Supplementary Material

The Novel Atypical Dopamine Uptake Inhibitor (S)-CE-123 Partially Reverses the Effort-related Effects of the Dopamine Depleting Agent Tetrabenazine

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1. Analytical characterization of S-CE-123

¹H and ¹³C NMR spectra were recorded on a Bruker Avance 500 NMR spectrometer (UltraShield) using a 5-mm switchable probe (PA BBO 500SB BBF-H-D-05-Z, 1H, BB = ¹⁹F and ³¹P - ¹⁵N) with z axis gradients and automatic tuning and matching accessory (Bruker BioSpin). The resonance frequency for ¹H NMR was 500.13 MHz and for ¹³C NMR 125.75 MHz. All measurements were performed for a solution in fully deuterated chloroform or DMSO at 298 K. Standard 1D and gradient-enhanced (ge) 2D experiments, like double quantum filtered (DQF) COSY, NOESY, HSQC, and HMBC, were used as supplied by the manufacturer. Chemical shifts are referenced internally to the residual, non-deuterated solvent signal for acetone ¹H (δ 2.09 ppm) and to the carbon signal of the solvent for acetone ¹³C (δ 30.60 and 205.87 ppm).

HRESIMS spectra were obtained on a maXis HD ESI-Qq-TOF mass spectrometer (Bruker Daltonics, Bremen, Germany). Samples were dissolved to $20 \,\mu$ g/mL in MeOH and directly infused into the ESI source at a flow rate of 3 μ L/min with a syringe pump. The ESI ion source was operated as follows: capillary voltage: 0.9 to 4.0 kV (individually optimized), nebulizer: 0.4 bar (N₂), dry gas flow: 4 L/min (N₂), and dry temperature: 200 °C. Mass spectra were recorded in the range of m/z 50 – 1550 in the positive-ion mode. The sum formulas were determined using Bruker Compass DataAnalysis 4.2 based on the mass accuracy ($\Delta m/z \leq 2$ ppm) and isotopic pattern matching (SmartFormula algorithm).

The purity of the compound was determined by HPLC on an UltiMate 3000 series system equipped with VWD detector (Dionex/Thermo Fisher Scientific, Germering, Germany). Separation was carried out on an Acclaim 120 C18, 2.1 x 150 mm, 3 µm HPLC column (Thermo Fisher Scientific) using LC-MS-grade water and acetonitrile as mobile phase A and B, respectively. The sample components were separated and eluted with a linear gradient from 10% to 90% B in 25 min followed by an isocratic column cleaning and re-equilibration step. The flow rate was 0.2 mL/min and the column oven temperature was set to 25°C. The purity was determined by HPLC chromatography with UV detector (254 nm), as being the ratio of the peak area of the compound and the total peak areas (i.e., the sum of the areas of all peaks that were not present in the solvent blank). For chiral measurements LC-2010A HT Liquid Chromatograph device

(Shimadzu Corporation, Tokyo, Japan) was used equipped with analytical Chiralpack IA (Daicel Inc., Tokyo, Japan). 100% ACN was used as mobile phase, the flow rate was 1 mL/min and the column oven temperature was set to 25°C. Melting points were measured on Leica Galen III apparatus (Leica Biosystems, Germany).

The final product, (*S*)-CE-123, has been unambiguously characterized. The overall purity of the (*S*)-enantiomer, determined by a C18 analytical column-based HPLC method using reversed-phase chromatography conditions was 99.3% The experimental molecular mass of the (*S*)-CE-123 was determined by HRESIMS from the precursor ion m/z 314.0667 [M+H]⁺(calculated for C₁₇H₁₆NOS₂⁺, 314.0668, $\Delta = 0.0$ ppm). Enantiopurity of the less retained (*S*)-enantiomer was determined to be 94.9% (*ee* 89.8%). Melting point was determined to be 106-107°C. Chemical shifts assignment by proton and carbon NMR are shown in **Supplementary Table 4**.

