

## *Supplementary Material*

# **Harnessing Metabolic Regulation to increase Hfq-dependent Antibiotic Susceptibility in *Pseudomonas aeruginosa***

**Petra Pusic<sup>+</sup>, Elisabeth Sonnleitner<sup>\*+</sup>, Beatrice Krennmayr, Dorothea A. Heitzinger, Michael T. Wolfinger, Armin Resch and Udo Bläsi<sup>\*</sup>**

<sup>+</sup> These authors contributed equally to this work

**\* Correspondence:**

Elisabeth Sonnleitner: [elisabeth.sonnleitner@univie.ac.at](mailto:elisabeth.sonnleitner@univie.ac.at)

Udo Bläsi: [udo.blaesi@univie.ac](mailto:udo.blaesi@univie.ac)

## 1 Supplementary Figures and Tables

### 1.1 Supplementary Tables

**Supplementary Table S1.** Susceptibility of PAO1 and PAO1 $\Delta hfq$  towards different classes of antibiotics in LB medium.

Classes/Subclass	Antibiotic	Mode of action	MIC	PAO1	PAO1 $\Delta hfq$	PAO1	PAO1 $\Delta hfq$
SCFM				High inoculum <sup>a</sup>		Low inoculum <sup>b</sup>	
<b><math>\beta</math>-Lactam antibiotics</b>							
Cephems/Cephalosporins IV	Cefepime	cell wall	mg/L	1.5/1.5	1.5/1.5	0.5/0.38	0.5/0.38
Penems/Carbapenems	Imipenem	cell wall	mg/L	2/2	1/1	1/1	0.5/0.25
<b>Non-<math>\beta</math>-Lactam antibiotics</b>							
Aminoglycosides	Gentamicin	protein synthesis	mg/L	4/4	0.25/0.5	2/2	0.5/0.25
Fluoroquinolones	Ciprofloxacin	DNA metabolism	mg/L	0.12/0.12	0.03/0.06	0.06/0.06	0.008/0.008
Fosfomycins	Fosfomycin	cell wall	mg/L	>1024/>1024	24/16	46/46	32/32
Lipopeptides/Polymixins	Colistin	cell membrane	mg/L	1/1.5	0.38/0.5	0.75/0.75	0.094/0.064
Tetracyclines	Tetracycline	protein synthesis	mg/L	16/16	8/8	8/16	4/4

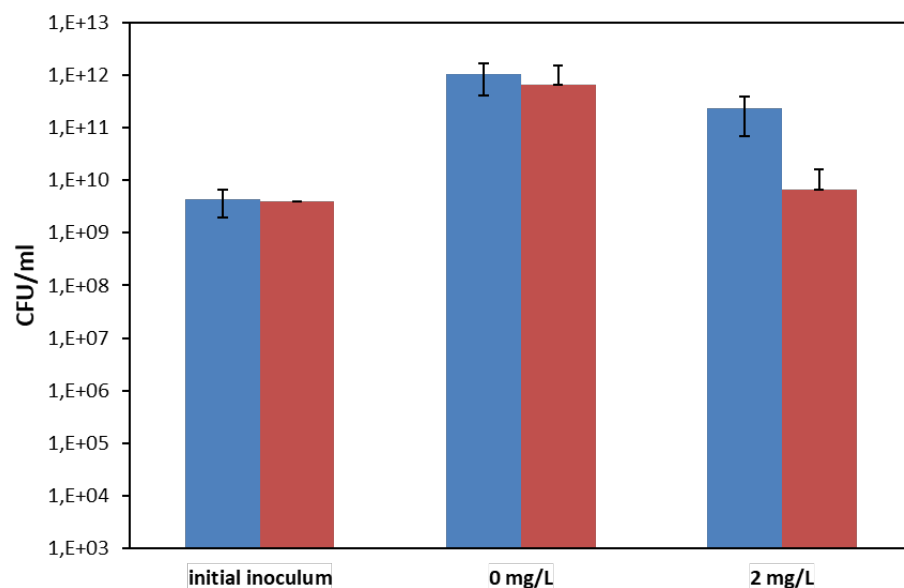
/, results of two independent experiments; >, more resistant than the highest concentration tested.

<sup>a, b</sup> See Table 1.

