

The Cytochrome *bd* complex is essential for sulfide, sulfate and chromate resistance and is regulated by a GbsR-type regulator, CydE, in *Alishewanella* sp. WH16-1

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TABLE S1 Bacterial strains and plasmids used in this study

Strains or plasmids	Relevant characteristic(s)	Source or reference
Bacterial strains		
<i>Alishewanella</i> sp.		
WH16-1	Wild type; Cr(VI) ^r ; Rif ^r , Km ^s , Tc ^s , Cm ^s	This study
Δ <i>cytbd</i>	Mutant of WH16-1(Rif); Tn5 inserted in <i>cydB</i> ; Rif ^r , Km ^r	This study
Δ <i>cytbd</i> -C	Mutant of WH16-1(Rif) complemented with the <i>Cytbd</i> oxidase operon cloned in pCT-Zori vector; Rif ^r , Km ^r , Cm ^r	This study
<i>Escherichia coli</i>		
<i>E. coli</i> S17-1(λ <i>pir</i>)	<i>Tpr Smr recA thi pro hsdR⁻ hsdM⁺</i> . RP4:2Tc:Mu:Km	(Chen et al., 2015a)
<i>E. coli</i> DH5α	F ⁻ <i>endA1 hsdR17 supE44 thi-1 recA1 deoR gyrA96 relA1</i> Δ (<i>argF-lacZYA</i>)U169 Φ80 <i>dlacZ</i> M15 λ ⁻	Promega
XL1-Blue	Δ(<i>mcrA</i>)183Δ(<i>mcrCB-hsdSMR-mrr</i>)173 <i>endA1 supE44 thi-1 recA1 gyrA96 relA1lac</i> [F' <i>proAB lacIqZ</i> ΔM15 Tn5 Km ^r]	Stratagene
BL21	B F ⁻ <i>dcm ompT hsdS</i> (r _B ⁻ m _B ⁻) <i>gal λ</i> (DE3)	Novagen
Plasmids		
pRL27	Tn5-based transposon vector, <i>oriR6K</i> , Km ^r	(Larsen et al., 2002)
pCT-Zori	Cm ^r	(Chen et al., 2015a)
pCT-Zori- <i>cytbd</i>	pCT-Zori vector cloned with the whole <i>cytbd</i> operon	This study
pLSP	<i>lacZ</i> reporter gene vector	(Li et al., 2015)
pLSP-Promoter	pLSP containing promoter region of <i>cytbd</i> operon	This study
pLSP-Promoter- <i>cydE</i>	pLSP containing promoter and <i>cydE</i> coding region of <i>cytbd</i> operon	This study
pTRG	Tet ^r , for bacterial one-hybrid assay	Stratagene
pTRG- <i>cydE</i>	pTRG containing <i>cydE</i> coding region	This study
pBXcmT	Cm ^r , for bacterial one-hybrid assay	(Guo et al., 2009)
pBXcmT-Promoter	pBXcmT containing <i>cytbd</i> operon promoter region	This study
pET28a	His6 Tag expression vector, Km ^r	Novagen
pET28a- <i>cydE</i>	<i>cydE</i> in frame fusion to the multiple sites of pET-28a	This study

TABLE S2 Primers used in this study

Primer	Sequence(5'-3')	Description
tpnRL17-1	AACAAGCCAGGGATGTAACG	Used for plasmid rescue
tpnRL13-2	CAGCAACACCTTCTTCACGA	
<i>cydE-cydAF</i>	TAGCCGCGGTAGTATCTG	Used to amplify <i>cydE-cydA</i>
<i>cydE-cydAR</i>	GATGGTAATGGTTGGGAA	
<i>cydA-cydBF</i>	GGTGTAGCCTGCGGTTGT	Used to amplify <i>cydA-cydB</i>
<i>cydA-cydBR</i>	TCCCAGAACGGACCAATAG	
<i>cytbdF</i>	AAAGAGCTCGGTAAACCTTGCATTAAGT	Used for complementation of WH16-1- <i>cydB</i> (Tn5)
<i>cytbdR</i>	AAAAAGCTTTCTCATTGTCTTATTAGTAATACTT	
LacPF	AAAGAATTCGGTTAAACCTTGCATTAAGT	Used for construction the <i>lacZ</i> reporter gene vector
LacPR	AAAGGATCCCATTGGAGTCATTGCATA	
LacP- <i>cydER</i>	AAAGGATCCACAGGTGGCAAAAACAG	Used for construction <i>CydE</i> over-expression vector with pET28a
<i>cydEF</i>	AAAGGATCCATGCAAATGACTCCAATG	
<i>cydER</i>	AAAAAGCTTGGCTTGTTTTGCCGGATC	Used for bacterial one-hybrid experiment of <i>cytbd</i> cluster regulatory region DNA
BohpF	AAAGGATCCATGCAAATGACTCCAATG	
BohpR	AAAGAATTCAGAACAACAGGTGGCAAA	Used for construction of <i>CydE</i> protein expression vector with pTRG
BohsF	AAAGGATCCATGCAAATGACTCCAATG	
BohsR	AAAGAATTCAGAACAACAGGTGGCAAA	Used to amplify promoter region sequence of <i>cytbd</i> cluster
PromoterF	GGTTAAACCTTGCATTAAGT	
PrompterR	CATTGGAGTCATTGCATA	Used to amplify FAM labeled promoter region of <i>cytbd</i> operon
FAM-PrompterR	FAM-CATTGGAGTCATTGCATA	