2. Attribution of absolute configuration

2.1. Vibrational-circular dichroism (VCD) analysis

Enantiomers of the racemic compound CE-123 were originally separated as described by *Nikiforuk et al. (2017)*. In short, this was achieved by using CHIRALPACK IA semipreparative column (10mm diameter x 20 mm length) (Daicel Inc, Tokyo, Japan) and 100% EtOH as mobile phase.

Determination of absolute configuration of individual enantiomers obtained by chiral separations was performed using the vibrational-circular dichroism (VCD) method. For this purpose, enantiomers were outsourced to European Center for Chirality, University of Antwerp as a part of BioTools Ltd. (Florida, USA).

2.2. X-ray analysis

To confirm the results from the VCD method, the less retained enantiomer of CE-123 was crystallized and X-ray measurement was performed. The X-ray intensity data were measured on Bruker D8 Venture diffractometer equipped with multilayer monochromator, Mo K/a INCOATEC micro focus sealed tube and Oxford cooling system. The structures were solved by *direct methods* and refined by *full-matrix least-squares techniques*. Non-hydrogen atoms were refined with *anisotropic displacement parameters*. Hydrogen atoms were inserted at calculated positions and refined with riding model. The following software was used: *Bruker SAINT software packageⁱ* using a narrow-frame algorithm for frame integration, *SADABSⁱⁱ* for absorption correction, *OLEX2ⁱⁱⁱ* for structure solution, refinement, molecular diagrams and graphical user-interface, *Shelxle^{iv}* for refinement and graphical user-interface *SHELXS-2013^{vi}* for structure solution, *SHELXL-2013^{vi}* for refinement, and *Platon^{vii}* for symmetry check. Experimental data, CCDC-Codes, Crystal data, data collection parameters, structure refinement details are given in Supplementary Material section.

Based on the agreement of VCD spectra with Confidence Level of 99%, absolute configurations (S) and (R) were assigned to the less and the more retained enantiomer of CE-123, respectively. Experimental details and report are available in the text below. By comparing VCD spectra of the enantiomer of CE-123 synthesized for this study with the VCD spectra of enantiomers of CE-123 to which the absolute configuration had been previously assigned, absolute configuration of the synthesized enantiomer was determined to be (S). Based on crystallographic data and following the *Cahn-Ingold-Prelog* rule, the configuration of the active substance was determined to be (S), which supports the findings of the VCD method.

ⁱ Bruker SAINT v8.37A Copyright © 2005-2016 Bruker AXS

ⁱⁱ Sheldrick, G. M. (1996). SADABS. University of Göttingen, Germany.

ⁱⁱⁱ Dolomanov, O.V., Bourhis, L.J., Gildea, R.J., Howard, J.A.K. & Puschmann, H., OLEX2, (2009), J. Appl. Cryst. 42, 339-341 ^{iv} C. B. Huebschle, G. M. Sheldrick and B. Dittrich, ShelXle: a Qt graphical user interface for SHELXL, J. Appl. Cryst., 44, (2011) 1281-1284

v Sheldrick, G. M. (1996). SHELXS. University of Göttingen, Germany.

vi Sheldrick, G. M. (1996). SHELXL. University of Göttingen, Germany.

vii Spek, A. L. (2009). Acta Cryst. D65, 148-155.

X-ray Analysis

Sample	Machine	Source	Temp.	Detector Distance	Time/ Frame	#Frames	Frame width	CCDC
	Bruker		[K]	[mm]	[s]		[°]	
PrKaSCE123A2	D8	Mo	100	34	30	2014	0.5	1837036

Supplementary Table 1. Experimental parameters and CCDC-Code.

(S)-5-((benzhydrylsulfinyl)methyl)thiazole for "Frontiers in Pharmacology"



Supplementary Figure 1. Asymmetric Unit of [**PrKaSCE123A2**], drawn with 50% displacement ellipsoid. The chiral space group $P2_1$ and the according results of Flack and Hooft parameter (-0.003(14), -0.002(12)) proof the chirality in S1 as "S". The bond precision for C-C single bonds is 0.0025Å. Main Residue disorder is 14%.



Supplementary Figure 2. Packing of [**PrKaSCE123A2**] in plane a-b. Characterized by alternating regions of polar (orange shaded) and nonpolar (blue shaded) fragments.

