

## APPENDIX

1 The equations describing the  $HVC_X$  neuron dynamics are taken from the work of Daou [1]. That paper  
 2 also has an extensive account of his experiments on the other two major classes of neurons in HVC. The  
 3 equations are of Hodgkin-Huxley (HH) form for a neuron without spatial extent; this is called a one-  
 4 compartment model. It is meant to apply to neurons in isolation of the network, here HVC, in which they sit  
 5 *in vivo*. The dynamical variables include the observable quantities: voltage across the cell membrane,  $V(t)$   
 6 and the intracellular concentration of  $[Ca^{2+}]_{in}(t) = C(t)$ .  $V(t)$  is directly connected to action potentials  
 7 or voltage spikes that communicate among cells in a network; the time scale of these spikes is a few ms.  
 8  $C(t)$  provides a slow background modulation that raises the cells potential (depolarizes the cell) or lowers  
 9 it (hyperpolarizes the cell) on time scales as long as 10's of ms.

10 The voltage equation, conservation of charge, relates the capacitance of the cell membrane  $C_m$  as it  
 11 separates concentrations of ions within and without the cells to the various currents which contain the  
 12 nonlinear voltage dependence of the permeability of ions to passing into and out of the cell. The model  
 13 represents these ion currents:  $\{Na, K, Ca\}$  in several different ways.

14 The general form of an HH current is

$$I_{ion}(t) = g_{ion} m_{ion}^{integer1}(t) h_{ion}^{integer2}(t) (E_{rev-ion} - V(t)), \quad (1)$$

15 where the reversal potential is the equilibrium Nernst potential [2, 3]. The gating variables  $\{m(t), h(t)\}$  lie  
 16 between zero and one and represent the amount the ion channel is open relative to the maximum opening it  
 17 may have. The maximal conductance  $g_{ion}$  represent the number or density of ion channels in the neuron  
 18 model. This form of ion current applies when the concentrations of the ion are not significantly different  
 19 outside and inside the cell. This is not so for  $Ca^{2+}$  ions, so there we use the Goldman-Hodgkin-Katz  
 20 (GHK) form of the current [4].

$$I_a(t) = -P_a z_a^2 F^2 \left( \frac{[ion]_{in}(t) - [ion]_{out} e^{-z_a F V(t)/RT}}{1 - e^{-z_a F V(t)/RT}} \right), \quad (2)$$

21 for ion  $a$ .  $z_a$  is the charge on the ion,  $F$  the Faraday constant,  $R$  the gas constant,  $T$  the temperature, and  $P_a$   
 22 the permeability of the cell membrane to ion  $a$ .

23 We use an approximation to the GHK equations for the two types of Calcium currents selected in [1]

$$\begin{aligned} I_{CaL}(t) &= g_{CaL} V s_{\infty}^2(V(t)) \left( \frac{[Ca]_{out}}{e^{2FV(t)/RT} - 1} \right) \\ I_{CaT}(t) &= g_{CaT} V(t) [a_T]_{\infty}^3(V) [b_T]_{\infty}^3(r_T^A) \left( \frac{[Ca]_{out}}{e^{2FV(t)/RT} - 1} \right) \\ b_{T\infty}(r_T) &= \frac{1}{1 + e^{\left(\frac{r_T - \theta_b}{\sigma_b}\right)}} - \frac{1}{1 + e^{\left(\frac{-\theta_b}{\sigma_b}\right)}} \end{aligned} \quad (3)$$

24  $a_T$  and  $b_T$  are instantaneous activating and inactivating gating variables, respectively.  $r_T$  is a slow gating  
 25 variable which takes the same functional form as  $a_T$  and other gating variables  $m(t)$  and  $h(t)$ . These gating

26 variables  $w(t)$  satisfy a first order kinetic equation

$$\frac{dw(t)}{dt} = \frac{w_{\infty}(V(t)) - w(t)}{\tau_w(V(t))}, \quad (4)$$

27 in which

$$w_{\infty}(V) = \frac{1}{2} \left[ 1 - \tanh \left( \frac{V - \theta_w}{2\sigma_w} \right) \right], \quad (5)$$

28 for all gating variables except  $h_{\infty}(V)$  appearing in  $I_{Na}(t)$  [1].  $\theta_w$  is the half-activation voltage and  $\sigma_w$   
 29 controls the slope of the activation function. For fast gating variables, such as  $m$  of  $I_{Na}$ , and  $s$  of  $I_{CaL}$  we  
 30 replace the time dependence by  $W_{\infty}(V)$ .