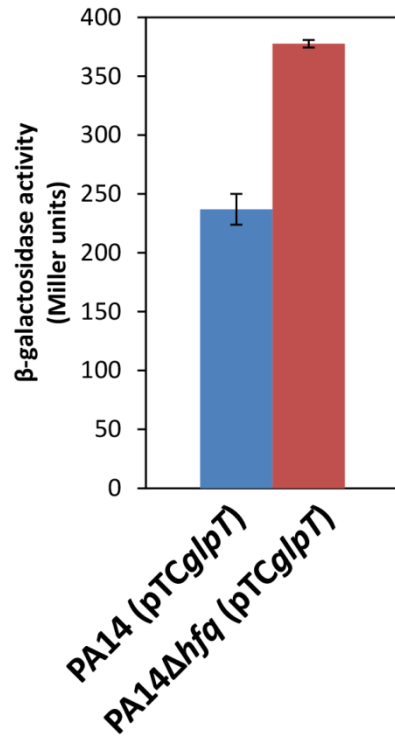
**Supplementary Table S2** Transcripts with increased abundance in PA14 $\Delta$ *hfq* versus PA14.

**Supplementary Table S3** Transcripts with decreased abundance in PA14 $\Delta$ *hfq* versus PA14.

## 1.2 Supplementary Figures

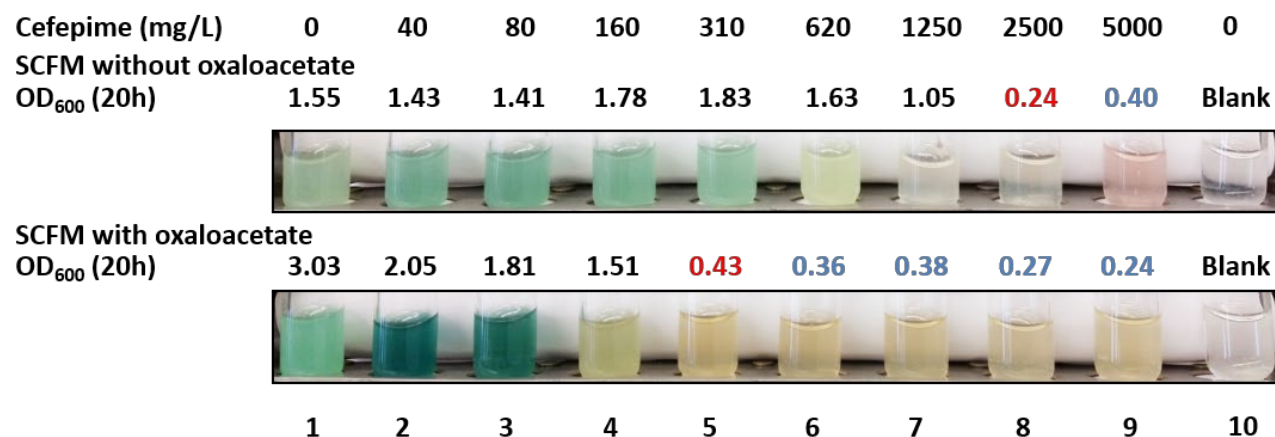


**Supplementary Figure S1** Viability of early stationary phase cells of PA14 and PA14Δ*hfq* after treatment with gentamicin. Determination of the CFUs of PA14 (blue bars) and PA14Δ*hfq* (red bars) at an OD<sub>600</sub> of 2.0 (initial inoculum) and in the absence (0 mg/L) or presence (2 mg/L) of gentamicin after 18 h of static growth in SCFM.



**Supplementary Figure S2** The transcription of *glpT* is increased in the absence of Hfq. The strains PA14 (pTC*glpT*) (blue bar) and PA14Δ*hfq* (pTC*glpT*) (red bar) were grown in SCFM. Samples were withdrawn at an OD<sub>600</sub> of 2.0. The bars represent the β-galactosidase values conferred by the plasmid pTC*glpT* encoded transcriptional *glpR-lacZ* fusion gene in the presence or absence of *hfq*. The error bars represent standard deviations from three independent experiments.

## PAO1



**Supplementary Figure S3** Addition of OAA to SCFM results in increased sensitivity of PAO1 towards cefepime. The PAO1 culture was diluted to an initial OD<sub>600</sub> of 0.5 and incubated for 20 h in SCFM with or without 40 mM OAA in the presence of different cefepime concentrations as indicated on top. Pictures were taken and the OD<sub>600</sub> was measured 20 h after inoculation. The antibiotic concentrations in the presence of which the cells did not grow above OD<sub>600</sub> of 0.5 (marked in red) were considered as MIC. All OD<sub>600</sub> values obtained above this cefepime concentration are depicted in blue indicating toxicity. The experiments were performed in duplicate, revealing the same MICs. Only one representative experiment is shown.