Supplementary Table 2	. Sample and crystal	l data of [PrKaSCE123A2].
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Chemical formula	C17H15NOS2	Crystal system		monoclinic
Formula weight [g/mol]	313.42	Space group	P21	
Temperature [K]	100	Z		2
Measurement method	f and w scans	Volume [Å ³]		755.27(8)
Radiation (Wavelength [Å])	MoKa ($\lambda = 0.71073$)	Unit cell dimensions [Å] and [°]	9.4062(6)	90
Crystal size / [mm ³]	$0.164 \times 0.079 \times 0.026$		8.4553(5)	116.145(2)
Crystal habit	clear colourless needle		10.5788(6)	90
Density (calculated) / [g/cm ³]	1.378	Absorption coefficient / [mm ⁻¹]	0.35	
Abs. correction Tmin	0.7134	Abs. correction Tmax	0.746	
Abs. correction type	multiscan	F(000) [e ⁻]	328	

Index ranges	$\begin{array}{c} \text{-13} \leq h \leq 13, \text{-11} \leq k \leq \\ 11, \text{-14} \leq l \leq 14 \end{array}$	Theta range for data collection [°]	4.824 to 60.176	
Reflections number	33075	Data / restraints / parameters	4416/4/217	
Refinement method	Least squares	Einal D indiana	all data	R1 = 0.0252, wR2 = 0.0593
Function minimized	$\Sigma \mathrm{w}(\mathrm{F_o}^2 - \mathrm{F_c}^2)^2$	Final K indices	I>2σ(I)	R1 = 0.0235, wR2 = 0.0585
Goodness-of-fit on F ²	1.041		$w=1/[\sigma^{2}(F$	o ²)+(0.0309P) ² +0.1482P]
Largest diff. peak and hole [e Å ⁻³]	0.21/-0.18	Weighting scheme	where $P = (F_0^2 + 2F_c^2)/3$	

Supplementary Table 3. Data collection and structure refinement of [PrKaSCE123A2].





99.27 % (with blank correction)

Supplementary Figure 3. HPLC-method determined purity of S-CE-123



Supplementary Figure 4. High-resolution mass spectrometry spectrum of S-CE-123



Supplementary Figure 5. HPLC-based method for chiral-resolution determines enantiopurity of S-CE-123 to be 94.9 %.



Supplementary Table 4. Chemical shifts and their attribution in ¹H and ¹³C NMR.

VD253 in d ₆ Aceton		$^{1}\mathrm{H}$	¹³ C
Ar 2	СН	8,98	156,16
Ar 4	СН	7,72	145,61
Ar 5	С		128,46
Ph 1	С		138,07
Ph 1	С		136,90
Ph 2,6	СН	7,55	131,23
Ph 2,6	СН	7,57	130,31
Ph 3,5	СН	7,47	130,74
Ph 3,5	СН	7,40	130,00
Ph 4	СН	7,40	129,78
Ph 4	СН	7,35	129,58
СН	СН	5,07	71,64
CH ₂	CH ₂	4,30/3,92	49,03

- 1.2499 - 1.2499		udd
- 5.0500 - 2.8430 - 3.9363 - 3.9363 - 4.1974		<u>1.24</u>)=
- 4 [.] 2401 - 4 [.] 2529		- N
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- 4.5477 - 4.5690 - 4.5766 - 5.6256 - 7.3332 - 5.6261 - 7.5332 - 7.5332 - 7.5090		4 <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10.1</u> <u>10</u>
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VD253 in d6Acetone (APT) 28.9.2017















VD253 in d6Acetone (HSQC) 28.9.2017











VD253 in d6Acetone (HMBC) 28.9.2017









VD253 in d6Acetone (HMBC) 28.9.2017













European Centre for Chirality, Antwerp, Belgium

Page **1** of 12

Title:

