

# Reconstitution of the tumor microenvironment in a microfluidic platform comprising a bone-mimetic hydroxyapatite/fibrin composite

Jungho Ahn<sup>1,2,†</sup>, Jungeun Lim<sup>1,2,†</sup>, Norhana Jusoh<sup>1,†</sup>, Jungseub Lee<sup>1</sup>, Tae-eun Park<sup>3</sup>, YongTae Kim<sup>2,4,5,6</sup>, Jangho Kim<sup>7,\*</sup> and Noo Li Jeon<sup>1,8,9,10,\*</sup>

<sup>1</sup>Department of Mechanical and Aerospace Engineering, Seoul National University, Seoul 08826, Republic of Korea

<sup>2</sup>George W. Woodruff School of Mechanical Engineering, Georgia Institute of Technology, Atlanta, GA 30332, USA

<sup>3</sup>Ulsan National Institute of Science and Technology, Ulsan 44914, South Korea

<sup>4</sup>Parker H. Petit Institute for Bioengineering and Bioscience, Georgia Institute of Technology, Atlanta, GA 30332, USA

<sup>5</sup>Institute for Electronics and Nanotechnology, Georgia Institute of Technology, Atlanta, GA 30332, USA

<sup>6</sup>Wallace H. Coulter Department of Biomedical Engineering, Georgia Institute of Technology, Atlanta, GA 30332, USA

<sup>7</sup>Department of Rural and Biosystems Engineering, Chonnam National University, Gwangju, 500-757, South Korea

<sup>8</sup>Division of WCU (World Class University) Multiscale Mechanical Design, Seoul National University, Seoul 08826, Republic of Korea

<sup>9</sup>Seoul National University Institute of Advanced Machines and Design, Seoul 08826, Republic of Korea

<sup>10</sup>Institute of Bioengineering, Seoul National University, Seoul, Republic of Korea

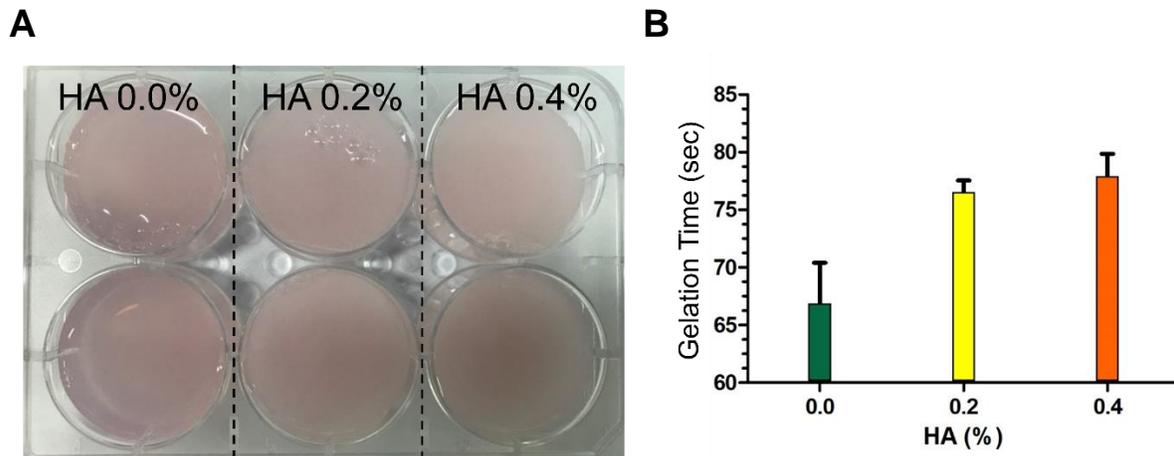
**\* Correspondence:**

Corresponding Author

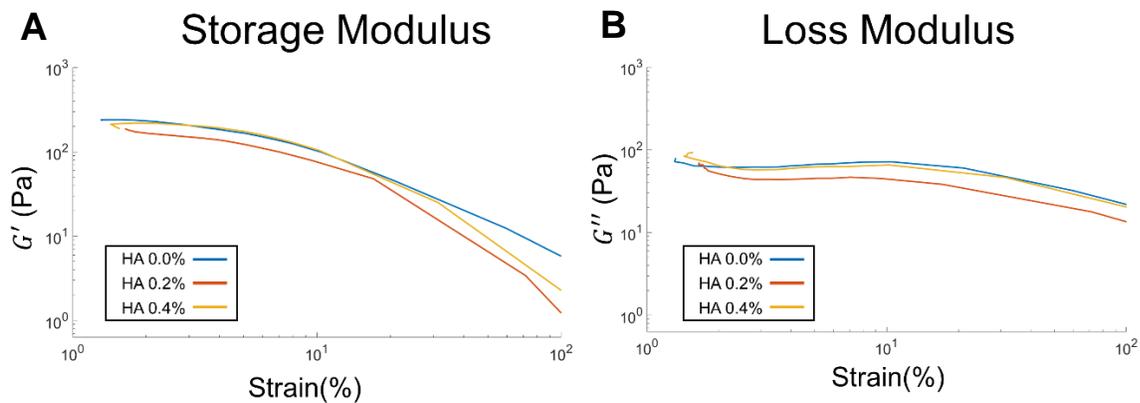
[njeon@snu.ac.kr](mailto:njeon@snu.ac.kr)

