAATA	To amplify <i>KanMX4</i> cassette; Rev. Homology tail to <i>CEN5</i> region.
JAO464	CATCGTGTAGTCAAGCAGCA	To confirm integration of GFP-KanMX4 cassette; Fwd. CEN5 region.
JAO467	CATTTACAGATTCATAGTTC	To confirm integration of GFP-KanMX4 cassette; Rev. CEN5 region.
JAO1563	ACAGTACTAGCTTTTAACTTGTATCCTAGGTTATCTATGCTGTCTCACCATAGGGAATAT TACCTATTTCAG CTTGCCTCGTCCCCGCCG	To amplify <i>AmdS</i> cassette; Fwd. Homology tail to <i>CEN4</i> region.
JAO1566	GTATTTTAAGTTTTTAAAAAAGTTGATTAAATAGCATGTGAC CTCCAGTATAGCGACCAG	To amplify <i>AmdS</i> cassette; Rev. Homology tail to region immediately upstream to <i>CEN4</i> .
JAO1567	AAAAAGTATTTTAAGTTTTTAAAAAAGTTGATTAAATAGCAT CTCCAGTATAGCGACCAG	To amplify <i>AmdS</i> cassette; Rev. Integration results in 5bp deletion of <i>CEN4</i> element 1.
JAO1568	GTATTTTAAGTTTTTAAAAAAGTTGATTAAATAGCATGTGAC AGTTATGGAGTAACAACG	To amplify <i>AmdS</i> cassette excluding its terminator region; Rev. Homology tail to region immediately upstream to <i>CEN4</i> .
JAO1569	CTGCAAAACAGTACTAGCTTTTAACTTG	To confirm integration of AmdS at CEN4 region; Fwd.
JAO1570	GGAATATATAGCAGTAGTCAATTTAGCAC	To confirm integration of <i>AmdS</i> at <i>CEN4</i> region; Rev. Used for Sanger sequencing.
JAO1574	TCCAAGGTGGTTGCATCATA	Chr4 710,025 SNP Bg/II RFLP marker Fwd
JAO1575	TGGCTGGAGTTTCGTCTTCT	Chr4 710,025 SNP Bg/II RFLP marker Rev
JAO1576	GTATCTTAATGAAACTATGCAATGG	Chr4 378,184 SNP BamHI RFLP marker Fwd
JAO1577	CCTCATCGGCACATTAAAGCTG	Chr4 378,184 SNP BamHI RFLP marker Rev
JAO1659	AAAGAAAAAAATTACTGCAAAAACAGTACTAGCTTTTAACTTGTATCCTAGGTTATCTATG CTGTCTCACCAT CTTGCCTCGTCCCCGCCG	To amplify <i>AmdS</i> cassette; Fwd. Homology tail to <i>CEN4</i> region
JAO1681	TGGAGGAAAAGCATTGGTATTAAGTACTTTGCATTCTCTTGGAGAAGAACTTGATCAATT GACGGGTATAGC CTTGCCTCGTCCCCGCCG	To amplify AmdS cassette; Fwd. Homology tail to CEN14 region
JAO1682	AAAATGTTTTAAAAATATTTTTTAAAAAGCTGCACGTGACTAAC AGTTATGGAGTAACAACG	To amplify <i>AmdS</i> cassette excluding its terminator region; Rev. Homology tail to region immediately upstream to <i>CEN14</i> .
JAO1683	GGACTACTGATGTACTGAAGTTTG	To confirm integration of AmdS at CEN14 region; Fwd
JAO1684	CCTGCTGTCTAATCCGTATTCATTC	To confirm integration of AmdS at CEN14 region; Rev
JAO1685	CTGAATTGGACGCTTTGGTTC	Inside AmdS gene. To confirm insertion of AmdS cassettes; Fwd.
JAO1686	TCATCACTGTTCTTTTCAGATACTAGTTTCAAAAATTCCTTGACAGAACCATTTCATGTTC AATAATGAAAA CTTGCCTCGTCCCCGCCG	To amplify <i>AmdS</i> cassette; Fwd. Homology tail to <i>CEN15</i> region
JAO1687	AATGTTTAAATATTTAATGTATATGACTTCCGAAAAATATAT AGTTATGGAGTAACAACG	To amplify <i>AmdS</i> cassette excluding its terminator region; Rev. Homology tail to region immediately upstream to <i>CEN15</i> .
JAO1688	GGTAGTATATAAGAGAATAACTTCCC	To confirm integration of AmdS at CEN15 region; Fwd
JAO1689	GCTCTCACCACATTTATCACC	To confirm integration of AmdS at CEN15 region; Rev
JAO1692	CACAAAATATGTTCGTACCCATCC	Chr14 259,198 SNP <i>Mfe</i> II RFLP marker Fwd
JAO1693	GTGACGGAGTTTGATATACTTAAC	Chr14 259,198 SNP <i>Mfe</i> II RFLP marker Rev