Absolute Configuration Determination Report

GENERAL INFORMATION	
Customer	University of Vienna Department of Pharmaceutical Chemistry Predrag Kalaba
Sales Order Number	N/A
Sample code (Our ref.)	S17130 / S17131
Sample description (Your ref.)	S17130 = CE-123A
	S17131 = CE-123B
VCD-spectrometer	ChiralIR-2X w/ Dual <i>PEM</i>
Report prepared by	Wouter Herrebout
Report validated by	Wouter Herrebout
Report signed by	NA
Date	May, 20 th , 2017
RESULTS	
Absolute Configuration of S17130: (<i>S</i>) Absolute Configuration of S17131: (<i>R</i>)	Confidence Level: 99 %
MEASUREMENT PARAMETERS	
Concentration	4.1 mg / 100 μL
Solvent	CDCl ₃
Resolution	4 cm ⁻¹
PEM setting	1400 cm ⁻¹
Number of scans/Measurement time	75.000 scans / 24.0 hours
Sample cell	BaF₂
Path length	100 µm
CALCULATION DETAILS	
Force fields used in MolMec conformational analyses	MMFF94S, MMFF, SYBYL
Number of conformations generated	18
Methodology and basis set for DFT calculations	SCRF-B3LYP/6-31G(d) SCRF-B3PW91/6-31G(d)
Enantiomer used for calculation	(<i>S</i>)
Number of conformations used in calculated spectrum	11
Number of low-energy conformations shown in report	2
COMMENTS	
VCD spectra were recorded using CDCl ₃ as a solvent. Ba spectra of both enantiomers.	aseline corrections were introduced by using

Calculations were performed using B3LYP and B3PW91 functionals, to check for consistency.





Absolute Configuration Determination Report

Molecular Formula: C₁₇H₁₅NOS₂

Chemical Structure:







Absolute Configuration Determination Report

Experimental IR and VCD spectra for S17130



IF-750-QSA-001





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Absolute Configuration Determination Report

Optimized geometries and relative populations of the two calculated lowest-energy conformers for the (S) enantiomer obtained at the SCRF-B3LYP/6-31G(d) level:



9 conformations have a Boltzmann population larger than 1%; 12 conformations have a Boltzmann population larger than 0.1%.





Absolute Configuration Determination Report

Calculated IR and VCD spectra for the (S) enantiomer: B3LYP/6-31G(d)



IF-750-QSA-001

BioTools, Inc. 17546 Bee Line Hwy | Jupiter, Florida 33458, USA ph. 561 625.0133 | fax. 561 625.0717 | www.btools.com





European Centre for Chirality, Antwerp, Belgium

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Title:

Absolute Configuration Determination Report

Optimized geometries and relative populations of the two calculated lowest-energy conformers for the (S) enantiomer obtained at the SCRF-B3PW91/6-31G(d) level:



9 conformations have a Boltzmann population larger than 1%; 11 conformations have a Boltzmann population larger than 0.1%.

IF-750-QSA-001





Absolute Configuration Determination Report

Calculated IR and VCD spectra for the (R,R) enantiomer: B3PW91/6-31G(d)







Absolute Configuration Determination Report

Inspection of the calculated and experimental spectra shows that the B3LYP IR and VCD spectra for the (*S*) enantiomer reproduce the experimental IR and VCD spectra of **S17130**.



The assignment of the AC of S17130 to (*S*) is confirmed by the neighborhood similarities and confidence levels calculated.





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Absolute Configuration Determination Report

Table 1. Numerical comparison describing the similarity in the range of 950-1700 cm⁻¹ between the calculated IR and VCD spectra and the experimental IR and VCD spectra of **S17130**.

S17130 / CE-123A			(S)		
Calculated level	σ ^a	ΣıR ^b	Σ _{VCD} ^c (%)	Δ^{d} (%)	CL ^e (%)
SCRF-B3LYP	0.967	85.7	66.8	51.4	99

^a σ : scaling factor. ^b \sum_{IR} : IR similarity, gives the similarity between the calculated and experimental IR spectra. ^c \sum_{VCD} : single VCD similarity, gives the similarity between the calculated and experimental VCD spectra. ^d Δ : enantiomeric similarity index, gives the difference between the values of \sum for both enantiomers of a given diastereoisomer. ^eCL: confidence level.