* The underlined sequence denotes the restriction enzyme sites.

A

GGTTAAACCTTGCATTAAGTTAACGGCGTAACCTTAACCTTGATCCGTAATGAATGTTTCGCGGGTGAGTTTCAC
TACTTTAGCAGTTCTATCAAGCCTTAACAGTCTGTGATAGAAAAGCCCAACTTTTTTCTGAATCGTGCGC
TCAACGCCCACTAAATTCTTTAATTAGTTTCGAGCCATACCTGCAAATTATGTTGATCTGGATCAAGCAACA
AGCGTATTATTATTCAGAAATTTCTGAAAGTTCAGTAACGGGATGCGCATATGCAAATGACTCCAATGATTG
AATCCTTTGTTTACCACCTTTGGCGAAATGGGCAGCCGTTGGGGCTTTAACCGTACCCTTGGCCAAATTACG
CCCTGCTGGTGATTAGCGAGCAACCGCTCTGTGCAATGAAATTGGTGAGCTGCTGGGGATTTCGCGCGG
CAATGTCAGTATGGGGTTAAAAGAGTTGGGTAGCTGGCAGCTGGTGCAGGTTAGCCATAAACCCGGCGAG
CGGCGCGAGTTTTATCATAGCCGCGGTAGTATCTGGGATATGGCCAACCGGGTGTGTTGAAGAGCGGCGCA
AACGGGAAATGGACCCGACGCTTAGCTTACTGCGTGACATTATGATGGACGAAGCGGCTAATCCGGAAGA
AGCCTTTGCCAAAAGCGCATGCGCGAAATTCACGATCTGCTGGAGAACATTACTGAATGGACGCAAAGCC
TGCAATCCTTGTCGCCGAGAGCTGAATACCTTAATGAAGTTAGGCTCCGGCGTTGGCAAGGTACTGGA
TTTACCGATAAGTTTTTAGGTAAAAAAGATCCGGCAAACAAGCCTAGCTGTTTTTGCCACCTGT

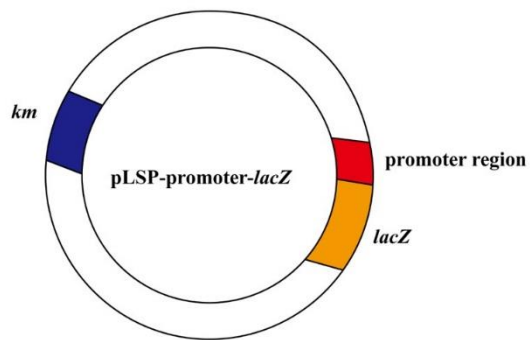
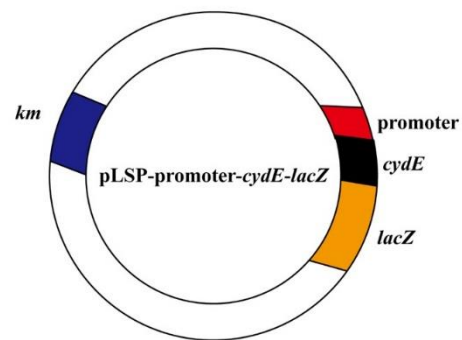
B**C**

FIG S1 Sequence of *cydE* and its promoter and the construction of related vectors. (A) The red nucleotides are the *cydE* promoter region and 18 nucleotides of *cydE*, and the underlined nucleotides are *cydE* coding sequence; (B) The *cydE* promoter sequence (red) was cloned into the pLSP-kt2*lacZ* plasmid to generate the pLSP-promoter-*lacZ* vector; (C) All of the sequence shown in “A” including *cydE* promoter and *cydE* was cloned into the pLSP-kt2*lacZ* plasmid to generate the pLSP-promoter-*cydE-lacZ* vector. For the construction of the pBX-promoter vector in the bacterial one-hybrid assay (FIG 5), the promoter sequence of *cydE* (red) was cloned into the pBXcmT plasmid.

