

Supplementary Figure 1. Phagocytosis by *C. robusta* haemocytes.

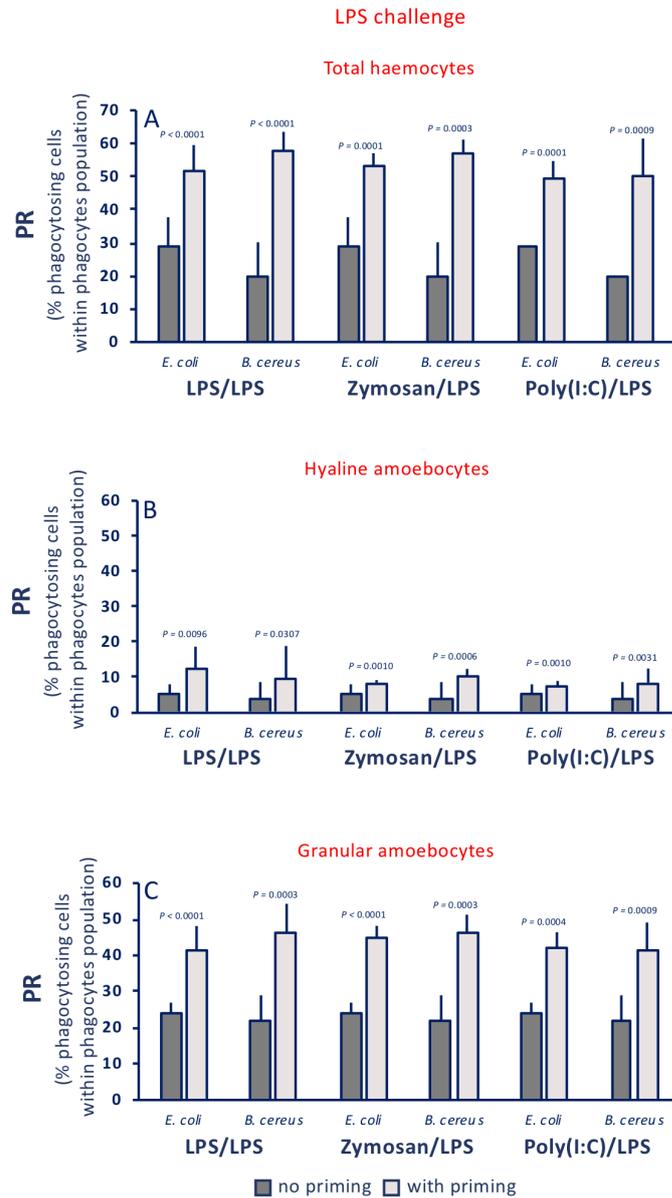
Phagocytosis was performed *in vitro* by exposing *C. robusta* haemocytes to fluorescent *B. cereus* bacteria for 1 h at 18°C.

Panels A and B: haemocytes from control animals phagocytosing fluorescent bacteria.

Panels C and D: haemocytes from LPS-treated animals phagocytosing fluorescent bacteria.

Panels A and C: dark field, panels B and D: white field.

Haemocyte nuclei were labelled with the nucleic acid dye DAPI (blue colour), bacteria were labelled with FITC (green colour). Scale bar 10 μm.



Supplementary Figure 2. Innate memory-induced variations in the phagocytic rate of *C. robusta* haemocytes challenged *in vivo* with LPS.

The phagocytic rate (PR), *i.e.*, the percentage of phagocytosing cells within the phagocyte population in haemolymph, was assessed 24 h after primary exposure to LPS, Zymosan or Poly(I:C) (dark grey columns, no priming), and after a 24-h LPS challenge of primed animals (light grey columns, with priming).

