

## Supplementary Materials

### **Rapid Nanopore Assay for Carbapenem-resistant *Klebsiella Pneumoniae***

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## **Expression and purification of MspA nanopore**

BL21 Competent *E. coli* is a widely used strain and is suitable for transformation and protein expression. The gene of MspA nanopore was cloned into the pET-28b plasmid, and the pET-28b plasmid carried with MspA gene was transferred into engineering bacteria *E. coli* BL21 (Niederweis et al., 1999). Successfully transferred *E. coli* was cultured in the LB media at 37°C, and the kanamycin was added to 50 µg/mL. While the optical density (600 nm) was close to 0.8, the 0.8 mM IPTG was added into the LB (lysogeny broth) media with an inducing temperature of 15°C. After 12h of induction, the *E. coli* was collected through centrifugation. The supernatant was collected after the *E. coli* was broken by the ultrasonicator, and further purified by anion-exchange column (Q-Sepharose) and molecular sieve (Superdex 200 16/90). The purified protein was analyzed by 10% SDS-PAGE (Sodium dodecyl sulfate polyacrylamide gel electrophoresis).

The purified MspA nanopore protein can be aliquoted and stored at – 80 °C. The aliquots remain stable for years, and the nanopore remains active upon thaw (Haque et al., 2013).

## **Probe design**

We design probes to combine with specific fragments of 16S rRNA, so that we can recognize specific signals regarding the target nucleic acid through nanopore. Since the target 16S rRNA has a long length of 932 bp, it is difficult to distinguish the 16S rRNA-probe complex without probes or by a single probe, thus two probes were designed to bind the specifically expressed 16S rRNA of KP (**Figure 2A**). Probe A bound to the conserved area of 16S rRNA (810 nt-937 nt) was used to help the 16S rRNA-probe complex keep consistent secondary structure and increase dwell time which can be distinguished easily by MspA ; Probe B bound to the specific area of 16S rRNA (10 nt-136 nt) has been reported to be 100% specific and we used it to increase the specificity (Poh et al., 2004).

## REFERENCES

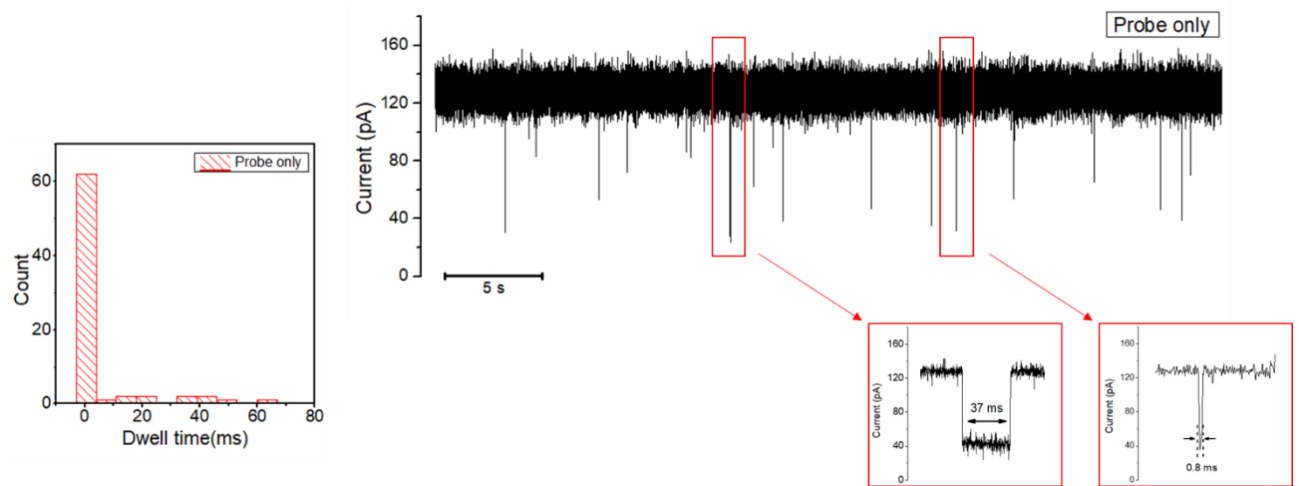
- Niederweis, M., Ehrt, S., Heinz, C., Klöcker, U., Karosi, S., Swiderek, K. M., et al. (1999). Cloning of the mspA gene encoding a porin from Mycobacterium smegmatis. *Mol. Microbiol.* 33, 933–945. doi:10.1046/j.1365-2958.1999.01472.x.
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**Table S1. Clinical sample information of CRKP.** The Sample ID is the patient ID in hospital and the Sample number is the corresponding number in our study.