31  $\tau_w(V)$  is the time constant of each gating variable. Time constants for the  $n$  and  $hp$  gating variables  
 32 (these names refer to [1]) are given below, where  $\bar{\tau}_w$  is an average time constant. Our model differs from [1]  
 33 by one time constant. Instead,  $\tau_{rs}(V)$  takes the form presented here:

$$\tau_w(V) = \frac{\bar{\tau}_w}{\cosh \left( \frac{V - \theta_w}{2\sigma_w} \right)}$$

for  $n$  or  $hp$

$$\tau_{rs}(V) = 0.1 + 193.0 \left( 1 - \tanh^2 \left( \frac{V(t) + 80}{-21} \right) \right)$$

$$\tau_{rf} = \frac{p_{rf}}{\frac{-7.4(V+70)}{e^{\frac{V+70}{-0.8}} - 1} + 65e^{\frac{V+56}{-23}}} \quad \tau_{rT}(V) = \tau_{r0} + \frac{\tau_{r1}}{1 + e^{\left( \frac{V - \theta_{rT}}{\sigma_{rT}} \right)}}$$

34 For our choice of ion currents we follow the results of experimental data [5, 6, 7] and generally reproduce  
 35 the model listed in [1].  $HVC_X$  spiking properties include fast rectifying current, sag in response to  
 36 hyperpolarizing current, and spike frequency adaption in response to depolarizing current.

$$C \frac{dV(t)}{dt} = I_{Na}(t) + I_K(t) + I_L(t) + I_{CaT}(t) + I_{CaL}(t) + I_A(t) + I_{SK}(t) + I_h(t) + I_{Nap}(t) + I_{injected}(t) \quad (6)$$

37  $I_{Na}(t)$  and  $I_K(t)$  are the standard HH currents. They produce fast spiking in response to injected currents.  
 38  $I_L(t)$  is a leak current meant to capture all linear currents of the neuron.  $I_{CaT}(t)$  is a low threshold T-type  
 39 calcium current that causes rebound depolarization in cooperation with  $I_h(t)$ .  $I_{CaL}(t)$  is a high threshold  
 40 L-type calcium current.  $I_{CaL}(t)$  works in conjunction with  $I_{SK}(t)$ , a calcium concentration dependent  
 41 potassium current, to create frequency adaptation in neuron spiking.  $I_A(t)$  is an A-type potassium current.  
 42  $I_{Nap}(t)$  is a persistent sodium current. From the model presented in [1], we eliminate  $I_{KNa}(t)$ , a sodium  
 43 dependent potassium current, and rewrite all sigmoidal functions as hyperbolic tangents.

44 The mass conservation equation for  $Ca^{2+}$  is written as

$$\frac{dC(t)}{dt} = \epsilon(I_{CaT}(t) + I_{CaL}(t)) + k_{Ca}(b_{Ca} - C(t)), \quad (7)$$

45 again following [1].

## REFERENCES

- 46 [1] Daou A, Ross M, Johnson F, Hyson R, Bertram R. Electrophysiological characterization and  
47 computational models of hvc neurons in the zebra finch. *Journal of Neurophysiology* **110** (2013)  
48 1227–1245.
- 49 [2] Johnston D, Wu SMS. *Foundations of Cellular Neurophysiology* (Bradford Books, MIT Press) (1995).
- 50 [3] Sterratt D, Graham B, Gillies A, Willshaw D. *Principles of Computational Modelling in Neuroscience*  
51 (Cambridge University Press) (2011), 390 .
- 52 [4] Goldman DE. Potential, impedance, and rectification in membranes. *The Journal of General Physiology*  
53 **27** (1943) 37–60. doi:10.1085/jgp.27.1.37.
- 54 [5] Dutar P, Vu HM, Perkel DJ. Multiple cell types distinguished by physiological, pharmacological, and  
55 anatomic properties in nucleus hvc of the adult zebra finch. *Journal of Neurophysiology* **80** (1998)  
56 1828–1838. doi:10.1152/jn.1998.80.4.1828. PMID: 9772242.
- 57 [6] Mooney R. Different subthreshold mechanisms underlie song selectivity in identified hvc neurons of the  
58 zebra finch. *Journal of Neuroscience* **20** (2000) 5420–5436. doi:10.1523/JNEUROSCI.20-14-05420.  
59 2000.
- 60 [7] Kubota M, Taniguchi I. Electrophysiological characteristics of classes of neuron in the hvc of the zebra  
61 finch. *Journal of Neurophysiology* **80** (1998) 914–923. doi:10.1152/jn.1998.80.2.914. PMID: 9705478.