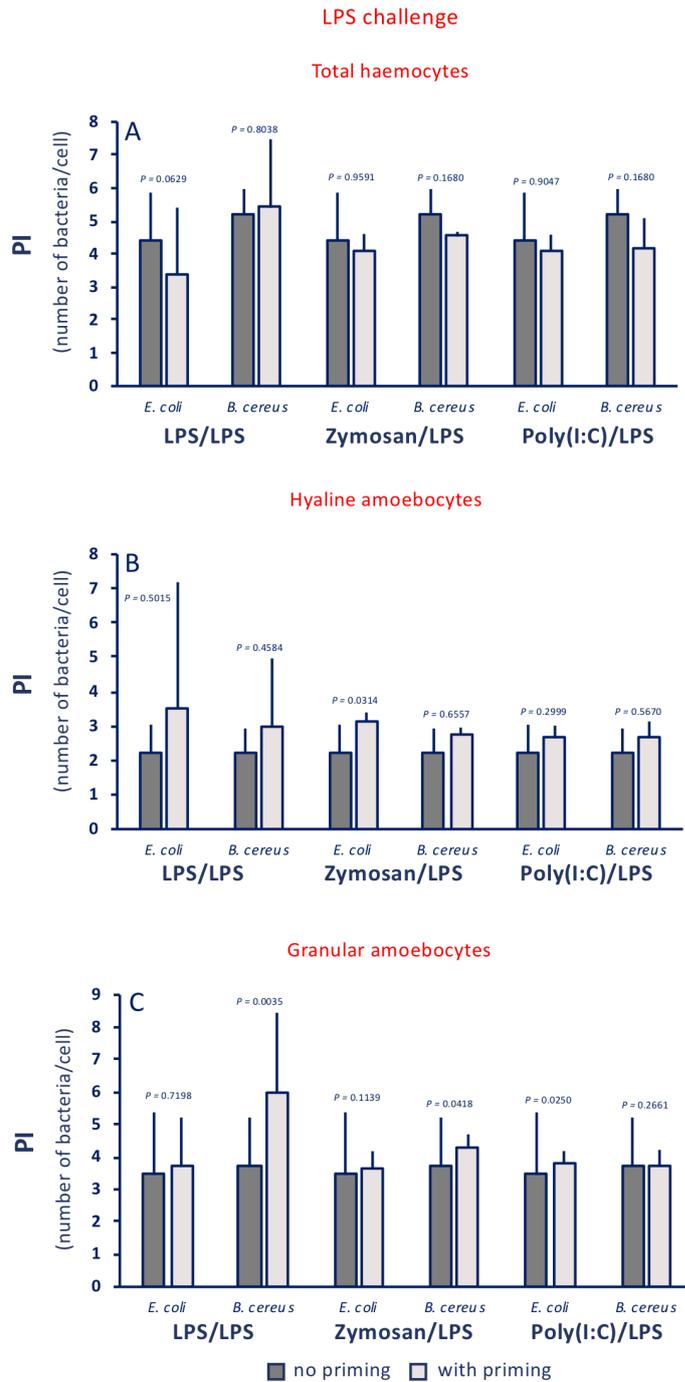
Panel A: PR of the total haemocyte population for two bacterial strains, the gram-negative *E. coli*, and the gram-positive *B. cereus*. The PR of control animals receiving MS once or twice (controls) was $17.9 \pm 3.7\%$ for *E. coli*, and $22.4 \pm 1.8\%$ for *B. cereus*. The mean values \pm SD of 5 animals are reported.

Panel B: PR of hyaline cells, *i.e.*, one of the two phagocytic amoebocyte subpopulations. The PR of control hyaline cells (from animals receiving MS once or twice) was 1.5 ± 1.1 for *E. coli* and 3.5 ± 0.4 for *B. cereus*.

Panel C: PR of granular cells, *i.e.*, the other phagocytic amoebocyte subpopulation. The PR of granular cells (from animals receiving MS once or twice) was 16.5 ± 1.8 for *E. coli* and 19.0 ± 0.9 for *B. cereus*.

The mean values \pm SD of 5 animals for each treatment are reported.

The data for the LPS/LPS combinations are the same reported in the Figure 3 and are repeated here for comparison.