**† Equally Contributed:** These authors contributed equally

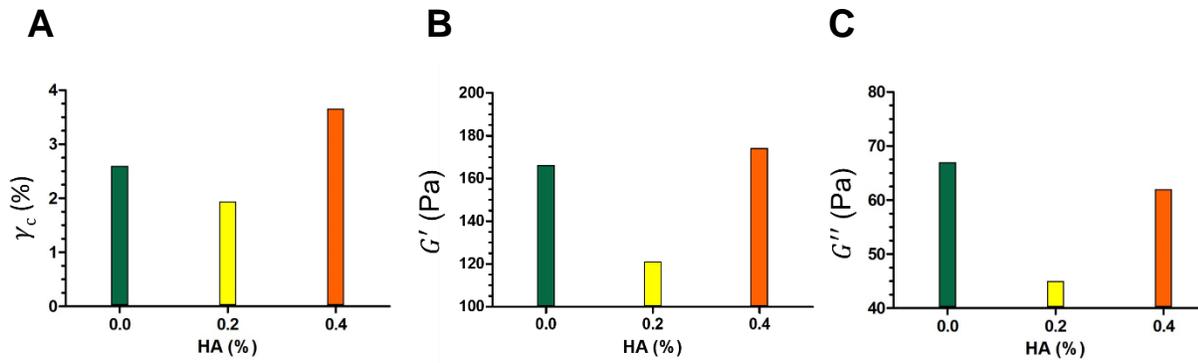
**Keywords:** Tumor microenvironment, Vascularized tumor, Hydroxyapatite, Fibrin matrix



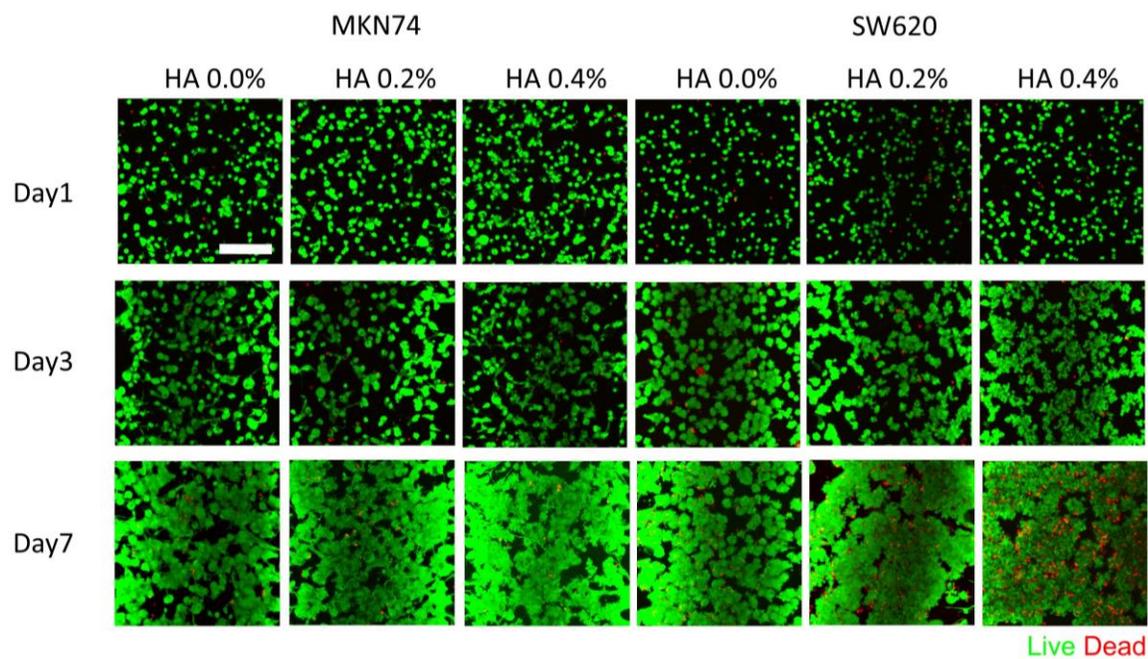
**Figure S1. Gelation time for HA/fibrin composite with each concentration of HA.** (A) HA/fibrin composite in 6-well plate with 0.0, 0.2 and 0.4% HA contents respectively. (B) Time taken to gelate completely after reacting with thrombin with varying concentration of HA components (0.0, 0.2 and 0.4 %).