Table S2 (continued). Oligonucleotides used in this study.

Name	5' – 3' sequence <sup>a</sup>	Description
JAO1694	GTTGAAGCAATGAAGGAATCTTCG	Chr14 758,547 SNP MseI RFLP marker Fwd
JAO1695	CCTCAGTCATGCCAAGTGCATCAAC	Chr14 758,547 SNP MseI RFLP marker Rev
JAO1696	CGTCTCGTATTATACTTGTCCTG	Chr15 16,733 SNP SalI RFLP marker Fwd
JAO1697	CATCGAGGAACAACAGCATCTAG	Chr15 16,733 SNP SalI RFLP marker Rev
JAO1983	CAATAATTCTCCTAGTGTTGTGC	Chr15 130,295 SNP PstI RFLP marker Fwd
JAO1984	CTGGTGTAAAGGCGTACTGAAC	Chr15 130,295 SNP PstI RFLP marker Rev
JAO1588	GAGTTATTGACAGGGAATAC	Chr15 1,058,946 SNP NdeI RFLP marker Fwd
JAO1589	GGAGCTTACTTCACTCATTTC	Chr15 1,058,946 SNP NdeI RFLP marker Rev
JAO1063	AGTCACATCAAGATCGTTTATGG	Chr3, mating type reverse primer; to amplify MATa and/or MAT $\alpha$
JAO1064	GCACGGAATATGGGACTACTTCG	Chr3, MAT $\alpha$ forward primer; to amplify MAT $\alpha$
JAO1065	ACTCCACTTCAAGTAAGAGTTTG	Chr3, MATa forward primer; to amplify MATa

a. Nucleotides in the 5' end (**bold**) have homology to the respective chromosomal insertion sites. 3' end nucleotides (not in bold) correspond to primer sequences to amplify the desired amplicons.

**Table S3.** Growth conditions used in phenotypic screening of inbred strain collection through plate spotting assay.

Base Media	Additional treatment/supplement	Temperature of incubation	Hours of incubation
2% YPD	-	30C	48
2% YPD	-	39C	96
2% YPGE	-	30C	48
2% YP Galactose	-	30C	48
2% YP Raffinose	-	30C	48
2% YPD	7 % Ethanol	30C	48
2% YPD	11 % Ethanol	30C	48
2% YPD	30 mM furfural	30C	48
2% YPD	0.75 mM menadione	30C	48
2% YPD	0.01% MMS	30C	48
2% YPD	100 mM hydroxyurea	30C	96
2% YPD	100 J/m <sup>2</sup> UV light	30C	48
2% YPD	150 J/m <sup>2</sup> UV light	30C	48

Table S4. Phenotypic data of inbreds from competition assays (CA) and tolerance to high temperature assays (HT).

