

Supplementary Material

CHOLESTEROL PROTECTS AGAINST ACUTE STRESS-INDUCED T-TUBULE REMODELING IN MOUSE VENTRICULAR MYOCYTES

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Supplementary Figure 1. Cardiomyocytes were incubated at different concentrations of M β CD at RT for 1 hour followed by membrane labeling with di-8-ANEPPS and confocal imaging. A clear deterioration of TATS was observed a 9 mM M β CD.



Supplementary Figure 2. Quantification of cellular cholesterol using filipin staining. *A*. Representative widefield images of filipin-stained cardiomyocytes. Cells were treated with M β CD at indicated concentrations for 1 hour at 37 °C. Note there is clear perinuclear fluorescence suggesting that filipin penetrates the membrane and stains intracellular cholesterol, ultimately leading to overestimation of the sarcolemmal component of cholesterol.

B. Quantification of the data. Right panel includes the data from cells incubated at room temperature (RT; ~20-22 °C). See Methods section in the main text for details. The decrease in the magnitude of filipin fluorescence seems to saturate at high concentrations of M β CD and/or temperature again suggesting significant staining of the intracellular cholesterol (not affected by membrane impermeable M β CD).



Supplementary Figure 3. Concentration and temperature effects of M β CD on dextran trapping. Cardiomyocytes were incubated for 1 hour at either room temperature (RT; light gray bars) or 37°C (dark gray bars) in C solution containing the indicated concentration of M β CD. Cardiomyocytes treated with 1 mM M β CD at 37°C displayed significant mortality after detubulation with standard hyposmotic Tyrode based solutions (e.g. 0.6 Na). To circumvent this issue, a modified hyposmotic stress protocol was developed using C solution (290 mOsm) as the base solution. Hyposmotic C solutions were prepared containing 60% (0.6 Na_C, 206 mOsm) and 70% (0.7 Na_C, 228 mOsm) of the base [NaCl]. After the 1-hour incubation, cells were detubulated using 0.7 Na_C solution. Control detubulation using 0.6 Na_C solution was performed with cells incubated at RT. All data were normalized to that obtained using 0.6 Na_C solution (black bar). N = 18-20 cells per group.



Supplementary Figure 4. Correlation between filipin staining (cholesterol) intensity and magnitude of dextran trapping. The dextran trapping data from Supplementary Figure 3 are plotted against the corresponding filipin fluorescence obtained from Figure 1H and Supplemental Figure 2B.