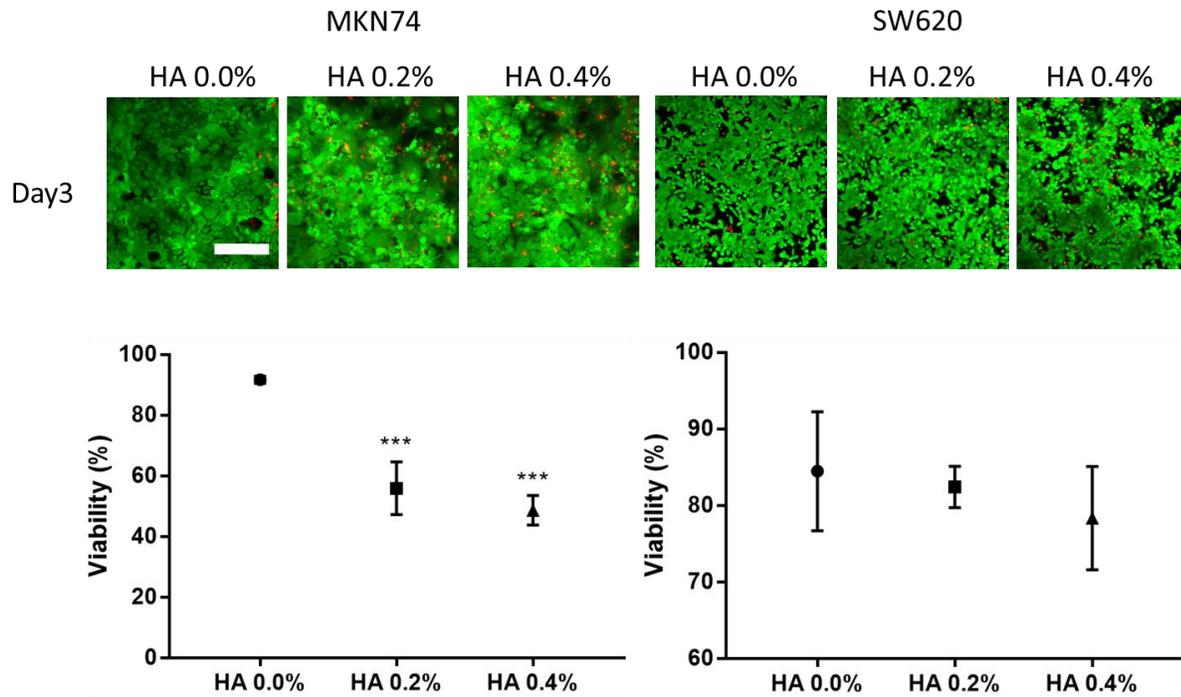
**Figure S2. Gel rheology analysis as a function of strain for HA/fibrin composite with each concentration of HA.** The (A) storage and (B) loss modulus as a function of strain at the same temperature and oscillation frequency in gel rheology analysis for HA/fibrin composite with varying concentration of HA contents (0.0, 0.2 and 0.4 %).



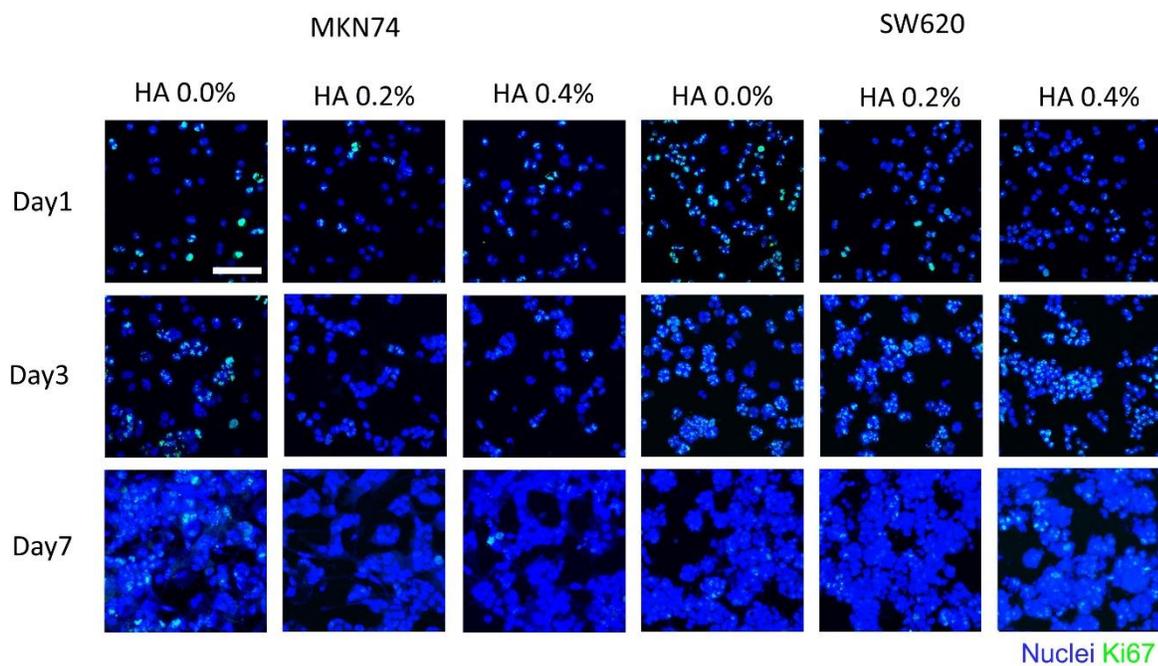
**Figure S3. Critical strain and value of modulus of HA/fibrin composite for each HA concentration.** (A) The critical strain of HA/fibrin composite for each concentration of HA. The (B) storage and (C) loss modulus at 5% strain state in gel rheology analysis for HA/fibrin composite with varying concentration of HA components (0.0, 0.2 and 0.4 %).