G4 ·					
Strain	Cycle 0	Cycle 2	Cycle 5	Cycle 8	<sup>b</sup> Median Score in HT
JAY1598	$50.50\pm0.40$	$33.72 \pm 0.41$	$19.15\pm0.17$	$13.32 \pm 0.49$	3.50 (0.42)
JAY1600	$54.40\pm0.31$	$36.87\pm0.12$	$22.27\pm0.16$	$10.03 \pm 0.19$	1.00 (0.46)
JAY1602	$50.65\pm0.26$	$18.00\pm0.18$	$5.72\pm0.27$	$2.39\pm0.06$	3.50 (0.28)
JAY1604	$54.45\pm0.19$	$38.85\pm0.30$	$25.68\pm0.27$	$12.97 \pm 0.26$	1.00 (0.43)
JAY1606	$54.00\pm0.10$	$59.88 \pm 0.40$	$74.95\pm0.19$	$78.28 \pm 0.34$	1.00 (0.25)
JAY1608	$54.10\pm0.41$	$43.32\pm0.15$	$32.38 \pm 0.35$	$26.13\pm0.30$	3.25 (0.25)
JAY1610	$54.20\pm0.13$	$39.92\pm0.40$	$37.22\pm0.39$	$25.55 \pm 0.70$	5.00 (0.00)
JAY1612	$49.95\pm0.30$	$36.57\pm0.28$	$25.97 \pm 0.50$	$14.63\pm0.32$	4.00 (0.31)
JAY1614	$47.15\pm0.09$	$11.48\pm0.14$	$1.27\pm0.07$	$0.81\pm0.07$	3.50 (0.28)
JAY1616	$50.35\pm0.38$	$57.08 \pm 0.54$	$73.27\pm0.41$	$70.05\pm0.26$	3.00 (0.26)
JAY1618	$53.55\pm0.08$	$57.93 \pm 0.32$	$73.47\pm0.15$	$76.57\pm0.61$	1.00 (0.39)
JAY1620	$47.45\pm0.21$	$22.00\pm0.06$	$6.85\pm0.17$	$3.11 \pm 0.14$	4.00 (0.62)
JAY1622	$49.75\pm0.18$	$52.28\pm0.15$	$62.38 \pm 0.48$	59.13 ± 0.30	0.00 (0.26)
JAY1624	$47.95\pm0.26$	$38.35\pm0.43$	$31.53\pm0.45$	$21.43\pm0.51$	3.00 (0.26)
JAY1626	$48.25\pm0.20$	$53.23 \pm 0.74$	$69.08 \pm 0.25$	$76.03\pm0.24$	1.50 (0.42)
JAY1628	$47.70\pm0.40$	$67.25\pm0.46$	$92.93 \pm 0.16$	$98.15\pm0.12$	2.00 (0.00)
JAY1630	$47.60\pm0.27$	$50.40\pm0.64$	$64.62\pm0.71$	$71.98 \pm 0.22$	3.00 (0.26)
JAY1632	$48.75\pm0.32$	$48.18\pm0.79$	$63.85\pm0.31$	$70.22\pm0.17$	5.00 (0.42)
JAY1634	$45.95\pm0.48$	$46.48\pm0.91$	$51.25\pm0.52$	51.07 ± 0.39	3.00 (0.41)
JAY1636	$49.05\pm0.32$	$33.93 \pm 0.99$	$16.63\pm0.34$	$7.22\pm0.70$	3.00 (0.63)
JAY1638	$46.60 \pm 1.87$	$58.07 \pm 0.44$	$83.22\pm0.50$	93.55 ± 0.37	2.25 (0.52)
JAY1640	$47.40\pm0.74$	$63.05\pm0.98$	$90.82\pm0.14$	$97.47\pm0.10$	1.00 (0.43)
JAY1642	$40.25 \pm 1.59$	$28.00\pm0.82$	$24.68\pm0.81$	$15.60\pm0.63$	3.50 (0.13)
JAY1644	$47.95\pm0.39$	$45.53 \pm 1.02$	$62.88 \pm 0.31$	$61.22\pm0.78$	4.00 (0.21)
JAY1646	$49.20\pm0.90$	$55.67\pm0.72$	$79.45\pm0.28$	$88.7\pm0.50$	3.25 (0.25)
JAY1648	$45.40\pm0.44$	$65.70 \pm 1.03$	$93.83 \pm 0.24$	$97.42\pm0.05$	4.00 (0.21)
JAY1650	$49.45\pm0.93$	$46.10\pm0.72$	$68.23 \pm 1.57$	$70.28 \pm 1.85$	4.25 (0.21)

a. Mean value between 3 replicates plus or minus standard error.

b. Median value between 3 replicates. Standard error of the median indicated in parentheses and calculated by multiplying standard error of mean by 1.253.