Supplementary Figure 3. Innate memory-induced variations in the phagocytic index of *C. robusta* haemocytes challenged *in vivo* with LPS. The phagocytic index (PI), *i.e.*, the number of bacteria ingested by single phagocytes in haemolymph, was assessed 24 h after primary exposure to LPS, Zymosan or Poly(I:C) (dark grey columns, no priming), and after a 24-h LPS challenge of primed animals (light grey columns, with priming). Panel A: PI of the total haemocyte population for two bacterial strains, the gram-negative *E. coli*, and the gram-positive *B. cereus*. The PI of haemocytes from control animals receiving MS once or twice (controls) was $17.9 \pm 3.7\%$ for *E. coli*, and $22.4 \pm 1.8\%$ for *B. cereus*. The mean values \pm SD of 5 animals are reported. Panel B: PI of hyaline cells, *i.e.*, one of the two phagocytic amoebocyte subpopulations. The PI of control hyaline cells 0.9 ± 0.4 for *E. coli* and 1.5 ± 0.7 for *B. cereus*. Panel C: PI of granular cells, *i.e.*, the other phagocytic amoebocyte subpopulation. The PI of control granular cells was 3.0 ± 1.1 for *E. coli* and 3.4 ± 0.9 for *B. cereus*. The mean values \pm SD of 5 animals for each treatment are reported. The data for the LPS/LPS combinations are the same reported in the Figure 4 and are repeated here for comparison.

Supplementary Table 1. Priming-dependent variations of gene expression levels in primed/challenged *C. robusta*

<i>Gene</i>	Priming-dependent changes in the response to any challenge		
	<i>LPS priming-dependent changes</i>	<i>LTA priming-dependent changes</i>	<i>Common priming changes</i>
<i>C3-1</i>	-	Up (above bkg)	-
<i>C3ar</i>	-	-	-
<i>Il17-2</i>	-	Up (above bkg)	-
<i>Il17r</i>	-	-	-
<i>Tnf</i>	-	-	Up (above bkg or normalised)
<i>Tgfb</i>	-	-	-
<i>Lbp</i>	-	-	Up (above bkg)
<i>Tlr-2</i>	-	-	-
<i>Tlr13</i>	-	-	-
<i>Cd36</i>	-	-	-

Supplementary Table 2. Challenge-dependent variations of gene expression levels in primed/challenged *C. robusta*

<i>Gene</i>	Challenge-dependent changes in the response to any priming		
	<i>LPS challenge-dependent changes</i>	<i>LTA challenge-dependent changes</i>	<i>Common challenge changes</i>
<i>C3-1</i>	-	Up (above bkg)	-
<i>C3ar</i>	Down (below bkg)	Up (above bkg)	-
<i>Il17-2</i>	-	Up (above bkg)	-
<i>Il17r</i>	Up (normalised)	-	-
<i>Tnf</i>	-	-	Up (above bkg or normalised)
<i>Tgfb</i>	-	-	-
<i>Lbp</i>	-	-	Up (above bkg or normalised)
<i>Tlr-2</i>	-	-	-
<i>Tlr13</i>	Up (above bkg)	-	-
<i>Cd36</i>	Up (above bkg)	-	-

Supplementary Table 3. Priming/challenge-specific variations of gene expression levels in primed/challenged *C. robusta*

Gene	Priming-dependent changes in the response to individual challenges			
	<i>LPS priming/LPS challenge-dependent changes</i>	<i>LTA priming/LTA challenge-dependent changes</i>	<i>LPS priming/LTA challenge-dependent changes</i>	<i>LTA priming/LPS challenge-dependent changes</i>
<i>C3-1</i>	none	-	-	-
<i>C3ar</i>	-	-	-	-
<i>Il17-2</i>	none	-	-	-
<i>Il17r</i>	-	-	-	-
<i>Tnf</i>	-	-	-	-
<i>Tgfb</i>	Down (below bkg)	-	Up (above bkg)	-
<i>Lbp</i>	-	-	-	-
<i>Tlr-2</i>	-	none	-	-
<i>Tlr13</i>	-	none	none	-
<i>Cd36</i>	-	Down (normalised)	-	-