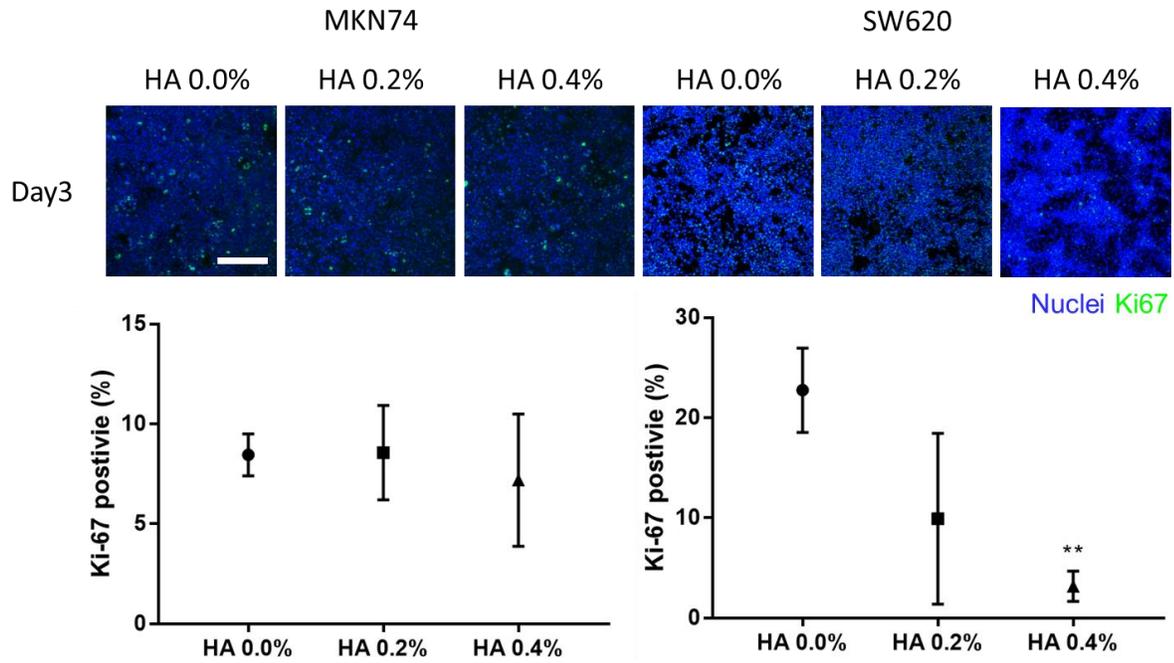
**Figure S4.** Confocal live/dead images of MKN74 and SW620 respectively in different HA concentration and culture time points. (green: calcein-AM, red: ethd-1) (n= 4-6 chips per condition) (Scale bar: 200  $\mu$ m)