Table S4 (continued). Phenotypic data of inbreds from competition assays (CA) and tolerance to high temperature assays (HT).

Stars in		bM. J'an Carne in TIT			
Strain	Cycle 0	Cycle 2	Cycle 5	Cycle 8	<sup>b</sup> Median Score in HT
JAY1652	$45.75\pm0.47$	$54.32\pm0.77$	$81.95\pm0.74$	$91.70\pm0.60$	4.25 (0.91)
JAY1654	$39.90 \pm 1.48$	$61.55\pm0.73$	$88.72\pm0.42$	$96.30\pm0.20$	4.00 (0.13)
JAY1656	$45.40\pm0.73$	$44.32\pm0.85$	$49.82\pm2.32$	$50.20\pm3.18$	4.25 (0.39)
JAY1658	$45.35\pm0.63$	$65.97 \pm 0.82$	$90.13 \pm 0.33$	$96.32 \pm 0.24$	4.00 (0.00)
JAY1660	$49.60\pm0.93$	$64.93 \pm 0.47$	$87.97 \pm 0.53$	96.30 ± 0.13	4.00 (0.26)
JAY1662	$40.60\pm0.88$	$30.02\pm0.95$	$20.12 \pm 1.01$	$12.95 \pm 0.99$	4.00 (0.26)
JAY1664	$47.05\pm0.69$	$67.30 \pm 1.42$	$92.93 \pm 0.49$	$97.93 \pm 0.20$	0.00 (0.00)
JAY1666	$51.83 \pm 0.42$	$41.30\pm0.85$	$28.87 \pm 0.30$	$18.93 \pm 0.39$	0.00 (0.00)
JAY1668	$52.20 \pm 0.58$	$28.53 \pm 0.56$	$5.56\pm0.46$	$2.29\pm0.31$	2.50 (0.75)
JAY1670	$52.02\pm0.42$	$63.97 \pm 0.81$	$71.73\pm0.68$	$74.48\pm0.56$	3.00 (0.10)
JAY1672	$50.53\pm0.36$	$51.68 \pm 0.99$	$45.70\pm0.93$	$25.42\pm0.72$	0.50 (0.42)
JAY1674	$49.07\pm0.40$	$37.38 \pm 0.49$	$18.28\pm0.53$	$9.05\pm0.32$	3.00 (0.13)
JAY1676	$50.70\pm0.48$	$51.02 \pm 1.44$	$45.40\pm0.58$	$29.93 \pm 0.55$	0.50 (0.42)
JAY1678	$49.55\pm0.20$	$41.27 \pm 1.36$	$50.87 \pm 0.58$	$46.52\pm0.79$	1.00 (0.39)
JAY1680	$45.35\pm0.47$	$59.17 \pm 0.39$	$28.92 \pm 1.55$	$38.80\pm0.58$	3.00 (0.21)
JAY1682	$50.23\pm0.71$	$45.62 \pm 1.01$	$47.33\pm0.62$	$34.42\pm0.52$	3.00 (0.42)
JAY1684	$49.00\pm0.31$	$36.33\pm0.68$	$14.77\pm0.68$	$5.20\pm0.36$	4.00 (0.26)
JAY1686	$52.93 \pm 0.42$	$31.18\pm0.96$	$28.03\pm0.29$	$17.82\pm0.52$	4.00 (0.26)
JAY1688	$52.82\pm0.42$	$30.47 \pm 1.46$	$24.83\pm0.45$	$14.92\pm0.22$	5.00 (0.00)
JAY1690	$49.32\pm0.37$	$28.15\pm0.42$	$8.07\pm0.38$	$2.89\pm0.29$	5.00 (0.13)
JAY1692	$43.13\pm0.34$	$32.80 \pm 1.55$	$24.40 \pm 1.12$	$15.27\pm0.74$	4.00 (0.00)
JAY1694	$48.32\pm0.67$	$30.17 \pm 1.76$	$27.12 \pm 1.13$	$14.50 \pm 1.14$	3.00 (0.00)
JAY1696	$44.93\pm0.49$	$44.48\pm0.98$	$31.73 \pm 1.56$	$33.77\pm0.86$	4.25 (0.25)
JAY1698	$52.42\pm0.53$	$48.17 \pm 1.22$	$31.33 \pm 1.40$	$16.68\pm0.68$	3.00 (0.00)
JAY1700	$48.68\pm0.31$	$32.05 \pm 1.64$	$26.65\pm0.45$	$2.93\pm0.22$	5.00 (0.00)
JAY1702	$50.48 \pm 0.73$	$30.12 \pm 1.45$	$17.13 \pm 0.34$	$2.96\pm0.46$	2.50 (0.51)

a. Mean value between 3 replicates plus or minus standard error.