Absolute Configuration Determination Report

Inspection of the calculated and experimental spectra shows that the B3PW91 IR and VCD spectra for the (S) enantiomer reproduce the experimental IR and VCD spectra of **S17130**.



The assignment of the AC of S17130 to (S) is confirmed by the neighborhood similarities and confidence levels calculated.

IF-750-QSA-001





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Absolute Configuration Determination Report

Table 2. Numerical comparison describing the similarity in the range of 950-1700 cm⁻¹ between the calculated IR and VCD spectra and the experimental IR and VCD spectra of **S17130**.

S17130 / CE-123 A			(S)		
Calculated level	σ ^a	∑ıR ^b	Σ _{VCD} ^c (%)	Δ^{d} (%)	CL ^e (%)
SCRF-B3LYP	0.965	85.1	61.1	44.0	99

^a σ : scaling factor. ^b \sum_{IR} : IR similarity, gives the similarity between the calculated and experimental IR spectra. ^c \sum_{VCD} : single VCD similarity, gives the similarity between the calculated and experimental VCD spectra. ^d Δ : enantiomeric similarity index, gives the difference between the values of Σ for both enantiomers of a given diastereoisomer. ^eCL: confidence level.



Statistics: S17130 in database (B3PW91)





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Absolute Configuration Determination Report

CONCLUSIONS

Absolute configuration of **S17130** (CE-123 A) was assigned as (S) based on the agreement of VCD spectra

Confidence Level: 99 %

Absolute configuration of **S17131** (CE-123 B) was assigned as (*R*) based on the agreement of VCD spectra. Confidence Level: **99** %

GENERAL INFORMATION			
Customer	University of Vienna Department of Pharmaceutical Chemistry Predrag Kalaba		
Sales Order Number	N/A		
Sample code (Our ref.)	S17310 / S17311		
Sample description (Your ref.)	S17310 = VD-253 S17311 = VD-257		
VCD-spectrometer	ChiralIR-2X w/ Dual <i>PEM</i>		
Report prepared by	Wouter Herrebout		
Report validated by	Wouter Herrebout		
Report signed by	NA		
Date	December, 14 th , 2017		
RESULTS			
Absolute Configuration of S17310: (<i>S</i>) Absolute Configuration of S17311 (R)	No Confidence Level		
MEASUREMENT PARAMETERS			
Concentration	2.5 mg / 100 μL , 4.4 mg / 100 μL		
Solvent	CDCl₃		
Resolution	4 cm ⁻¹		
PEM setting	1400 cm ⁻¹		
Number of scans/Measurement time	75.000 scans / 24.0 hours		
Sample cell	BaF ₂		
Path length	100 µm		
CALCULATION DETAILS			
Force fields used in MolMec conformational analyses	NA , calculations performed for S17130		
Number of conformations generated	NA , calculations performed for S17130		
Methodology and basis set for DFT calculations	NA , calculations performed for S17130		
Enantiomer used for calculation	NA _ calculations performed for S17130		
Number of conformations used in calculated spectrum	NA , calculations performed for S17130		
Number of low-energy conformations shown in report	NA, calculations performed for S17130		
	, I		
S17130 = CE-123A was confidently assigned to S (earlier report , 99% CL) S17131 = CE-123B was confidently assigned to R (earlier report , 99% CL)			

IR and VCD spectra of S17310 / VD-253 and S17311 / VD-257 were recorded. Assignment of the AC was performed by comparing the experimental IR and VCD spectra of S17130 / S17131 and S17310 / S17311.

Due to additional spectral features in the IR and VCD spectra, assigned to unidentified impurities, no confidence levels could be obtained for S17310 / S17311. The characteristic pattern observed in VCD allows the AC of S17310 and S17311 to be confidently assigned.

Molecular Formula: C₁₇H₁₅NOS₂

Chemical Structure:





Experimental IR and VCD spectra for S17130 – taken from earlier report



Experimental IR and VCD spectra for S17310 - new data



Experimental IR and VCD spectra for S17310 (black) and S17130 (red) - impurities in S17310/311