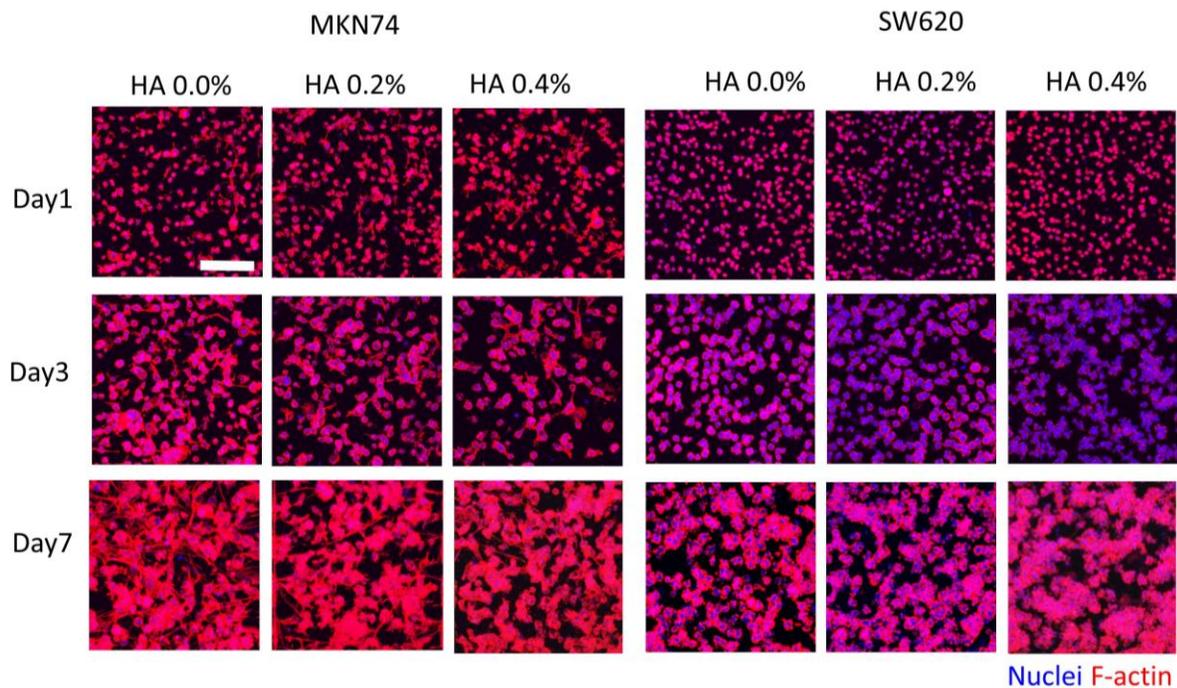
**Figure S5.** Tumor cell viability in 2D culture with varying HA concentration.



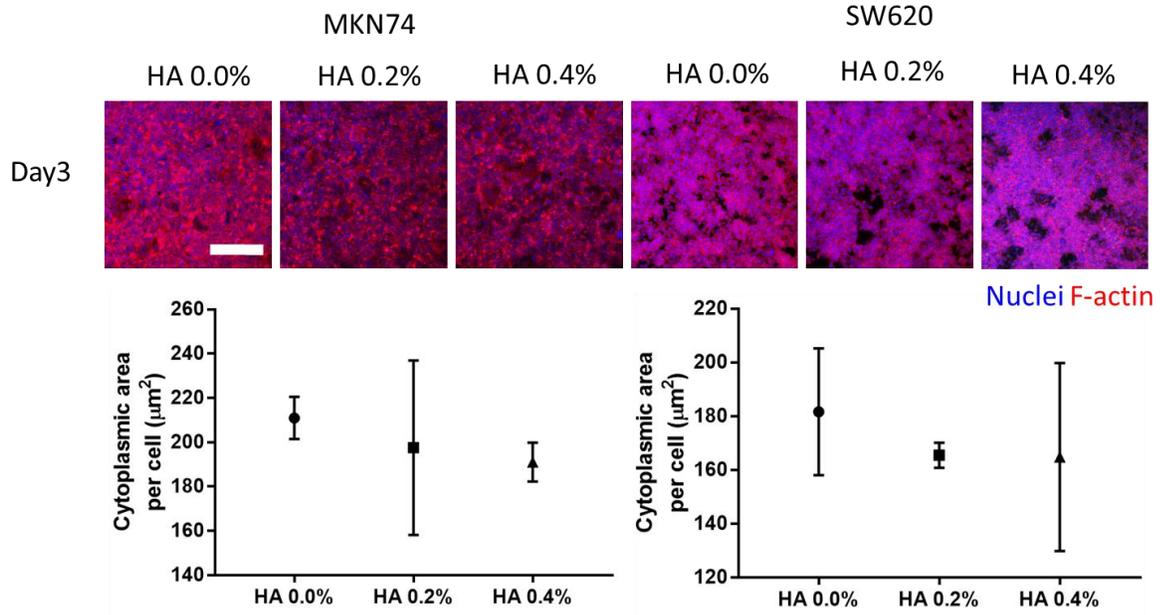
**Figure S6.** Confocal images for cell proliferation of MKN74 and SW620 respectively in different HA concentration and culture time points. (green: Ki-67, blue: nuclei) (n= 4-6 chips per condition) (Scale bar: 200  $\mu$ m)