b. Median value between 3 replicates. Standard error of the median indicated in parentheses and calculated by multiplying standard error of mean by 1.253.

Table S4 (continued). Phenotypic data of inbreds from competition assays (CA) and tolerance to high temperature assays (HT).

Strain					
	Cycle 0	Cycle 2	Cycle 5	Cycle 8	<sup>b</sup> Median Score in HT
JAY1704	$49.87\pm0.45$	$55.72\pm0.49$	$71.50\pm0.42$	84.67 ± 0.30	4.50 (0.38)
JAY1706	$50.80\pm0.96$	$64.72\pm0.45$	$86.90\pm0.29$	$96.1\pm0.41$	3.50 (0.13)
JAY1708	$53.53 \pm 0.48$	$81.92\pm0.25$	$92.57 \pm 0.25$	$97.25\pm0.21$	3.00 (0.26)
JAY1710	$46.03\pm0.37$	$25.37\pm0.54$	$25.25\pm0.79$	$24.93 \pm 1.63$	5.00 (0.21)
JAY1712	$51.80\pm0.32$	$19.22 \pm 0.67$	$16.70\pm0.50$	$16.70\pm0.72$	4.00 (0.00)
JAY1714	$52.30\pm0.34$	$24.70\pm0.54$	$22.67\pm0.60$	$23.55 \pm 0.37$	3.00 (0.63)
JAY1716	$51.90\pm0.22$	$7.05\pm0.32$	$3.56\pm0.51$	$4.00\pm0.56$	2.50 (0.65)
JAY1718	$51.47\pm0.27$	$46.05 \pm 0.35$	$57.65\pm0.55$	$75.33 \pm 0.87$	2.00 (0.52)
JAY1720	$49.82\pm0.28$	$41.17 \pm 1.24$	$43.68 \pm 1.19$	$35.63 \pm 1.01$	1.50 (0.54)
JAY1722	$46.68\pm0.39$	$7.64\pm0.76$	$5.25\pm0.38$	$4.94\pm0.53$	3.00 (0.00)
JAY1724	$43.30\pm0.75$	$18.58\pm0.55$	$14.02\pm0.42$	$8.49\pm0.40$	1.50 (0.28)
JAY1726	$43.73\pm0.59$	$8.52\pm0.35$	$4.18\pm0.43$	$5.11 \pm 0.45$	4.00 (0.13)
JAY1728	$39.18\pm0.94$	34.82 ± 2.23	$25.07 \pm 1.19$	$16.42\pm0.36$	3.00 (0.26)
JAY1730	$40.37\pm0.27$	33.60 ± 2.30	$23.30\pm0.78$	$17.52 \pm 1.46$	1.00 (0.26)
JAY1732	$38.78 \pm 2.23$	$13.20 \pm 0.85$	$4.97\pm0.73$	$1.68\pm0.17$	3.00 (0.26)
JAY1734	$51.42\pm0.35$	$36.17\pm0.54$	$12.00\pm0.40$	$3.91\pm0.49$	4.00 (0.21)
JAY1736	$50.88 \pm 0.33$	$59.33 \pm 0.47$	$69.98 \pm 0.49$	$79.22\pm0.42$	3.00 (0.00)
JAY1738	$51.72\pm0.35$	$55.88 \pm 0.70$	$58.42 \pm 1.26$	$61.85 \pm 1.41$	3.00 (0.26)
JAY1740	$47.28\pm0.35$	$35.82\pm0.53$	$19.05\pm0.62$	$10.22\pm0.56$	4.00 (0.00)
JAY1742	$53.95\pm0.25$	$69.72 \pm 0.25$	$90.65\pm0.28$	$97.78 \pm 0.10$	1.50 (0.28)
JAY1744	$55.13\pm0.27$	$65.48 \pm 0.32$	$86.57\pm0.46$	$95.35\pm0.05$	0.50 (0.36)
JAY1746	$48.22\pm0.24$	$42.45 \pm 0.53$	$41.25\pm0.61$	$38.33 \pm 0.80$	3.00 (0.13)
JAY1748	$51.38\pm0.38$	38.30 ± 0.33	$22.48\pm0.18$	$12.52\pm0.36$	2.50 (0.23)
JAY1750	$48.55\pm0.59$	$15.60 \pm 0.43$	$4.05\pm0.47$	$1.83\pm0.33$	4.00 (0.13)
JAY1752	$48.17\pm0.81$	$15.15 \pm 0.13$	$4.11 \pm 0.49$	$2.31 \pm 0.50$	3.00 (0.13)