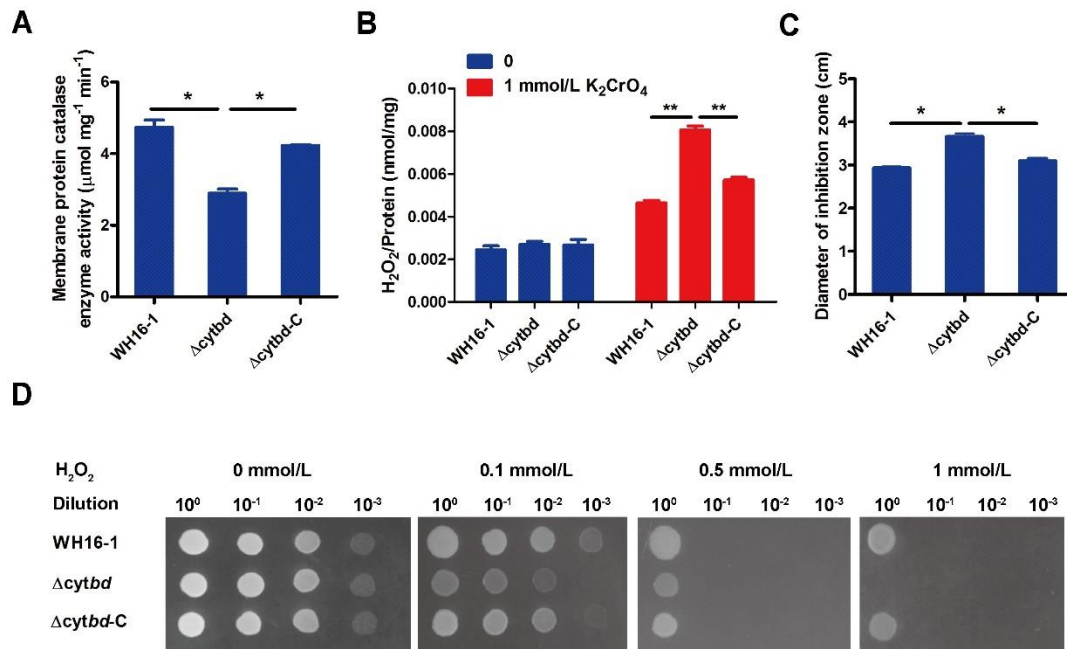


FIG S2 Effects of Cytbd on the reduction of cellular oxidative stress in strain WH16-1, mutant Δcytbd and complementary strain $\Delta\text{cytbd-C}$. (A) The H_2O_2 decomposition ability of membrane proteins. (B) Cellular H_2O_2 contents with 0 and 1 mmol/L K_2CrO_4 , respectively. (C) The inhibition zones when exposed to 3% H_2O_2 . (D) H_2O_2 sensitive tests on LB plates. Every sample was prepared in triplicates and the results are presented as mean values. *, remarkable difference; **, extremely remarkable difference.

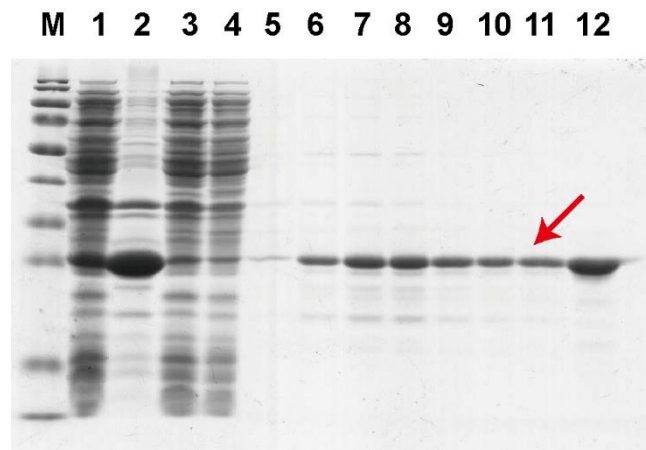


FIG S3 Purification of CydE. Lanes 1 and 2 show cell lysis product suspensions and precipitates after centrifuging, respectively. Lanes 3-5 and 6-12 show proteins eluted from a nickel-ion affinity chromatography column by 200 and 500 mM imidazole, respectively. The red arrow marks proteins that were used in a subsequent EMSA test.