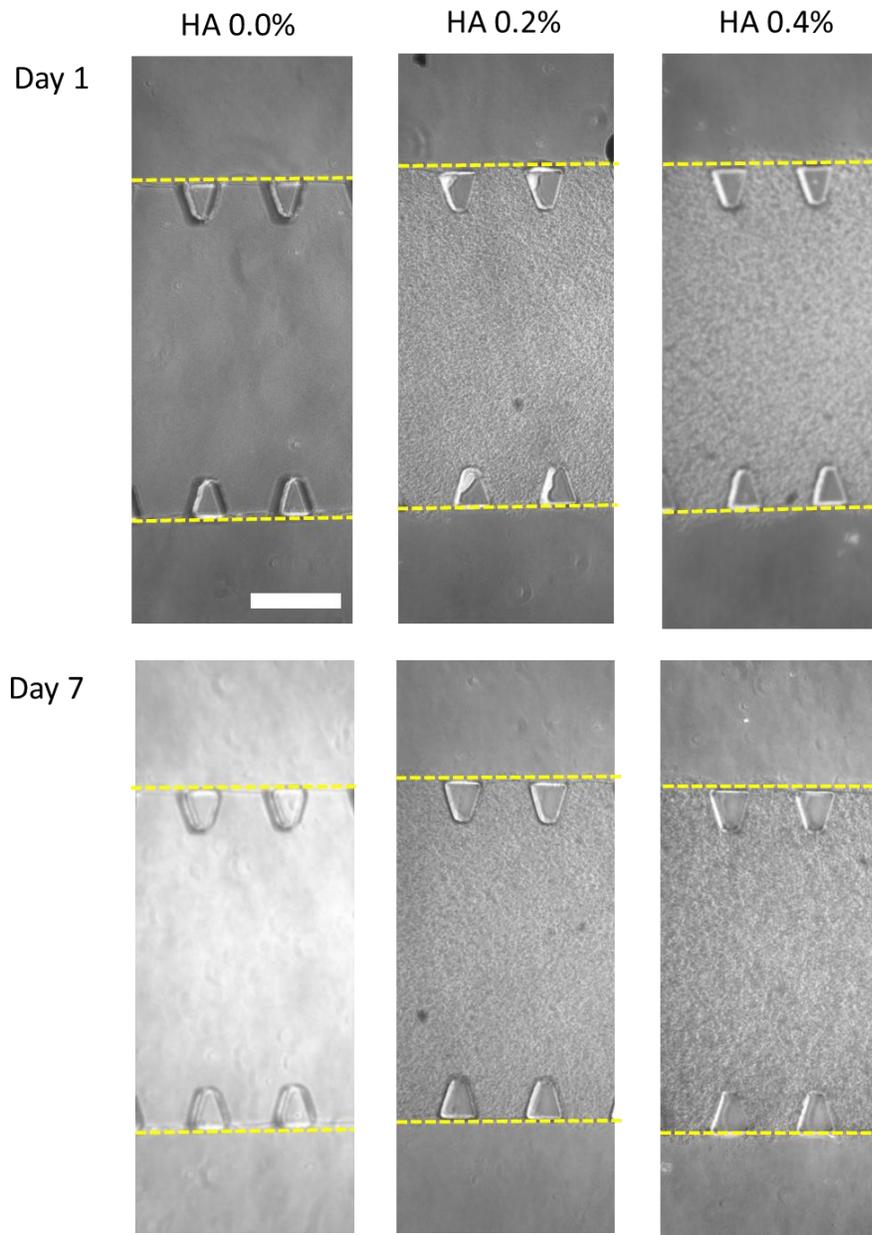
**Figure S7.** Tumor cell proliferation in 2D culture with varying HA concentration. (Scale bar: 200  $\mu\text{m}$ )



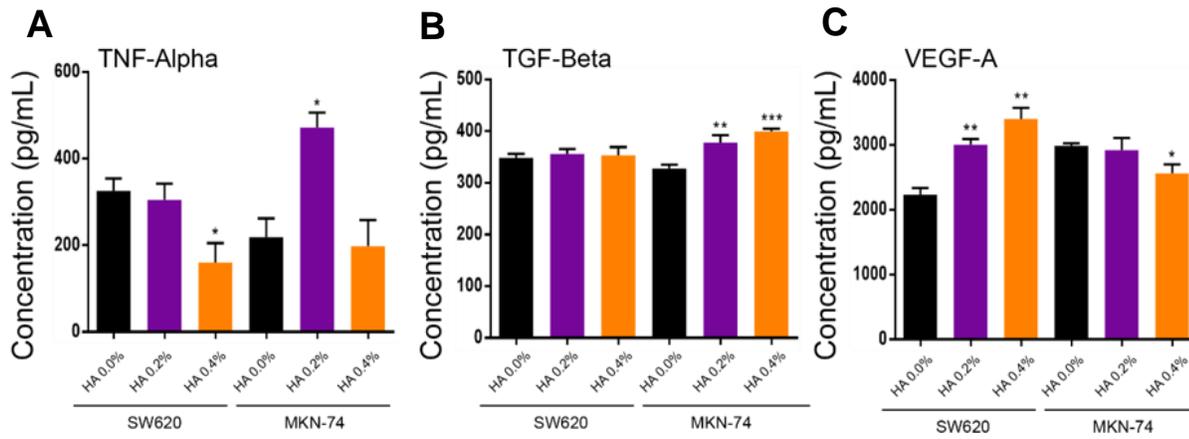
**Figure S8.** Confocal image analysis of tumor cell morphology of MKN74 and SW620 respectively in different HA concentration and culture time points. (red: F-actin, blue: nuclei) (n= 4-6 chips per condition) (Scale bar: 200  $\mu$ m)