a. Mean value between 3 replicates plus or minus standard error.

b. Median value between 3 replicates. Standard error of the median indicated in parentheses and calculated by multiplying standard error of mean by 1.253.

**Table S5.** List of phased JAY270 HetSNPs interrogated for confirmation of UPD strains.

Marker SGD	coordinates	PCR primers	Polymorphism	<sup>a</sup> SGD Watson	JAY270 Watson phased bases	
Chromosome	Nucleotide	rckprimers	detection method	bases	Chr-P	Chr-M
Chr4	378,184	JAO1576/JAO1577	BamHI	<u>C</u>	Т	<u>C</u>
Chr4	710,025	JAO1574/JAO1575	BglII	<u>T</u>	G	<u>T</u>
Chr14	259,198	JAO1692/JAO1693	MfeI	<u>T</u>	<u>T</u>	А
Chr14	758,547	JAO1694/JAO1695	MseI	A	G	<u>A</u>
Chr15	16,733	JAO1696/JAO1697	SalI	<u>C</u>	Т	<u>C</u>
Chr15	130,295	JAO1983/JAO1984	PstI	<u>A</u>	<u>A</u>	С
Chr15	1,058,946	JAO1588/JAO1589	NdeI	Т	Т	<u>C</u>

a. Nucleotides that are <u>underlined</u> indicate the sequences that are cut by the respective restriction endonucleases, whereas lack of underlining corresponds to no cut site.

Table S6. Regions showing significant association to the heat tolerance and competitive growth phenotypes and their gene contents.

Phenotype	Chr	Genome Coordinates	HetSNPs	Max LOD	Total Genes	Genes with HetSNPs	Genes with Nonsynonymou s Substitution(s)	List of Genes with Nonsynonymous Substitution(s)
	2	Chr02_207698214801 (7.1 Kb)	14	4.45	2	2	1	YBL008W-A
	5	Chr05_549677565641 (16.0 Kb)	40	5.47	9	2	2	ISC10, SLO1
	6	Chr06_7481497759 (22.9 Kb)	91	6.23	14	13	8	HAC1, CAF16, GYP8, STE2, BST1, BUD27, FRS2, GAT1
	7	Chr07_8275388718 (6.0 Kb)	11	4.61	4	2	1	KIP3
	7	Chr07_673392730469 (57.1 Kb)	9	7.12	30	2	0	none
	7	Chr07_849524887971 (38.4 Kb)	14	5.56	20	3	0	none
	12	Chr12_176348201613 (25.3 Kb)	88	4.28	14	10	6	PRS2, YEH2, IRC25, SDO1, UBR2, ADE16
Heat Tolerance	13	Chr13_7504092531 (17.5 Kb)	49	8.67	12	9	4	TAF13, YML096W, UTP14, RPM2
Tolerance	13	Chr13_715996749967 (34.0 Kb)	13	8.70	19	5	2	MRE11, BCH1
	13	Chr13_754723814438 (59.7 Kb)	59	5.20	37	19	13	YMR244W, YMR253C, GFD1, PET111, TRM732, TPS3, RSN1, PPA2, PRP24, RRN9, URA10, SCS7, ZDS1
	14	Chr14_528514605480 (77.0 Kb)	66	11.06	47	14	9 (1 nonsense)	COG5, SFB2, ALG11, SLM2 (nonsense), YNL034W, KTR5, CRZ1, SAM50, FAP1
	14	Chr14_691878695118 (3.2 Kb)	6	5.26	3	1	0	none
	15	Chr15_764174781222 (17.0 Kb)	13	6.01	10	1	1	YOR228C
	3	Chr03_7784984690 (6.8 Kb)	44	4.34	5	3	2	KCC4, YCL021W-A
	4	Chr04_556996592486 (35.5 Kb)	99	6.00	24	18	11	DBF4, PST1, MAK21, YDR061W, LCB2, AIM7, RRG1, RRTR2, DOS2, DOA4, IPT1
	8	Chr08_3514951314 (16.2 Kb)	26	4.68	7	2	2	ECM29, WSC4
Competitive Growth	10	Chr10_467589569699 (102.1 Kb)	48	8.30	56	17	8	REC107, YJR030C, GEA1, CPR7, RAD26, URB2, TOR1, YAE1
	15	Chr15_882911889659 (6.7 Kb)	3	4.13	2	1	0	none
	15	Chr15_10248201069385 (44.6 Kb)	138	4.31	23	12	10	YOR365C, GFB1, NDD1, NUD1, YOR376W-A, ATF1, FRE3, YOR381W-A, YOR385W, PHR1
	16	Chr16_289873361102 (71.2 Kb)	134	5.93	40	22	15	SPP1, GIP3, ISU1, COX11, SPO19, TBF1, HHO1, NAN1, KAP120, MRP51, HOS3, CAR1, YPL109C, YPL108W, FMP30