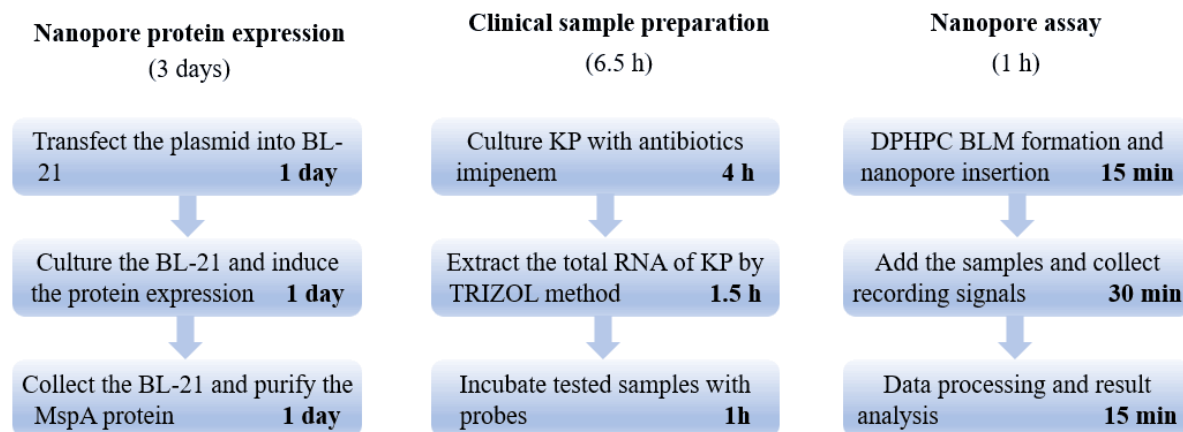
Sample ID	Sample #	SCIM (mm)	Drug resistance gene	Drug resistance gene
17012889-3	1	6	KPC	KPC-2
17019349-3	3	6	KPC	KPC-2
1810143046	4	6	KPC	KPC-2
15043287-1	5	6	KPC	KPC-2
15057156-1	6	6	KPC	KPC-2
15083593-1	7	6	KPC	KPC-2
1807191036	8	6	KPC	KPC-2
1807271015	9	24	Negative	-
17008404-1	11	6	KPC	None
17012837-3	12	6	KPC	KPC-2
17020362-3	20	6	KPC	KPC-2

**Table S2. Clinical sample information of CSKP.** The Sample ID is the patient ID in hospital and the Sample number is the corresponding number in our study.

Sample ID	Sample #	Detected bacteria	Drug-resistant form	SCIM-IPM (mm)
1809163029	2	CSKP	ESBL	34
1807171076	10	CSKP	ESBL	33
1807163074	13	CSKP	-	-
1806163019	14	CSKP	-	-
1807043013	15	CSKP	ESBL	33
1806301100	16	CSKP	ESBL	34
1806133062	17	CSKP	-	-
1806083069	18	CSKP	ESBL	32
1806163021	19	CSKP	ESBL	31



**Figure S1. Translocation signal of probe only group.** The dwell time of the DNA probe translocation through nanopore is in the range of 0 to 70 ms.



**Figure S2. Flow chart of the experiment and time cost in total.** The MspA protein is stored in the refrigerator at -80°C for use after expression and purification for further test. There is no need to prepare the nanopore for each experiment.