**Figure S9.** Tumor cell cytoplasmic area in 2D culture with varying HA concentration. (Scale bar: 200  $\mu\text{m}$ )

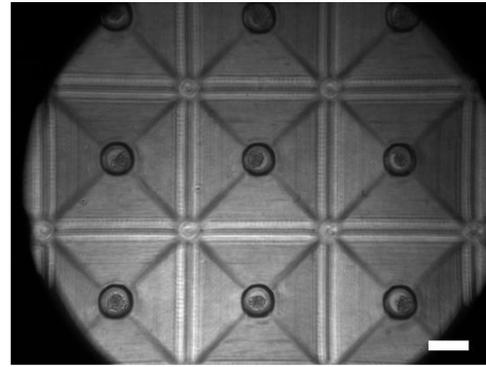
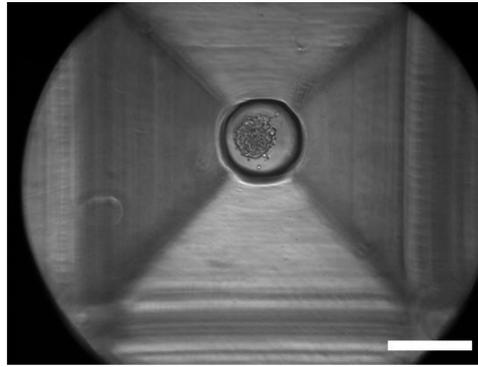


**Figure S10.** Gel construct observation under experiment conditions. HA-fibrin composite with varying HA concentration (0.0%, 0.2%, and 0.4%) was filled into middle channel and no cells were injected to other side channels. Scale bar: 200  $\mu\text{m}$

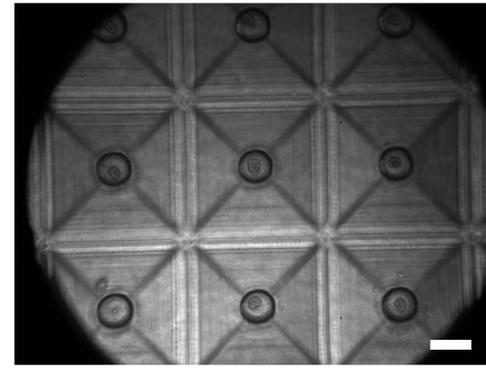
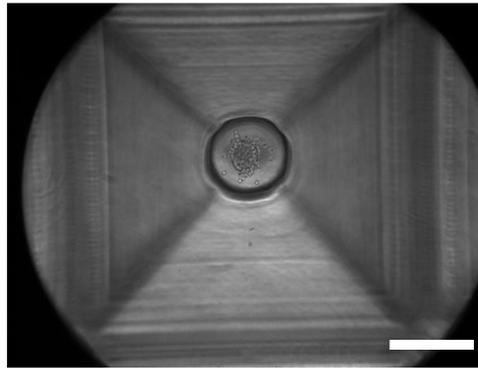


**Figure S11.** ELISA quantification of growth factors/cytokines. (A) TNF- $\alpha$  (B) TGF- $\beta$  (C) VEGF-A for each concentration of HA (0.0, 0.2 and 0.4 %) in HA/fibrin composite.