Table S7. Single locus analysis of variance for heat tolerance and competitive growth phenotypes in the inbred strains.

Phenotype	Locus	Significance	<sup>a</sup> PVE	<sup>b</sup> Relative PVE	Inheritance model	Parent Contributing high value allele
	Chr02_209988	< 0.0001	23.12	0.48	Additive	Р
	Chr05_564935	< 0.0001	27.59	0.58	Dominant	Р
	Chr06_87106	< 0.0001	30.77	0.64	Additive	М
	Chr07_86432	< 0.0001	23.82	0.50	Additive	М
	Chr07_677969	< 0.0001	34.34	0.72	Additive	Р
	Chr07_858098	< 0.0001	27.99	0.58	Additive	Р
Heat Tolerance	Chr12_188842	< 0.0001	22.31	0.47	Dominant	Р
Toterance	Chr13_83905	< 0.0001	40.04	0.84	Additive	М
	Chr13_743742	< 0.0001	40.16	0.84	Additive	Р
	Chr13_799417	< 0.0001	26.45	0.55	Additive	Р
	Chr14_572098	< 0.0001	47.95	1.00	Additive	М
	Chr14_691878	< 0.0001	26.68	0.56	Dominant	М
	Chr15_764348	< 0.0001	29.86	0.62	Additive	М
	Chr03_82057	< 0.0001	22.6	0.58	Additive	Р
	Chr04_560433	< 0.0001	29.82	0.77	Additive	М
	Chr08_49196	< 0.0001	24.13	0.62	Additive	Р
Competitive Growth	Chr10_488369	< 0.0001	38.74	1.00	Additive	Р
Giowin	Chr15_888521	< 0.0001	21.64	0.56	Additive	Р
Į Į	Chr15_1055015	< 0.0001	22.47	0.58	Dominant	М
	Chr16_340133	<0.0001	29.52	0.76	Additive	М

a. The PVE values obtained through this single-QTL model are likely over-estimates of the values.b. PVE values normalized to highest value PVE for each phenotype.

**Table S8.** Multi-locus QTL models that best explain variation seen in heat tolerance and competitive growth inbred strains.

Phenotype	Locus	Significance	Total Percent Variance Explained
	Chr06_87106	< 0.0001	
Heat Tolerance	Chr13_743742	< 0.0001	75.88
	Chr14_572098	< 0.0001	
	Chr04_560433	0.0015	
Competitive Growth	Chr10_488369	< 0.0001	57.27
	Chr15_1055015	0.0047	