MKN74

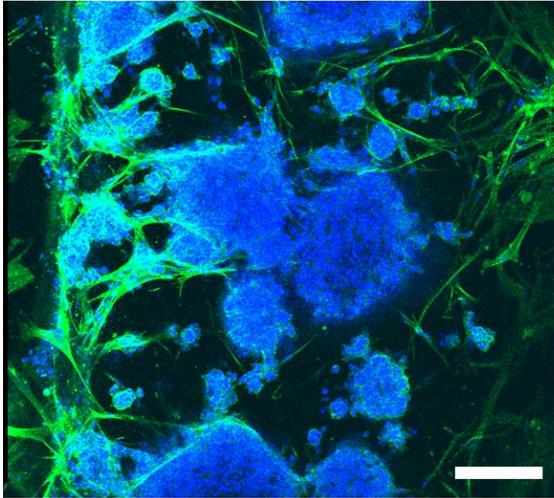


SW620

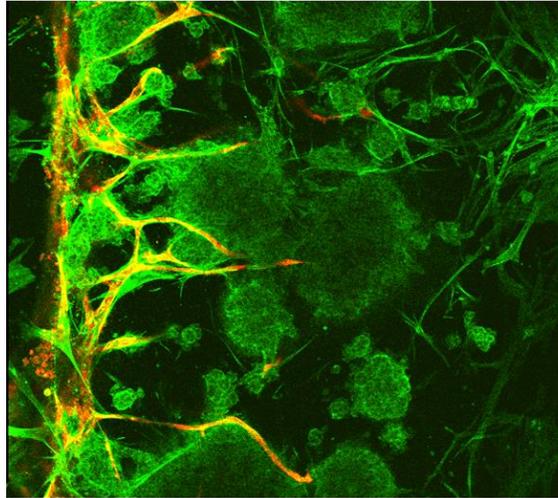


**Figure S12.** Tumor-fibroblast microspheroid formation using SpheroFilm(incyto) at day 2. Scale bar: 300  $\mu\text{m}$

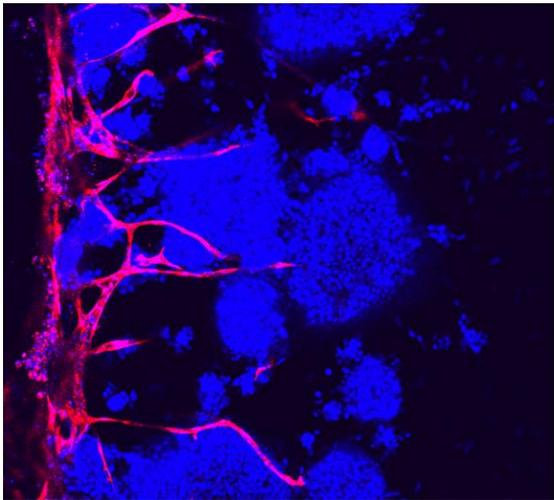
F-actin Hoechst 33342



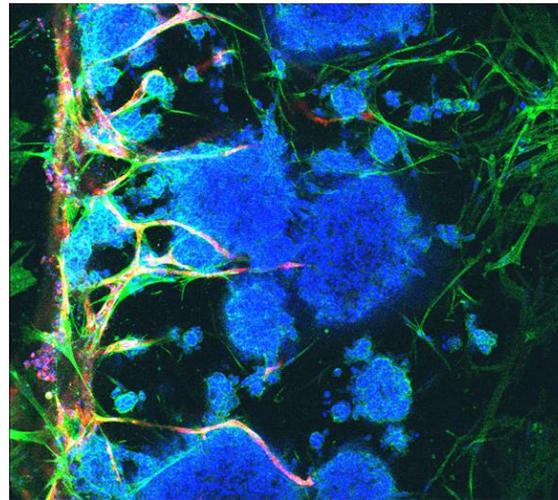
F-actin RFP expressing HUVEC



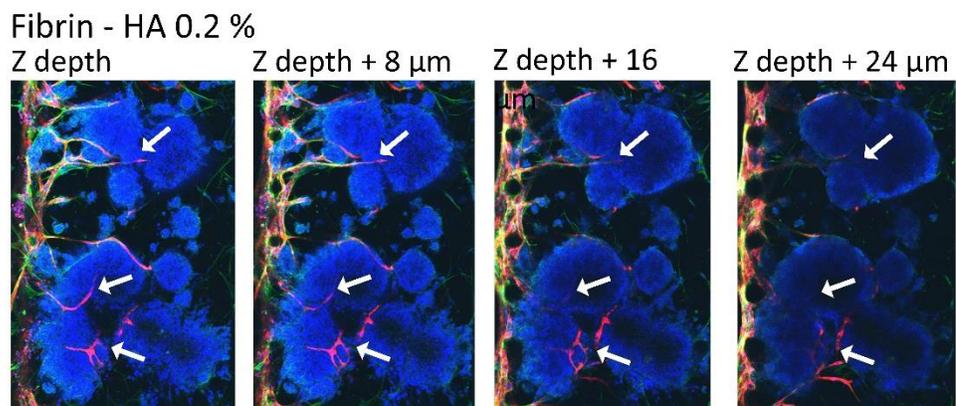
Hoechst 33342 RFP expressing HUVEC



Merge



**Figure S13.** Representative confocal images of direct interaction between tumor-fibroblast microspheroid and blood vessel. Scale bar: 100  $\mu$ m.



**Figure S14.** Representative confocal z-section images of direct interaction between blood vessel and tumor microspheroid (white arrows indicate direct interaction between endothelial cells and the microspheroids). (Scale bar: 200  $\mu\text{m}$ )